

Effect of dietary supplementation with selenium-enriched yeast or sodium selenite on ruminal enzyme activities and blood chemistry in sheep

Zita Faixová¹, Elena Piešová¹, Zuzana Maková¹, Klaudia Čobanová², Štefan Faix²

¹University of Veterinary Medicine and Pharmacy in Košice, Institute of Pathological Physiology, Košice, Slovak Republic

²Slovak Academy of Sciences, Institute of Animal Physiology, Košice, Slovak Republic

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The experiment was conducted to evaluate the effect of feeding a diet supplemented with different forms of selenium on the rumen fluid, blood and serum enzyme activity and osmotic fragility of red blood cells in sheep. The experiment was carried out on 18 sheep of the Valashka breed at the age of 18 months, divided into 3 groups. The first group was given basal diet (BD) with a Se content of 0.17 mg/kg of dry matter (DM). The second group received BD supplemented with 0.4 mg Se/kg of (DM) in the form of sodium selenite. The third group received BD supplemented with 0.4 mg Se/kg of (DM) in the form of Se-yeast extract. Duration of the trial was 12 weeks. Selenium concentration in blood and total rumen fluid were elevated in both supplemented groups with the highest values in Se-yeast-treated sheep. Blood glutathione peroxidase (GPx) activity was significantly increased, regardless of the source of selenium. Osmotic resistance of red blood cells was not affected by selenium supplementation. The results indicate that feeding a diet supplemented with selenium from Se-yeast or selenite improved selenium status in blood and total rumen fluid. Selenium from sodium selenite was as effective as selenium from Se-yeast in the availability of selenium for the blood GPx activity. The effect of selenium supplementation on the ruminal enzyme activity depends on the selenium form; GGT and GDH were significantly higher in the Se-yeast supplement group, AST and ALP were significantly higher in the selenite supplement group.

Rumen, blood enzymes, ruminant

Selenium is an essential trace element which plays an important role in the antioxidative, reproductive, endocrine and immune systems of species.

In most European Union countries the natural Se content in feed is only 0.03–0.12 mg/kg of dry matter (Pappas et al. 2008). The intake of such feeds can result in serious Se deficiencies and health problems especially in young growing and highly producing animals.

For this reason commercial animal feeds are routinely supplemented with various Se sources up to the maximum European Union authorized total Se content of 0.5 mg/kg of feed.

Currently, sodium selenite (Na_2SeO_3) and sodium selenate (Na_2SeO_4) as inorganic and selenium-enriched yeast (Se-yeast) as organic Se are principal selenium supplements of animal feed.

It is established that selenocompounds from dietary inorganic and organic Se sources follow different metabolic pathways (Qin et al. 2007; Wang et al. 2009). Intestinal absorption of Se is much lower in ruminants than in nonruminants. For selenite, the absorption is 79% and 80% in poultry and swine, respectively, whereas only 29% in sheep. For selenomethionine and selenate the absorption is higher than 90% in monogastrics and poultry. Low absorption of selenium in ruminants is believed to result from reduction of dietary selenium (selenite and selenate) to unsoluble forms such as elemental selenium or selenides in the rumen environment (Mehdi et al. 2013).

Address for correspondence:

Zita Faixová
Institute of Pathological Physiology
University of Veterinary Medicine and Pharmacy in Košice
Komenského 73, 041 81 Košice, Slovak Republic

Phone: +421 915 984 704
E-mail: zita.faixova@uvlf.sk
<http://actavet.vfu.cz/>

There are several methods of evaluation of the selenium status in species. It can be measured in plasma, serum, whole blood, milk, rumen fluid or in tissues. These methods are direct. Selenium may also be measured by an indirect method, assessing the glutathione peroxidase (GPx) activity in erythrocytes. Glutathione peroxidases catalyze the reduction of lipid and hydrogen peroxides to hydroxides via the oxidation and subsequent reduction of selenocysteine (SeCys), which is the active center of the enzyme (Arteel and Sies 2001).

