EFFECT OF FOOD INTAKE, FASTING, DEULECTOMY AND ENVIRONMENTAL TEMPERATURE ON SATURATED AND UNSATURATED FATTY ACID DISTRIBUTION IN THE BLOOD PLASMA OF COCKERELS IN THE FIRST WEEK AFTER HATCHING

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Received April 8, 1980

Abstract


The effect of food intake, fasting; ambient temperatures of 18°C and 35°C and deuctectomy on the distribution of saturated and unsaturated fatty acids in the blood plasma of cockerels was investigated in 200 birds aged 1 to 5 days. Total plasma fatty acids were determined by gas chromatography.

In newly hatched cockerels the quantitatively most important (C14 - C30) saturated fatty acids accounted for 68.2 ± 4.2 % and unsaturated acids for 27.1 ± 4.7 % of total fatty acids.

This distribution changed considerably within the first week after hatching in that the saturation of fatty acids decreased at a rate obviously dependent on the onset of food intake: it started to change later in fasted than in fed birds. Environmental temperature and deuctectomy did not affect these changes.

Fatty acids (C14 - C30), age-dependent changes, saturation.

Fatty acids serve mainly as an energy source to the body. Small amounts of fatty acids (FA) are used in addition for other purposes such as membrane formation or prostaglandin synthesis (Lindsay 1975). Data on their role in lipid metabolism of birds are rather scarce; their contribution to egg lipids of the domestic fowl and other avian species were studied e. g. by Christie and Moore (1972). The bulk of fatty acids in body fats of various migrating avian species was found to consist of oleic (36 %) and palmitic (21 %) acids (Caldwell 1973). In depot fats of adult fowls 69 % unsaturated and 31 % saturated fatty acids was found (Feigenbaum and Fisher 1959).

There are no data, however, on the effects of age, fasting, and various ambient temperatures on the distribution of saturated and unsaturated fatty acids in the blood plasma of chickens. We therefore investigated this distribution in newly hatched cockerels and followed its development with age in fed and fasted birds and the effects of deuctectomy and two ambient temperatures.

Materials and Methods

In the experiment 200 newly hatched Shaver Starcross cockerels were employed. The birds were kept and treated as previously described (Baranyiová, Standara 1980).

Five birds each of all experimental groups were blood-sampled invariably at the same time of the day after minimum handling on days 1, 2, 3, 4 and 5. The blood plasma samples for determination
of total fatty acids were processed according to Hammarstrand (1966). The methyl esters of fatty acids were purified on Florisil (Johnson and Davenport 1971). Gas chromatograph Chrom IV (LP Praha) equipped with a flame ionization detector and flash heater temperature of 235 °C was used for determination of methyl esters of fatty acids. A stainless steel column (2.5 m × 3 mm) packed with 5 % DEGS on Inerton AW-DMCS (100—125 mesh) (Lachema Brno, CSSR) was run isothermically at 195 °C. Qualitative analysis was based on relation between the elution time and number of carbon atoms in the molecules of individual homologues (Purnell 1967; Barvíř et al. 1968), for quantitative analysis the method of internal normalization was employed (Forman 1970). The contents of NEFA (C_n—C_m) are expressed as % w/v of total fatty acids. Results were statistically evaluated by Student’s t-test.

Results

In the blood plasma of newly hatched chickens 68.2 ± 4.2 % saturated and 27.1 ± 4.7 % unsaturated fatty acids was found (Fig 1). As soon as 24 hours later a sharp decrease (P < 0.001) of saturated fatty acids and an increase (P < 0.01) of unsaturated fatty acids occurred (33.1 ± 1.0 and 54.7 ± 2.2 % respectively) and this distribution of fatty acids remained practically unchanged until day 5, with significantly more unsaturated acids (P < 0.001; P < 0.001; P < 0.001; and P < 0.02) in the plasma of intact fed chickens. A similar distribution of fatty acids was also found in intact chickens exposed to 18 °C for one hour (P < 0.001; P < 0.001; P < 0.01; —). This pattern of development was also observed in deutectomized birds, a significantly larger proportion of unsaturated fatty acids being observed at both temperatures on days 3, 4 and 5 (P < 0.001; P < 0.01; P < 0.001) (Fig. 2).

In the plasma of fasted intact chickens (Fig. 3), kept at 35 °C the proportions of the two groups of fatty acids began to change slowly between days 2 and 3 so that a smaller proportion of saturated (34.3 ± 1.6 %) and a larger proportion of unsaturated (52.4 ± 1.7 %) FA vas found on day 5. Almost the same values

![Graph 1](image1)

Distribution of saturated (closed symbols) and unsaturated (open symbols) fatty acids in the blood plasma of intact fed chickens

![Graph 2](image2)

Distribution of saturated (closed symbols) and unsaturated (open symbols) fatty acids in the blood plasma of deutectomized fed chickens
were obtained in birds exposed to 18 °C. In deutectomized fasted cockerels kept at 35 °C the development was similar as in the previous group. In deutectomized fasted birds exposed to 18 °C the shift in the proportions of FA began to occur between day 3 and 4 (Fig. 4).

