EFFECTS OF BASIC ABIOTIC FACTORS ON DEVELOPMENT OF THE EGGS AND ON SURVIVAL OF THE INFECTIVE LARVAE OF COMMON HELMINTHS OF SHEEP IN THE EXTERNAL ENVIRONMENT

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Abstract

H o l a s o v á E v a, I. P a v l á s e k, B o ž e n a K o t r l á : Effects of Basic Abiotic Factors on Development of the Eggs and on Survival of the Infective Larvae of Common Helminths of Sheep in the External Environment. Acta vet. Brno, 57, 1988: 153-168.

Development of the eggs of nematodes of the genera <u>Stron-gyloides</u>, <u>Haemonchus</u>, <u>Trichostrongylus</u>, <u>Ostertagia</u> and <u>Chabertia</u> to the infective larval stage was observed on 50 x 50 cm grass-covered plots set out experimentally at monthly intervals during one year. The development was most rapid, taking 5 to 15 days, in May to September at mean monthly air temperatures of 15.6 to 23.8°C. The infective larval stage was reached even at a mean monthly air temperature of 0.6°C and relative humidity of 76.3% in 65 days. No infective larvae were found in sheep faecal samples set out in November to February.

The survival time of the infective larvae of nematodes of the genera <u>Haemonchus</u>, <u>Trichostrongylus</u>, <u>Ostertagia</u> and <u>Chabertia</u> in the external environment was as long as 11 months. In summer months the larvae were destroyed in 2 to 4 months in consequence of a higher air temperature and lower relative humidity. Lower temperatures combined with higher relative humidity were withstood better: the first signs of destruction were observed after 5 to 7 months.

Repeated administration of Nilverm to naturally-infected sheep did not affect the rate of development of the larvae to the infective stage and probably did not even impair their viability.

Infective larvae, external environment, survival of infective larvae, NILVERM.

Table 1

Development of the eggs of gastrointestinal nematodes to the infective larval stage

s of nematodes of the genera Air temperature Soil temperature Relative Str. Haem. Ost. Trich. Chab. (°C) 2 cm below the air humidity (°C) (°C) (°C) (°C)	od min max od m	21.5 · 15.8 10.1 19.5	16.1 10.0	30.7 23.8 21.0 28.0 25.3 38.3 65.7 9	25.0 22.9 18.2 22.5 21.0	15.6 9.8 14.7 11.7 47.3 88.3	11.7 0.6 -14.2 7.5	9.0 -2.4 -14.7 4.0 -4.4 63.3 90.0	7.7 1.3 -7.0 3.0 -2.2 43.7 89.3	2.2 -2.9 -8.7 -2.0 -6.1 60.0 86.0	3.7 -2.4 -10.2 -2.5 -4.9 53.3 85.0	-3.0 11.0 2.7 43.3	18.7 9.0 2.7 14.2	11 - 11.7 19.5 15.0 11.5 20.5 15.7 46.7 84.3 60.9	Str Strongyloides Haem Haemonchus Ost Ostertagia Trich Trichostrongylus Chab Chabertia
Soil te 2 cm h	min	10.1	10.0	21.0	18.2	9.8	-14.2	-14.7	-7.0	-8.7	-10.2	-3.0	2.7	11.5	Ē
ture	6	• 15.8	16.1	23.8	22.9	15.6	0.6	-2.4	1.3	-2.9	-2.4	5.1	0.0	15.0	''
tempera (°C)	max	21.5	23.0	30.7	25.0	21.7	11.7	0.0	7.7	2.2	3.7	13.2	18.7	19.5	
Air	min	8.2	0.0	18.5	19.7	12.0	-10.5	-11.7	-4.5	-8.2	-10.7	-1.7	3.5	11.7	ongyloi(ertagia abertia
genera Chab.	nt to (days)	ı	15	15	13	14	I	o	o	o	0	ı	I	ı	'
f the f Trich.	elopmen stage	15	15	12	11	13	65	0	0	0	0	40	27	11	
codes c Ost.	ation of the deve infective larval	15	11	13	11	14	65	0	0	0	0	07	27	11	e found
nemat Haem.	n of t ctive	12	11	12	11	14	65	0	0	0	0	ı	ı	I	rvae were occurred
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Egg: Month	цр Тр	٨	ΝI	ΛII	ΛIII	XI	x	IX	XII	I	II	III	ΛI	٨	no infective larvae were found no development occurred
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154