There are reports that whole blood GPx was significantly higher in ruminants offered Se-yeast compared to those that received sodium selenite (Juniper et al. 2008). Similar results were reported by Qin et al. (2007). However, Van Ryssen et al. (1989) and Gunter et al. (2003) found no differences in the GPx activity in whole blood of animals given organic or inorganic selenium.

Previous findings (Mihaliková et al. 2005; Panev et al. 2013) have shown that rumen microbial population is capable of incorporating Se into their cells. Several studies have dealt with the effects of various Se forms on the rumen enzyme activity.

It has been reported that supplementation with sodium selenite together with vitamin E increased production of fatty acids and number of protozoa (Naziroglu et al. 1997). Our previous results showed an increase of the glutamate dehydrogenase enzyme in the ruminal fluid of lambs fed Se-yeast enriched diet with 0.3 mg Se/kg of DM compared to the group given basal diet (Faixová et al. 2007). Mihaliková et al. (2005) reported that selenomethionine has a protective effect on the growth of some protozoa species in young ruminants.

On the other hand, other authors did not find any significant changes after *in vitro* application of selenium (Feasenhaser 2005). It has been also reported that higher Se concentration can have a harmful effect on ruminal microflora and/or ruminal mucosa. The increase in enzymes of ruminal fluid results from damaged ruminal bacteria and/or mucosa in the rumen. In addition, the chemical form of Se in the supplement did not affect ruminal fermentation indicators in dependence on different doses of Se contained in feeding rations. However, a negative effect of a sudden start of feeding high contents of organically bound Se on the infusoria count was reported, which was accompanied by the increase of GMT, LDH and AST enzymes activity in ruminal fluid (Panev et al. 2013).

Several studies have shown that selenium deficiency leads to increased oxidative degradation of compounds susceptible to oxidation. Polyunsaturated fatty acids (PUFA) in biological membranes are highly susceptible to peroxidation. They play an important role in membrane properties such as structure, fluidity, permeability and activities of enzymes located within membranes. Moreover, autooxidative degradation products can damage membranes such as erythrocyte membranes.

Our previous experiment showed that the total erythrocyte count was significantly higher in Se-yeast supplemented lambs than in the control group, and the total white blood cell count was significantly higher in the control group than in the Se-yeast supplemented group (Faixová et al. 2007). According to literature, selenium is essential in regulating haematopoiesis by differentiation of erythroid progenitors of C57/BL6 mice (Kaushal et al. 2011).

The present study evaluates the effect of forms of supplemented selenium on selected indicators of the ruminal fluid and blood in sheep. The aim was to find whether selenium availability of the organic compound is higher compared to the inorganic one and whether higher concentration of selenium in one or the other form affects the rumen enzyme activity and fragility of red blood cells in sheep.

Materials and Methods

The experiment was carried out on 18 sheep of the Valashka breed at the age of 18 months, divided into three groups of six animals and kept on diets that differed in the form of selenium supplement.

Animals were housed in individual pens with free access to fresh potable water and specially prepared trace mineral lick without Se, and were fed diets used for 12 weeks before sampling. The sheep weighed 40 kg at the end of experiment. The composition of mineral lick (g/kg) without Se: calcium (Ca) 16.2, sodium (Na) 316, magnesium (Mg) 32, copper (Cu) 0.7, manganese (Mn) 2.5, zinc (Zn) 3.1, cobalt (Co) 0.06, and iodine (I) 0.02.

The first group (control) was given a basal diet (BD) containing (g/day per animal): 1 500 g grass hay and 500 g ground barley with the Se content of 0.17 mg/kg of dry matter (DM) coming from the natural occurrence of Se in the dietary component only. The second group received basal diet supplemented with 0.4 mg Se/kg of DM of inorganic selenium in the form of sodium selenite (Na_2SeO_3), resulting in 0.53 mg Se/kg of DM. Sodium selenite with 99% purity (Sigma-Aldrich, USA) was dissolved in water and top dressed on ground barley during each feeding. The third group received basal diet supplemented with 0.4 mg Se/kg of DM in the form of Se-yeast extract (Se-Plex, Alltech Inc., USA), resulting in 0.46 mg Se/kg of DM (Table 1).