**Discussion**

In newly hatched chickens the quantitatively most important saturated fatty acids (C<sub>14</sub>-C<sub>20</sub>) averaged 68 % and the unsaturated ones about 27 %. However, using the data reported by Christie and Moore (1972) for triglyceride composition of Shaver Starbro and Shaver Starcross eggs we calculated that in their study the reverse was the case: saturated fatty acids accounted for 33.9 and 36.1 % and unsaturated acids for 66.1 % of total fatty acids, respectively. Similarly, in depot fats of adult hens Feigenbaum and Fisher (1959) found 32 % saturated and 67 % unsaturated fatty acids. Nevertheless, there is evidence to indicate that intensive transformation and synthesis of lipid components occurs during the embryonic development (Budowski et al. 1961; Noble and Moore 1964, 1967) and that the developing embryo preferentially absorbs from the yolk a phosphatidyl ethanolamine fraction that is relatively rich in polyunsaturated docosahexaenoic acid (Noble and Moore 1965). Moreover, the fatty acid synthetase activity was found to be low in chick liver during the entire embryonic development (Joshi and Sidbury 1975).

Our finding of a larger proportion of saturated fatty acids along with a smaller proportion of unsaturated acids immediately after hatching may reflect specific metabolic events in the embryo leading to exhaustion of its unsaturated fatty acid supplies.

This view is further supported by the fact that having changed significantly
during the first 24 hours after hatching the proportions of saturated and unsaturated fatty acids in all groups of fed birds became similar to the values reported by Feigenbaum and Fisher (1959) for adult fowls. Also the fatty acid synthetase increased by about three-fold on hatching and thereafter in fed, newly hatched chickens by about 35-fold, over the basal embryonic activity (Joshi and Sidbury 1975).

The data on distribution of saturated and unsaturated fatty acids in blood plasma of chickens in the first posthatching week reported here provide the basic information about the distribution and movement of these metabolites in the internal environment in the early post-incubation period of the domestic fowl.

In the two groups of fasted cockerels the shift in the proportions of saturated and unstaturated fatty acids did occur too, though later, and reached values similar to those found in the fed birds not until day 5. This finding indicates that the mechanisms involved in induction of post-hatching fatty acid synthesis begin to operate at or in consequence of hatching and are therefore not related to food intake only. This conclusion is supported by data of Joshi and Sidbury (1975). Exposure to 18 °C did not affect the changes in FA saturation in intact cockerels. In deutectomized fasted birds the change in saturation of plasma fatty acids was delayed as compared with intact fasted birds.

In conclusion in can be stated that saturation of fatty acids in the blood plasma of chickens decreases considerably within the first five post-hatching days, its rate depending on the onset of food intake. This process was not affected by either environmental temperature or deutectomy.

Vplyv príjmu potravy, hladovania, deutektómie a teploty prostredia na obsah nasýtených a nenasýtených mastných kyselín v krvnej plazme kurčiat v prvom týždni po vyliahnutí

Na 200 kohútikoch línie Shaver Starcross sme sledovali vplyv príjmu potravy, hladovania a teploty prostredia (18 °C a 35 °C) na obsah plazmatických nasýtených a nenasýtených mastných kyselín (MK) od 1. do 5. dňa po vyliahnutí. Čelkové mastné kyseliny sme stanovovali pomocou plynovej chromatografie.

U kurčiat po vyliahnutí činil podiel kvantitatívne najvýznamnejších (C₁₄—C₂₀) MK nasýtených 68.2 ± 4.2 %, nenasýtených MK 27.1 ± 4.7 %. V priebehu prvého týždňa po vyliahnutí sa však tento podiel MK v plazme podstatne zmenil, a to tak, že saturácia mastných kyselín klesla. Rýchlosť tejto zmeny je zrejme závislá na zahájení príjmu potravy; u hladujúcich kurčiat sa objavila neskôr ako u kŕmených. Teplota prostredia ani deutektómia ju však neovplyvnila.

Влияние приема пищи, голодания, дейтерэктомии и температуры окружающей среды на содержание насыщенных и ненасыщенных жирных кислот в кровной плазме цыплят на первой неделе

На 200 петушках линии Shaver Starcross нами проводились наблюдения за влиянием приема пищи, голодания и температуры окружающей среды на содержание плазматических насыщенных и ненасыщенных жирных кислот с первого по пятый дни после выплупивания. Общие жирные кислоты нами были определены с помощью газовой хроматографии.
У цыплят после вылупления доля качественно самых значимых (C₁₄—
C₂₀) насыщенных жирных кислот составляла 68,2 ± 4,2 %, ненасыщенных жирных кислот 27,1 ± 4,7 %.
В течение первой недели после вылупления она, однако, существенно меняется, а именно так, что их сатурация понижается. Скорость данного изменения зависит от начала приема пищи, температура окружающей среды на нее не оказывает влияния, у голодащих она появляется позже, чем у кормленных цыплят.

References