Parasitic nematodes of the sub-order <u>Strongylata</u> are characterized by a wide range of geographic distribution resulting from their ability to adapt themselves to various environmental factors. Recent adoption of intensive husbandry practices in animal and vegetable production has brought changes in the environmental conditions in which the development of parasites occurs. In spite of major progress achieved in the control of helminths parasitic in the gastrointestinal tract of farm ruminants environmental factors including pasture, the density of animal populations in large herds and flocks and the use of dung as manure for growing various plants to be fed to animals are still much-feared sources of helminth infections. Further studies are therefore needed to investigate the development, survival and viability of the infective larvae of individual helmith species under various outdoor conditions.

The first comprehensive study on survival of the larval stages of parasitic worms on pasture in Czechoslovakia were reported by E r h a r d ov á (1961) who found that helminth larvae survived winter, the most resistant being those of the genera <u>Ostertagia</u>, <u>Nematodirus</u> and <u>Chabertia</u>; the infective larvae of these nematodes were viable even if the environmental temperature fell to -30 °C. The bulk of relevant literature shows that in studies on ecological adaptation of the free-living stages of helminths it is necessary to consider the effects of both temperature and relative humidity. According to a number of writers the optimum temperature for the development of helminth larvae is 20 to 30 °C (G u p t a 1961; W a l l a c e 1961), averaging 25 °C. T r a č (1975) reported that strongylate eggs developed in the temperature range between 4 to 5 °C and 35 to 41 °C. The relative humidity required for development of the eggs ranges reportedly between 20 and 50%. B a s h i r o v (1979) found that eggs of the genera <u>Ostertagia</u> and <u>Trichostrongylus</u> survived at -16 and -25 °C and that in April the larvae developed within one month at an average month temperature of 6.4 °C and were, according to his view, the source of infection of sheep and cattle in May or July.

Development of a new helminth infection is possible only where infective larvae are present. Therefore the survival and duration of viability of the infective larvae of most pathogenic helminth species continues to be studied from various points of view. Although the results published so far have often been contradictory, the evidence thus obtained contributes to a better understanding of the principles underlying the development and survival of the free-living stages of parasites in the external environment.

The present study was designed to assess the effects of basic ecological factors on the rate of development of the infective larvae of the helminths occurring most frequently in our large herds of cattle and flocks of sheep and test their survival during a one-year period. Consideration was also given to NILVERM, an anthelmintic commonly used to treat naturally-infected animals, as to its effects on the course of development of the eggs to the infective larval stage.

Materials and Methods

Effects of basic abiotic factors on the development of nematodes of the genera <u>Strongyloides</u>, <u>Haemonchus</u>, <u>Ostertagia</u>, <u>Trichostrongylus</u> and <u>Chabertia</u> were investigated during one year. Values of some meteorological elements (Fig. 1) were obtained from a climatological station about 8 km distant from the observation place and were used to supplement our own observations on air temperature, relative humidity and soil temperature conducted on the spot (Table 1). 1. Methods used to assess the rate of development of the eggs to the infective larval stage and the survival of the infective larvae in dependence on environmental outdoor conditions

The study was carried out on a field workplace of the Parasitological Institute of the Czechoslovak Academy of Sciences at 425 m above sea level. At monthly intervals sheep faecal pellets positive for the presence of eggs of the aforementioned helminths were placed in 1 kg quantities in the middle of small (50 x 50 cm) experimental plots to occupy in each plot a circular area of about 20 cm in diameter. In this way a total of 13 experimental plots were set out on sand-and-clay soil with grass allowed to reach maximally 10 cm in height.

From each plot sheep faecal samples of circa 3 to 5 g were collected daily till the first infective larvae appeared. Their survival and morphology were investigated at monthly intervals during the following period.