Table 1. The experimental design and selenium concentration in the experimental groups.

Group/treatment	Group 1 Basal diet	Group 2	Group 3
Selenium source		Sodium selenite (SS)	Se-yeast
Selenium concentration			
Additive ($\text{mg} \cdot \text{kg}^{-1}$ feed)	0.0	0.4	0.4
Analysed ($\text{mg} \cdot \text{kg}^{-1}$ dry matter)	0.17	0.53	0.46

Selenized yeast was directly mixed with ground barley

The major Se-compound of selenized yeast is selenomethionine (SeMet). The preparations of selenized yeast are reported to contain 54–74% of total Se in the form of SeMet, depending on the extraction efficiency of the techniques used by the analytical laboratories (Rayman 2004).

Animals were fed the diets divided into two portions offered at 7.00 h and 15.00 h. The daily ration for all treatments was prepared fresh daily and differed only in the form of selenium supplement. There were no dietary refusals and the feed was completely consumed by animals.

Blood collection

Blood samples were collected from each sheep from vena jugularis into heparinized and non-heparinized tubes. Blood collected in non-heparinized tubes was centrifuged at $2,000 \times g$ for 20 min and the serum was decanted and stored at -65°C until analysis.

Activity of blood glutathione peroxidase (GPx, EC 1.11.1.9) was determined using the method of Paglia and Valentine (1967) with a RANSEL kit (Randox, United Kingdom). Erythrocyte osmotic fragility test was performed by the method of Coles (1986).

Ruminal fluid collection

Ruminal fluid was obtained by stomach tube 2 h after the morning feeding. Duplicate samples of the ruminal fluid were strained through four layers of cheesecloth with 30 min of collection.

The ruminal enzyme activities of alanine aminotransferase (ALT, EC 2.6.1.2) and aspartate aminotransferase (AST, EC 2.6.1.1) were measured by the method of Reitman and Frankel (1957) and glutamate dehydrogenase (GDH, EC 1.4.1.3) was determined by the method of Schmidt and Schmidt (1983) with commercial kits (Randox, Ardmore, UK). The activities of gamma-glutamyl transferase (GGT, EC 2.3.2.1) was determined according to Dimov and Kulhanek (1967) and alkaline phosphatase (ALP, EC 3.1.3.1) was measured using the method of Schmidt et al. (1990) with commercial kits (Pliva-Lachema, Brno, Czech Republic).

Selenium determination

Selenium contents in the feed components, blood, and rumen fluid were analysed fluorimetrically according to the method of Rodriguez et al. (1994).

Statistical analysis

The results are expressed as means \pm standard errors of the means (SEM). Statistical analysis was done by one-way analysis of variance (ANOVA) with Tukey's post hoc multiple comparison test (GraphPad Software, La Jolla, USA).

The experiments were carried out in accordance with the established standards for the use of experimental animals. The protocol was approved by the local ethics and scientific authorities.

Results

Selenium concentration in the rumen fluid and selenium level in blood (Fig. 1) and glutathione peroxidase (GPx) activity in blood (Table 2) were significantly increased compared to the control group. Between experimental groups, the selenium concentration in blood and rumen fluid was higher in the Se-yeast group. There were no differences between groups in the osmotic resistance of red blood cells (Table 2).

