PSEUDORABIES VIRUS GROWTH FACTOR (PRGF) FACILITATES THE GROWTH AND POSTNATAL DEVELOPMENT OF MICE AND RATS

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Abstract


PRGFA, one of two components of pseudorabies virus (PRV) growth factor (PRGF) facilitated the growth and postnatal development of newborn BALBc mice and outbred albino Wistar rats, when administered subcutaneously. The growth proceeded much more rapidly and the initiation of hair growth appeared considerably sooner in PRGFA treated than in control animals. Susceptibility to PRGFA decreased with age, two days old animals proved to be the most susceptible and became non-susceptible at the age of 12 days.

Two monoclonal antibodies (moabs) directed against gII glycoprotein of PRV neutralized the growth stimulating activity of PRGFA. Considering these findings, PRGFA is discussed as a potential novel growth stimulator.

Pseudorabies virus, viral growth factor, newborns, mice, rats, growth stimulation

Pseudorabies virus growth factor (PRGF), possessing dual, transforming and transformed phenotype repressing activity was detected in certain pseudorabies virus (PRV) infected and transformed cells (Golais et al. 1990; Golais et al. 1992). Purification of PRGF by discontinuous recycling chromatography revealed that this factor consists of two components, PRGFₐ and PRGFₐ. Each of these components alone is sufficient for transforming activity, whereas for transformed phenotype repressing activity both components are required. Two monoclonal antibodies (moabs) No. 36 and 68 (Quist et al. 1989) directed against gII glycoprotein of PRV were shown to neutralize both PRGF activities, thus indicating that PRV gene for gII might be involved in PRGF synthesis (Gašperík et al. 1994).

The described effect of PRGF on normal and transformed cells (Golais et al. 1990) might be of particular interest in studies of tumour formation, progression or inhibition. Before testing PRGF in tumour bearing animals, its effect on normal suckling mice and rats has been studied and presented in this paper.

Materials and Methods

Preparation of PRGF. Semipurified PRGF containing both components as well as its two purified components PRGFₐ and PRGFₐ were prepared by procedure developed by Gašperík et al. (1994). All three PRGF samples were further used for inoculation of animals.

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Authors state that all procedures using animals were performed in accordance with European Convention for the Protection of Vertebrate Animals Used for Experimental and Other Scientific Purposes (1986).
Animals. Conventional BALB/cJ/Ham mice, originated from Laboratory Animal Institute Hannover, Germany and conventional Wistar rats were obtained from VELAZ, Prague, Czech Republic. The cages (VELAZ, Prague) of TII type (bottom area 333 cm²) were used for housing of mice and TIII type (bottom 738 cm²) were used for rats in “open systems”. The cages made of polypropylene were transparent for daily light. Sterilized, dried, soft and dust-free wood shavings used for bedding were changed daily. The room temperature was kept at 22 ± 1°C, relative humidity at 55 ± 10%. Natural daylight, and an 8-10-fold air change in the room per hour was provided. The animals were maintained on a standard laboratory diet (ST - 1 feed, VELAZ, Prague) with drinking water available ad libitum.

Inoculation of animals. Phosphate buffered saline (50 μl) at pH 7.2, containing 10⁶ units of PRGF, PRGF_A or PRGF_B (for definition of PRGF unit see Golais et al. 1992) was injected subcutaneously, intraperitoneally or given orally to two-day-old BALBc mice, or subcutaneously to three-day-old outbred albino Wistar rats, observing the guidelines recommended by NIH Animal Care and Use Committee. Six mice or 8 rats from each litter were always randomly chosen for inoculation and kept in one cage. Sham-inoculated animals were given 50 μl intact saline, a group of the control animals was kept intact, i.e. without any inoculation. The body mass and body length of animals were determined daily. The animals repeatedly inoculated with the same amounts of PRGF (administered at 48 hrs intervals) were compared with those inoculated once only. The results were statistically evaluated by Student’s t-test.

Neutralization of PRGF. All three PRGF samples were neutralized with two mabs, No 36 and 68 directed against gII of PRV. Moabs were obtained from Veterinary Serum Laboratories Copenhagen, Denmark (Qvist et al. 1989) and were previously shown to neutralize the PRGF activity in vitro (Gašperík et al. 1994). This neutralization method was described previously (Golais et al. 1992).

Results

The effect of PRGF and its separated components on newborn mice

PRGF administered subcutaneously was shown to enhance the growth and to facilitate the postembryonal development of mice. PRGF-treated animals grew much more rapidly as compared to the sham-treated ones whose growth did not substantially differ from intact controls. Moreover, PRGF-treated mice became pilous as soon as 3-4 days after inoculation while hair growth in the sham-inoculated animals initiated 4-5 days later. However, there were some differences in growth enhancement between the non-resolved PRGF and its two separated components. PRGF_A proved to be the most effective growth stimulator, whereas

Fig. 1. The effect of PRGF on suckling BALBc mice 24 hrs after subcutaneous administration. a) mouse treated with PRGF_A; b) mouse treated with non-resolved PRGF; c) sham-inoculated mouse.
non-resolved PRGF and PRGF_B were less efficient (Fig. 2). Mice inoculated with PRGF and PRGF_A are shown in Fig. 1.

Following intraperitoneal administration no significant growth differences were obtained with non-resolved PRGF, PRGF_B and sham control, only PRGF_A caused a limited short-term effect (Fig. 3). Both resolved and non-resolved PRGF samples elicited no effect in mice when administered orally (data not shown).