2. Methods used to investigate the effects of administration of NILVERM (Bioveta, Czechoslovakia) on the rate of development of the eggs till the larval infective stage in sheep naturally infected with nematodes of the genera <u>Trichostrongylus</u>, <u>Ostertagia</u>, <u>Chabertia</u>, <u>Strongyloides</u> and <u>Haemonchus</u>

A total of 7 sheep (3 males and 4 females) were included in the experiment. Faecal samples were taken from all of them repeatedly and examined by culture before NILVERM administration. The males were injected with NILVERM at the dose of 4 ml per animal and the ewes served as controls. Two males in which nematode eggs reappeared in the faeces were treated repeatedly with the same dose of NILVERM, one on day 28 and then again on day 43 and the other one on day 81 after the first treatment. All the sheep were faeces-sampled individually from the rectum three times a week throughout the observation period of 132 days.

3. Methods used for examination and culture of sheep faecal samples

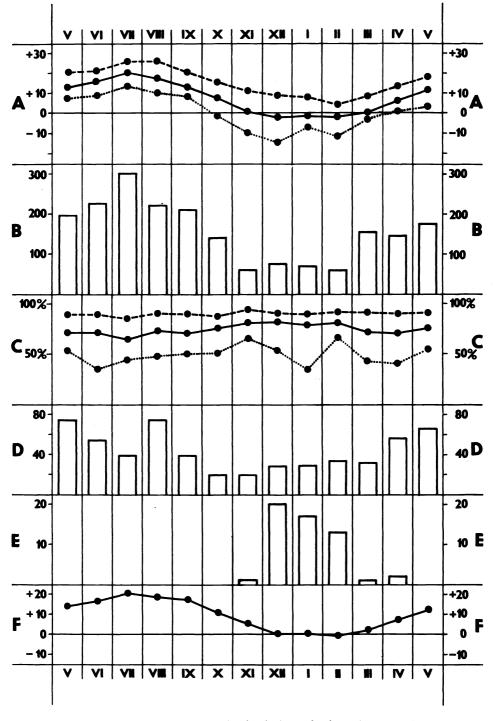
- a Coprological method based on flotation and centrifugation according to Breza (1957) using 1 g faeces.
- b Infective larvae of the helminths were obtained from the faeces on the individual experimental plots by a modification of the Baermann technique.
- c To test the effects of NILVERM on the rate of development from the eggs to the infective larval stage, the faeces from naturally-infected sheep treated with NILVERM were incubated under laboratory conditions at 25°C and 100% relative humidity. Infective larvae were recovered by a modification of the Baermann technique.

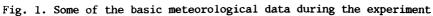
Results

1. The rate of development of helminths of the sub-order Strongylata from the eggs to the infective larval stage

The rate of development from the eggs to the infective larval stage in dependence on the air and soil temperatures and relative humidity is shown in Table 1. It can be seen







that in May to September the development to the infective larval stage occurred most rapidly (taking 5 to 7 days) in <u>S. papillosus</u>. Larvae of the remaining species (<u>H. contortus</u>, <u>Trichostrongylus</u> spp., <u>Ostertagia</u> spp. and <u>C. ovina</u>) began to hatch on days 3 to 4 and 2nd stage larvae were observed on days 6 to 8. The first infective larvae (<u>H. contortus</u> and <u>Trichostrongylus</u> spp.) developed 11 to 15 days after the experiment was started. During this period the mean monthly temperature ranged from 15.6°C (September) to 23.8°C (July) and the relative humidity ranged from 50.3% (July) to 67.2% (June).

In the experiment set up in October the first solitary larvae hatched on day 20 and the infective larvae (<u>H. contortus</u>, <u>Ostertagia</u> spp. and <u>Trichostrongylus</u> spp.) were first observed on day 65. Almost half of the embryonated eggs did not develop any further. The mean air temperature was 0.6 °C (ranging from -10.5 to 11.7 °C) and the mean relative humidity was 75.3%.