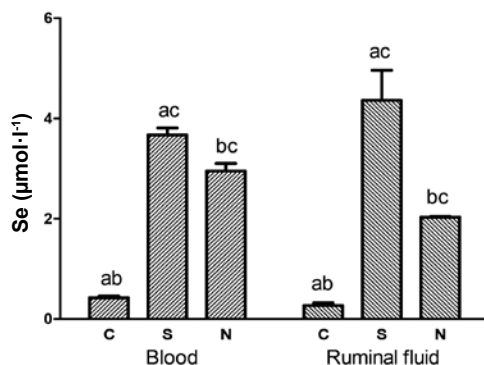


Fig. 1. Selenium concentration in blood and ruminal fluid in control and selenium supplemented groups. Significant differences between columns of respective indicators are indicated by the same letter, $P < 0.05$, results are expressed as the mean of six determinations \pm SEM. C- control group, S – selenized yeast, N – sodium selenite

The source of selenium fed to sheep differently affected the alkaline phosphatase (ALP) activity in rumen fluid; the Se-yeast supplemented diet significantly decreased, whereas the Na_2SeO_3 -supplemented diet significantly increased the enzyme activity.

Activities of gamma-glutamyl transferase (GGT) and glutamate dehydrogenase were significantly increased in the rumen fluid of sheep supplemented with Se-yeast and the aspartate amino transferase activity was significantly higher in sheep fed the diet supplemented with Na_2SeO_3 . Alanine amino transferase activity in the rumen fluid was not affected by the treatment (Table 3).

Table 2. The effect of supplemental selenium on the osmotic resistance of red blood cells and the glutathione peroxidase activity in blood.

	Control (basal diet)	Selenized yeast (Sel-Plex)	Na_2SO_3
The osmotic resistance of RBCs			
a) minimal	0.59 ± 0.015	0.63 ± 0.10	0.64 ± 0.015
b) maximal	0.44 ± 0.015	0.46 ± 0.016	0.50 ± 0.022

Significant differences within a row are indicated by the same superscript letter, $P < 0.05$. Results are expressed as mean of six determinations \pm SEM.

RBC – red blood cell, GPx – glutathione peroxidase

Discussion

Rumen

Selenium concentration in the total rumen fluid was significantly higher in sheep supplemented with selenium from the organic (Se-yeast) or the inorganic (Na_2SeO_3) source in comparison with the control group of sheep. Comparing Se-supplemented groups, the

Table 3. The effect of supplemental selenium on the enzyme activity and selenium content in the ruminal fluid of sheep.

Indicator	Control (basal diet)	Selenized yeast (Sel-Plex)	Na ₂ SO ₃
GGT (μkat·l ⁻¹)	1.26 ± 0.046 ^a	3.48 ± 0.210 ^{ab}	1.77 ± 0.124 ^b
ALT (μkat·l ⁻¹)	1.57 ± 0.10	1.84 ± 0.03	1.79 ± 0.04
AST (μkat·l ⁻¹)	2.76 ± 0.060 ^a	2.735 ± 0.101 ^b	3.33 ± 0.128 ^{ab}
GDH (μkat·l ⁻¹)	0.25 ± 0.012 ^a	0.38 ± 0.015 ^{ab}	0.29 ± 0.014 ^b
ALP (μkat·l ⁻¹)	1.37 ± 0.067 ^{ab}	0.95 ± 0.061 ^{bc}	2.27 ± 0.087 ^{ac}

Significant differences within a row are indicated by the same superscript letter, $P < 0.05$, results are expressed as the mean of six determinations ± SEM.

GGT – gamma-glutamyltransferase; ALT – alanine aminotransferase; AST – aspartate amino transferase; GDH – glutamate dehydrogenase; ALP – alkaline phosphatase

selenium concentration in the rumen fluid was significantly higher in the Se-yeast group.

Selenium added to the feeding ration is not fully utilized by the animal's organism, as its significant part is absorbed and metabolized by ruminal bacteria, thus the highest selenium concentrations are found in ruminal biomass, whereas the lowest are in the ruminal fluid (Serra et al. 1994; Panev et al. 2013).

In ruminants, inorganic Se compounds are partly reduced by ruminal microbes to unabsorbable elemental selenium resulting in a lower Se apparent absorption. Selenoamino acids are subject to ruminal microbial degradation also decreasing apparent selenium absorption from the digestive tract (Van Ryssen and Schroeder 2003).

It is well established that dietary composition influences the microbial population in the rumen which in turn can modify the availability of dietary Se for absorption and also its subsequent retention in the body of ruminants.