In the experiment set up in November some signs of embryonation were observed in <u>H. contortus</u>, <u>Ostertagia</u> spp., <u>Trichostrongylus</u> spp. and <u>C. ovina</u>. Larvae within the eggs (some of them motile) were observed very rarely on day 12. The process of embryonation occurred at the time that the mean daily temperature ranged between 3.2 and 9.0°C, which lasted only four days (days 8 to 11 after the experiment was started). On the remaining days the air temperature was low, falling below zero. The mean air temperature was -2.4°C and the mean relative humidity was 77.7%.

Fig. 1.	Abscissa:	<pre>some basic meteorological elements months of the year A - air temperature (°C)</pre>
		B - hours of solar radiation
		C - relative humidity (%) ——— mean monthly relative humidity (%) maximum monthly relative humidity (%) minimum monthly relative humidity (%)
		D - total precipitation (mm)
		E - No. days with snow cover
		F - mean monthly soil temperature measured 2 cm beneath the soil surface (°C)

In the experiment set up in December the presence of larvae was found in 50% of the eggs on day 4. These larvae were non-motile and did not develop any further. The mean air temperature was 1.3°C and the mean relative humidity was 75.6%.

In the experiment set up in January and February daily examination of the faecal samples revealed no development of the larvae within the eggs during the 30-day observation period. The mean air temperatures were -2.9 and -2.4°C and the relative air humidity values were 69 and 69.8%, respectively. In the four last-mentioned months (November to February) no infective larvae were found at the following monthly examinations which were carried out till June.

In the experiment set up in March the first signs of embryonation were observed on day 5. Beginning day 9 hathing occurred sporadically and the first infective larvae of <u>Trichostrongylus</u> spp. and <u>Ostertagia</u> spp. appeared on day 40 after the start of the experiment.

In the experiment set up in April embryonation began on day 3 and the first infective larvae (<u>Ostertagia</u> spp.) were observed on day 27. The mean air temperature was $9^{\circ}C$ and the relative humidity was 60.9%.

In the experiment set up in May the first infective larvae of <u>Trichostrongylus</u> spp. were found on day 11, which is in keeping with the observations made in May of the preceding year.

2. Viability and survival of the infective larvae

As can be seen from Table 2 the infective larvae of <u>H</u>. <u>contortus</u>, <u>Trichostrongylus</u> spp. and <u>Ostertagia</u> spp. from experimental plots (set out in May and June) showed reduced motility and the first vacuoles in their bodies as early as in September. In <u>C. ovina</u> infective larvae, vacuoles were very rare. Although the infective larvae gradually died, examination of the faeces from these plots revealed specimens of the infective larval stage of <u>Trichostrongylus</u> spp. and <u>Ostertagia</u> spp. up to April of the following year (i.e. after 11 months including overwintering).

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Survival and viability of the infective larvae in the external environment		III	t,o,CH		t,o	h,t,o	h,t o,ch	t,o								CH (ch) Chabertia ovina Small letters vacuolized
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	Experi- mental	PLOC	1	2	ę	4	5	6	7	ω	6	10	11	12	13	H (n) T (t)

160

Table 2

On experimental plots 4 to 6, set out in August to October, infective larvae with vacuoles and reduced motility were found 5 to 7 months after the start of the experiment (i.e. ater overwintering of the infective larvae).

On experimental plots 11 to 13, set out in March to May of the following year, development and survival of the infective larvae were observed in <u>Trichostrongylus</u> spp. and <u>Ostertagia</u> spp.

3. Effect of NILVERM administration on the course of development of the eggs

The results of cultivation up to the infective larval stage of sheep faeces containing eggs of five nematode species before treatment with NILVERM are shown in Table 3. The most rapid development of the infective larvae (on day 4) was shown by <u>S. papillosus</u>. The first infective larvae of <u>Trichostrongylus</u> spp., <u>C. ovina</u>, <u>Ostertagia</u> spp. and <u>H.</u> <u>contortus</u> in the faecal samples were observed on days 5, 8, 10 and 11 of culture, respectively.

Faecal samples from the three NILVERM-treated sheep contained <u>S. papillosus</u>, <u>Trichostrongylus</u> spp. and <u>H.</u> <u>contortus</u> eggs on day 2 after administration. Examination on the 3rd post-treatment day yielded negative results. On day 6, however, sheep No. 2 was found to excrete <u>Trichostrongylus</u> spp. and <u>Trichuris</u> ovis eggs and did so for 28 days.

Sheep No. 3 was found to excrete <u>Trichostrongylus</u> spp., <u>Ostertagia spp.</u>, <u>S. papillosus</u> and <u>C. ovina</u> eggs 70 days after NILVERM administration. Sheep No. 1 showed no evidence of nematodes after a single dose of NILVERM till the end of the 132-day observation period.

Sheep No. 2 received a second 4 ml dose of NILVERM 28 days after the first treatment and began to excrete <u>S.</u> papillosus, <u>Trichostrongylus</u> spp., <u>Ostertagia</u> spp., <u>H.</u> contortus, <u>T. ovis</u> and <u>Capillaria</u> sp. eggs 6 days later. After being treated with NILVERM for a third time (43 days after the first administration and 15 days after the second administration), this animal excreted <u>Trichostrongy-lus</u> spp., <u>Ostertagia</u> spp., <u>H.</u> contortus, <u>C.</u> ovina and

Table 3	Re	sults of from natu	cultiv. Irally-	ation to infected	the infect sheep befo	tive la ore adr	urval sta ninistrat	Results of cultivation to the infective larval stage of the faeces from naturally-infected sheep before administration of NILVERM	eces RM		
Helminth				Cultiva	Cultivation (days)	Ģ					
		1	2	ŝ	4	5	9	7	8	10	11
Strongyloides papillosus	illosus	ı	1	I	+	+	+	+	+	+	+
Trichostrongylus spp	spp.	ı	I	I .	ı	+	+	+	+	+	+
Chabertia ovina		ı	ı	ı	ı	I	ı	ı	+	+	+
Ostertagia spp.		ı	•	ı	l	ı	ı	ı	ı	+	+
Haemonchus contortus	tus	ı	ı	ı	ı	ı	ı	ı	•	I	+
Table 4	fae	Results of cultivation to t faeces from naturally-infected	of cult natura	tivation 11y-infec	Results of cultivation to the infective larval es from naturally-infected sheep after administ Administration of	fective after nistra	e infective larval stage c neep after administration Administration of NILVERM	che infective larval stage of the sheep after administration of NILVERM Administration of NILVERM	LVERM		
Helminth	2nd day	Cultivation L ₃ (days)	1 tion ys)	6th day	Cultivation L ₃ (days)		6th day	2 Cultivation L ₃ (days)	1 2nd day	3 Cult: L ₃ (3 Cultivation L ₃ (days)
S. papillosus	+	5		ı	ı		+	12	'n		ı
Trichostrongylus spp.	+	7		+	8		+	9	+		7
H. contortus	+	13		ı	ı		+	L ₂ till the end	+		7
Ostertagia spp.	ı	I		I	ı		+	12	+		7
Chabertia ovina	ı	1		ı	ı		ı	ı	+		8
Capillaria sp.	I	I		ı	ı		+	l	ı		
Trichuris ovis	ı	I		+	ı		+	1	+		ı

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162

<u>T. ovis</u> eggs from 2 to 27 days after the third treatment. It was only from this time onward (i.e. beginning 71, 43 and 28 days after the first, second and third treatments, respectively) that no eggs were revealed in the faeces of this sheep till the end of the observation period.

Control untreated ewes excreted eggs of nematodes of the genera <u>Strongyloides</u>, <u>Trichostrongylus</u>, <u>Ostertagia</u>, <u>Haemonchus</u>, <u>Trichuris</u> and <u>Chabertia</u> throughout the observation period.

The effect of NILVERM administration on the rate of development of the eggs at 25 °C and 100% relative humidity under laboratory conditions is shown in Table 4. On culture of the faecal samples collected from the three NILVERM-treated sheep 24 h after the first administration infective larvae of <u>S. papillosus</u>, <u>Trichostrongylus</u> spp. and <u>H. contortus</u> were first observed on days 5, 7 and 13, respectively. On culture of the faeces (containing eggs of nematodes of the genera <u>Trichostrongylus</u> and <u>Trichuris</u> collected from sheep No. 2 on day 6 after the first NILVERM administration the infective larval stage was reached only by Trichostrongylus spp. in 8 days.