Rumen microorganisms have been shown to incorporate Se from both inorganic and organic forms into their own protein and into the microbial cell wall components (Koenig et al. 1997).

There is some evidence indicating that Se metabolism in the rumen is dependent on the prevalence of particular species of rumen bacteria. It has been demonstrated that pure cultures of *Selenomonas ruminantium* and *Butyrivibrio fibrisolvens* incorporated Se into selenoamino acids whereas *Bacteroides rumenicola* reduced seleno-compounds into elemental Se only (Hudman and Glenn 1985).

It has been suggested that the release of selenomethionine from Se-yeast proteins in the digestive tract should correspond to the digestion of other total proteins in ruminants. Factors affecting the ruminal degradation of proteins such as heat processing of feeds can therefore modify the fate of Se compounds in the rumen and affect the proportion of Se amounts available for microbial reduction or incorporation into their own proteins and enable Se-containing compounds to escape rumen microbial fermentation (Van Ryssen and Schroeder 2003). Mihaliková et al. (2005) reported that Se supplementation showed protective effects against the development of some ciliate species in young ruminants. The inclusion of Se-yeast into sheep's feed had a more pronounced effect on the population of *Ophryoscolex* and *Diploplastron* than feed supplementation with an equivalent amount of sodium selenite. The most sensitive rumen ciliate genera to Se dietary intake appear to be *Ophryoscolex* which did not even occur in sheep with deficient Se intake.

The protective effect of Se on the rumen microflora was demonstrated in sheep fed a basal diet (BD) containing 0.07 mg Se/kg of DM, while the experimental group was supplemented with Se at a dose of 0.3 mg/kg of dry matter in the form of sodium selenite. Se level and glutathione peroxidase (GPx) activity was significantly higher and malondialdehyde (MDA) concentration was significantly lower in bacterial and protozoal fractions of the rumen fluid in Se-supplemented animals, which confirms the ability of rumen micro-

flora to accumulate dietary Se, and the effectiveness of rumen microflora to utilize Se from sodium selenite and to use it for synthesis of the seleno aminoacid Se-cysteine. Se-amino acids incorporated into microbial proteins are highly available sources of selenium for their absorption in the lower part of the gastrointestinal tract (Čobanová – Boldižárová et al. 2008).

Se-yeast supplemented diet significantly affected rumen fluid enzyme activity and increased the activity of enzymes involved in nitrogen metabolism in rumen (e.g., GGT and GDH). Our results show that the effect of selenium supplementation on ruminal enzyme activity depends on the selenium form. Gamma-glutamyl transferase and glutamate dehydrogenase were significantly higher in the Se-yeast supplemented group, and aspartate amino transferase and alkaline phosphatase were significantly higher in the selenite supplemented one.

There are several reports available on the relationship between dietary Se and rumen fermentation. Kim et al. (1997) found that Se (as SeMet, 2 mg/kg) can stimulate *in vitro* fermentative activity of rumen microorganisms compared to the addition of elementary selenium or sodium selenite, or no selenium supplementation. Naziroglu et al. (1997) reported an increase of total volatile fatty acids and propionate production, the total counts of protozoa of the ruminal fluid of lambs supplemented by 0.3 mg Se/kg diet (Na_2SeO_3) *in vitro*. The ammonia nitrogen content was not affected by the Se-treated diet. Similarly, Liu et al. (2007) reported reduction of ratio of acetate to propionate due to an increase in propionate production when steers were fed 300 and 600 mg Se-yeast/kg dietary DM.

Later Xun et al. (2012) reported that ruminal pH, ammonia N concentration, molar proportion of propionate, and ratio of acetate to propionate were decreased ($P < 0.01$) and total VFA concentration were increased with 4 g nano-Se (4 mg Se/kg of dry matter) and 4 g Se-yeast supplementation (provide 4 mg Se/kg of DM) in ruminally cannulated sheep. This agrees with the results of Shi et al. (2011) who obtained higher VFA concentration and lower pH in the sheep fed with the basal diet supplemented with 3 g of nano-Se (provide 3 mg Se/kg of DM) in sheep. Similar results were also found in cows (Wang et al. 2009).

However, Serra et al. (1994) showed that the concentration of total volatile fatty acids VFA and propionate decreased slightly by administration 0.2 mg Se (Na_2SeO_3 or Na_2SO_4) per kg of DM in sheep. Dutt and Chhabra (2008) report that supplementation of 0.5 ppm selenium either in the inorganic or the organic form had no significant effect on the nutrient intake, rumen microbial population, and protein synthesis in cattle and buffaloes.

The effect of selenium supplementation on ruminal enzyme activity in the present study may be explained as follows. We did not evaluate other indicators of rumen fermentation or feed digestibility so we can only speculate on the hypothesis.

According to literature reports, supplementation of selenium in organic (Se-yeast) in cows (Wang et al. 2009) or Se-yeast and nano-Se in sheep (Xun et al. 2012) in the diet switched the rumen fermentation pattern from acetate to propionate and decreased the ammonia N concentration, which suggested that selenium supplementation in diet particularly increased microbial activity in propionate production. In addition, Xun et al. (2012) reported a decrease of concentration of volatile fatty acids with high selenium-yeast versus medium selenium-yeast, indicating the increase of Se-yeast dose from 0.3 mg Se/kg of dietary DM to 0.45 mg Se/kg of dietary DM was not beneficial to further increase of feed fermentation in the rumen.

It can be suggested that stimulation of microbial activity by Se-yeast may be in dose-dependent.

In addition, Panev et al. (2013) studied the effects of various doses of Se on selected variables of the ruminal fluid and blood in sheep. They reported that the form of supplemented Se did not have a significant effect on ruminal fermentation indicators (pH, total acidity, volatile fatty acids, and ammonia) except for the negative effect of the

sudden start of feeding high contents of organically bound Se on the infusoria count, which was accompanied by an increase of gamma-glutamyl transferase, lactate dehydrogenase, and aspartate amino transferase activity in the ruminal fluid. It can be suggested that enhanced growth of the ruminal microbial population by Se-yeast could increase the ammonia nitrogen consumption and increase the activity of enzymes of nitrogen metabolism in the rumen fluid such as glutamate dehydrogenase and gamma-glutamyl transferase found in the present study. An increase of the glutamate dehydrogenase enzyme in the ruminal fluid was reported in our previous study on lambs fed a Se-yeast enriched diet with 0.3 mg Se/kg diet DM compared to the group given basal diet (Faixova et al. 2007).

Selenium in blood

Blood Se concentration was significantly higher in sheep supplemented with selenium from the organic (Se-yeast) or inorganic (Na_2SeO_3) source in comparison with the control group of sheep. However, between experimental groups the Se concentration in blood was significantly higher in the Se-yeast group.

This finding is in agreement with the study by Hall et al. (2012) who studied the effects of the Se source (inorganic vs. organic) and supplementation rate (the Food and Drug Administration, FDA vs. supranutritional rates) on whole – blood and serum – Se concentrations in mature ewes. Their results showed that the dose range whereby Se supplementation increased blood Se concentrations was more limited for the inorganic Na-selenite than for the organic Se-yeast. The smallest rate (FDA recommended quantity) of organic Se supplementation was equally effective as the supranutritional rates of Na-selenite supplementation in increasing whole blood Se concentrations, demonstrating greater oral bioavailability of organic Se.

Rock et al. (2001) reported that Se supplementation in pregnant ewes either by sodium selenite or selenized yeast significantly increased the Se concentration in whole blood and serum, increased the activity of glutathione peroxidase (GPx) in both the pregnant ewes and the lambs of ewes given Se. When the two Se supplements were compared, lambs of the ewes which were fed the Se-yeast had higher ($P < 0.001$) concentrations of Se and activities of GPx ($P < 0.05$) in blood than lambs of the ewes fed selenite.