The faeces from the same animal containing eggs of 6 nematode species (see above) was cultured again on day 6 after the second NILVERM administration. Infective larvae of <u>Trichostrongylus</u> spp. were first observed on day 6 and those of <u>S. papillosus</u> and <u>Ostertagia</u> spp. on day 12. <u>H. contortus</u> larvae. were still in the second stage 13 days after the cultivation was started.

On culture of a positive faecal sample collected from the same animal 24 h after the third NILVERM administration infective larvae of <u>Trichostrongylus</u> spp., <u>Ostertagia</u> spp. and <u>H. contortus</u> were first observed on day 7 and those of <u>C. ovina</u> on day 8.

Discussion

Helminth infections continue to be a problem particularly under intensive husbandry practices because most 3rd stage (infective) larvae are able to survive under outdoor conditions. Investigations into the biology and development of their pre-parasitic stages under the present environmental conditions are therefore essential for organization of optimal preventive measures. Although a number of relevant reports have been published, the results are often contradictory. To be effective, any helminth control program must therefore take into account the actual environmental conditions in which the development of gastrointestinal nematodes takes place.

From our study on the rate of development of the eggs of five species of gastrointestinal nematodes of ruminants during a 13-month period (May to May) it appears that the infective larval stage was reached most rapidly, in 5 days, by <u>S. papillosus</u> and in 11 days by <u>H. contortus</u> and <u>Trichostrongylus</u> spp. According to G old berg and Lucker (1963) the development of infective larvae from <u>H. contortus</u> eggs takes 7 to 21 days at 15 to 16°C, according to Pustovoj (1970) it takes 7 to 8 days at the mean air temperature of 15.1 to 16.7°C. In keeping with a number of other writers we found that development of the eggs to the infective larval stage became longer as the air temperature decreased. Thus Morgan et al. (1951), fo example, found that <u>Ostertagia circumcinta</u> eggs developed to the infective larval stage at 5°C and Shumakovich et al. (1968) reported that helminth eggs developed to the infective larval stage at 10°C in 12 to 31 days.

In our study the development of <u>Ostertagia</u> spp. and <u>Trichostrongylus</u> spp. eggs proceeded till the infective larval stage not only at the mean air temperature of $5.1^{\circ}C$ (taking 40 days but also at the mean temperature of $0.6^{\circ}C$ and at the relative humidity of 75.3° . Under these outdoor conditions <u>H. contortus</u> and <u>Ostertagia</u> spp. infective larvae were first detected on day 65.

Larvae did not develop in the period November to February at which time the mean air temperature fell below -2.0 °C. From our results it appears that under these conditions helminth eggs of the genera <u>Haemonchus</u>, <u>Trichostrongylus</u> and <u>Chabertia</u> are unable to develop and lose their viability at temperatures below zero. Our findings are in keeping with the observations of Persson (1974) and Trač (1975) but differ from those of Bashirov (1979) who reported that eggs of the genera <u>Ostertagia</u> and <u>Trichostrongylus</u> survived at -16.0 to -25.0 °C. From our results it also appears that under our conditions development of the eggs to the infective larval stage is most rapid in May to September, i.e. during the grazing season.

In our study the infective larvae survived under outdoor conditions for as many as 11 months, depending on the period during which the faeces were put on experimental plots. High air temperatures in combination with a low relative humidity in summer led to the destruction and vacuolization of infective larvae in as few as 2 to 4 months. On the other hand, the infective larvae were able to withstand lower temperatures combined with a high relative humidity much better, showing the first signs of vacuolization after 5 to 7 months. Similar observations were reported by W i ll i a m s and B i l k o v i c h (1971) and P e r s s o n (1974). The shortest survival times of infective larvae under our outdoor conditions were recorded for <u>H. contortus</u> (8 to 9 months) followed by <u>C. ovina</u> (10 months). Infective larvae of <u>Trichostrongylus</u> spp. and <u>Ostertagia</u> spp. survived for 11 months.

To combat helminth infections of cattle and sheep, many anthelmintics have been used under a variety of chemo-therapeutic regimens. From a number of studies it appears their effects are often short-lived, in consequence that of which repeated treatment has been frequently recommended (Kotrlá and Koždoň 1981). Up to now the effects of maybe inadequately large and repeated doses of anthelmintics on development of the eggs to the infective larval stage remain undisclosed. Our preliminary results suggest that repeated administration of NILVERM to naturally-infected sheep had no marked effect on the rate of development of the eggs to the infective larval stage and presumably did not impair the viability of infective larvae either.

Our findings are of value to a better understanding of the spread of helminth infections. The ability of the infective larvae to survive and overwinter in the external environment, practically unimpaired development of the eggs to the infective larval stage without loss of infectivity after repeated anthelmintic treatment and particularly rapid development to the infective larval stage during the grazing season are responsible for contamination of the grassland throughout the year, making it the source of helminth infection of young animals. Vliv základních abiotických faktorů na vývoj vajíček a schopnost přežívání invazních larev významných druhů helmintů ovcí ve vnějším prostředí

Vývoj vajíček hlístic rodů Strongyloides, Haemonchus, Trichostrongylus, Ostertagia a Chabertia do vzniku invazních larev byl sledován v průběhu jednoho roku v měsíčních intervalech na experimentálně zakládaných travních plochách o rozměrech 50 x 50 cm. Nejrychleji, 5 - 15 dnů, probíhal vývoj u jednotlivých druhů v měsících květnu až září při průměrné teplotě vzduchu od 15,6 °C - 23,8 °C. Vývoj larev do invazního stadia probíhal po dobu 65 dnů i při průměrné teplotě vzduchu 0,6 °C a relativní vlhkosti vzduchu 76,3 %. V období měsíců listopad až únor jsme žádné invazní larvy v zakládaných vzorcích trusu nenalezli.

Ve vnějším prostředí přežívaly až po dobu 11 měsíců invazní larvy hlístic rodů <u>Haemonchus</u>, <u>Trichostrongylus</u>, <u>Ostertagia</u> a <u>Chabertia</u>. V letním období dochází vlivem vyšší teploty a nižší relativní vlhkosti vzduchu k destrukci larev během 2 - 4 měsíců. Nižší teploty s vyšší relativní vlhkostí vzduchu snášejí invazní larvy lépe. První známky destrukce byly pozorovány po 5 - 7 měsících.

Opakované podávání NILVERMU spontánně invadovaným ovcím výrazně neovlivnilo rychlost vývoje larev do invazního stadia a pravděpodobně nedochází ani k narušení jejich životaschopnosti.

Влияние основных абиотических факторов на развитие яичек и способность выживания инвазионных личинок важных видов гельминтов овец и крупного рогатого скота во внешней среде

Развитие яичек нематод рода <u>Strongyloides</u>, <u>Haemonchus</u>, <u>Trichostrongylus</u>, <u>Ostertagie</u> и <u>Chabertia</u> до возникновения инвазионных личинок исследовали в течение одного года с месячными интервалами на экспериментально созданном травянистом покрове размером 50 x 50 цм. Быстрее всего, <u>пять</u> - пятнадцать суток, протекало развитие отдельных видов с мая по сентябрь при средней температуре воздуха 15,6 °С - 23,8°. Развитие личинок до инвазионной стадии протекало 65 суток даже при средней температуре воздуха 0,6 °С и отностительной влажности воздуха 76,3%. С ноября по февраль никаких инвазионных личинок в образцах кала нами не было обнаружено.

Bo внешней среде выживали до 11 месяцев инвазионные личинки нематод рода Haemonchus, Trichostrongylus, Ostertagia и Chabertia. В летний период под воздействием более высокой температуры И более низкой относительной влажности воздуха происходит в течение 2 - 4 месяцев деструкция личинок. Более высокой относительной низкую температуру с более влажностью инвазионные личинки переносят лучше. Первые признаки деструкции наблюдались СПУСТЯ 5 - 7 месяцев.

дача НИЛЬВЕРМА Повторная спонтанно инвадированным овцам не оказала существенного влияния на личинок до инвазионной скорость развития стадии по всей вероятности, не нарушается их жизне-И, способность.

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