Similarly, there were significant differences ($P < 0.05$) between the two Se sources in whole blood Se concentration, with values being higher in the Se-yeast supplemented animals (0.30 mg Se/kg of DM) compared to those that received a comparable dose of sodium selenite, indicating a better uptake and incorporation of Se derived from Se-yeast in beef cattle (Juniper et al. 2008). These observations are in line with those of Van Ryssen et al. (1989) in sheep, Falkowska et al. (2000) and Gunter et al. (2003) in cows, Ortman and Pehrson (1999) in dairy cows and Pechova et al. (2012) in goats, who reported higher whole blood Se concentrations in animals supplemented with organic Se compared to those administered diets of a comparable Se content where sodium selenite was the source of supplementary Se.

On the other hand, Pavlata et al. (2011) reported that Se supplementation in goats either by sodium selenite or the new organically bound (lactate-protein complex) form of selenium for three months significantly increased the concentration of selenium in blood and no differences were found regarding the source of Se. However, the GPx activity increased much faster with selenite, which therefore appears to be a more biologically available form of Se for the synthesis of biologically active selenoproteins.

Glutathione peroxidase activity in blood

Our results showed that the activity of GPx in blood was significantly elevated in both groups supplemented with sodium selenite or Se-yeast compared to the animals fed the control diet. There were no differences between the animals given inorganic or organic

selenium. Similar results were reported by other authors in sheep (Van Ryssen et al. 1989; Gunter et al. 2003).

In contrast to our results, other authors reported that whole blood GPx was significantly higher in beef cattle offered Se-yeast compared to those that received a comparable dose of sodium selenite (Juniper et al. 2008). Qin et al. (2007) also showed a significantly higher whole blood selenium concentration and GPx activities in lambs fed Se-yeast enriched diet (0.10 mg/kg of diet) compared to lambs fed similar concentrations of sodium selenite. The differences in results obtained by the researchers might be explained by the fact that although whole blood Se concentration is closely correlated with the GPx activity (Brigelius-Flohe et al. 1994), there is evidence that the percentage of Se in erythrocytes associated with GPx may vary depending on the Se source (Belstein and Whanger 1986; Van Ryssen et al. 1989).

Osmotic fragility of red blood cells

Erythrocyte osmotic fragility was not significantly affected by selenium supplementation.

In agreement with the current study, Saini et al. (2007) reported no effect in the membrane permeability and physicochemical alteration of erythrocytes in the presence of Se and vitamin E in goats.

In contrast, fragility of erythrocyte membranes was significantly decreased in SeO₂-treated rats compared to control (Chen and Lin 2000). In addition, Li et al. (1990) reported that the Se concentration in blood of children living in the endemic areas of the Kaschin-Beck disease (KBD) was lower than that in the non-endemic areas. Moreover, the fragility of erythrocytes was increased, indicating membrane defects and membrane damage in KBD. Recently Kaushal et al. (2011) suggested that Se is essential in regulating haemopoiesis by differentiation of erythroid progenitors in C57/BL6 mice.

These discrepancies may be due to differences in species bioavailability of Se (Pavlata et al. 2001), the amount and chemical form of supplemental Se (Pavlata et al. 2005) and erythrocyte osmotic fragility between species.

In conclusion, feeding diets supplemented with selenium from organic or inorganic sources significantly increased the Se concentrations in the total rumen fluid and blood, and the blood GPx activity. Comparing comparable doses from different Se sources, Se concentration in the blood and total rumen fluid was higher in the Se-yeast group. However, there were no differences in the blood GPx activity between the Se sources, indicating that Se from sodium selenite was as effective as Se from Se-yeast in the availability for the blood GPx activity of sheep. The effect of Se supplementation on the ruminal enzyme activity depends on the Se form – GGT and GDH were significantly higher in the Se-yeast supplemented group, and AST and ALP were significantly higher in the selenite supplemented group. These preliminary results contribute to existing knowledge about the effects of Se supplementation on the ruminal enzyme activity in sheep. Further studies are needed in order to clarify the mechanisms of various Se forms on the rumen fermentation processes.

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