The growth stimulating effect of PRGF following subcutaneous inoculation was observed only during a 17-19-day long time period with maximum differences from control animals between days 2 and 12 after inoculation. The body mass and length of all tested animals became approximately equal at about day 18 after initial PRGF administration (Figs 2 and 3ab).

![Fig. 2. The effect of PRGF and its two separated components (PRGF_A, PRGF_B) on growth of newborn mice. Subcutaneous administration. Hair growth initiation is shown by arrows. The means of body mass and standard deviations were calculated from 6 repeated experiments including more than 35 animals in each time point. The t-test was used for statistical analysis. The means of the weight of PRGF_A treated and control animals were different at the P<0.05 level at 4 - 18 days. The body mass means of the PRGF-treated and control animals were different at the P<0.05 level at 4 - 12 days. ○○ sham-inoculated controls, ●● PRGF_B, ▼▼ non-resolved PRGF, ▼▼ PRGF_A.](image1)

![Fig. 3a (for text see p. 252)](image2)
Fig. 3b Comparison of subcutaneous and intraperitoneal administration of PRGF\textsubscript{A} on the size (Fig.3a) and body mass of mice. The body mass means and standard deviations were calculated from 6 repeated experiments including more than 35 animals in each time point. The t-test was used for statistical analysis. At subcutaneous administration of PRGF\textsubscript{A}, the means of the body mass and size of treated and control animals were different at the \( P < 0.05 \) level at 4 - 16 days. ○–○ sham-inoculated controls, ●–● intraperitoneal administration, ▽–▽ subcutaneous administration.

Repeated inoculation of mice at 48 hrs interval was a prerequisite for observed growth enhancement. Otherwise, when after the first inoculation no further challenges took place, the growth enhancement stopped within the next 4-5 days.

All PRGF samples lost their ability to enhance the growth of mice when treated either with moab 36 or 68, or both. Fig. 4 shows the effect of moab 36 on PRGF\textsubscript{A} activity (other data are not shown).

Fig. 4. The effect of moab 36 on growth stimulating activity of PRGF\textsubscript{A}. Subcutaneous administration of mice. The body mass means and standard deviations were calculated from 6 repeated experiments including more than 35 animals in each time point. The t-test was used for statistical analysis. At subcutaneous administration of PRGF\textsubscript{A}, the body mass means and size of treated and control animals were different at the \( P < 0.05 \) level at 4 - 20 days. ○–○ sham-inoculated controls, ▽–▽ PRGF\textsubscript{A}, ●–● moab treated PRGF\textsubscript{A}.
Age-dependent susceptibility to PRGF<sub>A</sub>

In these experiments, mice of different ages were treated with the same amount \((10^6 \text{ units})\) of PRGF<sub>A</sub> and weighed after 24 hrs. The susceptibility of animals markedly decreased with age. Mice aged 2 days showed highest susceptibility, their body mass was about 150% higher as compared to 100% in sham-inoculated controls. Animals aged 4 days reached about 75%, and those aged 6 days about 50% body mass increase. Mice at 8 days of age showed about 45% body mass increase, and at the age of 12 days all animals became non-susceptible to PRGF<sub>A</sub> (see Fig. 5).

![Fig. 5. Susceptibility of mice of different age to PRGF<sub>A</sub>.](image)

Fig. 5. Susceptibility of mice of different age to PRGF<sub>A</sub>. The animals were inoculated by PRGF<sub>A</sub> at different ages and the next day the body mass of treated animals were measured and compared to their respective controls. The body mass of the controls at each age is 100%. Repeated experiments included about 30 animals from each age category. ●● PRGF<sub>A</sub> treated animals, ○○ sham-inoculated controls.

![Fig. 6. Growth stimulating effect of PRGF<sub>A</sub> on 3-day-old rats and the effect of moab 68.](image)

Fig. 6. Growth stimulating effect of PRGF<sub>A</sub> on 3-day-old rats and the effect of moab 68. Subcutaneous administration. The means body mass means and standard deviations were calculated from 6 repeated experiments including more than 45 animals in each time point. The t-test was used for statistical analysis. At subcutaneous administration of PRGF<sub>A</sub>, the means of the body mass and size of treated and control animals were different at the \(P < 0.05\) level at 5 - 19 days.

●● PRGF<sub>A</sub> treated with moab.
The effect of long-term administration of PRGF₁₆

Sixty mice aged 2 days were inoculated subcutaneously with PRGF₁₆ and kept for 4 months along with 18 sham-treated and 18 non-treated animals (male and female animals were kept separately). In the first 4 weeks, \(10^6\) units were inoculated into each animal at 48 hrs intervals, later on \(2 \times 10^6\) units were administered twice weekly. During the period under study, PRGF-treated animals did not differ in any respect (weight, size, behaviour) from those sham-inoculated or non-inoculated (besides the above mentioned 17 days time span). No tumors or visible pathological findings as revealed by autopsy were found in animals kept in the long-term experiment under PRGF treatment.

The effect of PRGF on suckling rats

PRGF₁₆ introduced subcutaneously into 3-day-old rats induced a similar growth stimulating effect as was observed in mice. It was similarly neutralized by two moabs against gII of PRV. Fig. 6. shows the effect of moab 68 on PRGF₁₆.

Discussion

PRGF, a novel viral growth factor detected in certain PRV infected and transformed cells was shown to have two effects on cultured cells in vitro. Non-transformed cells cultivated in the presence of PRV acquired the phenotype pertain to transformed cells ("criss-cross" pattern of growth, anchorage independence in soft agar), whereas the phenotype of transformed cells became converted towards the normal one (Golaïs et al. 1990).

Purification of PRGF by discontinuous recycling chromatography revealed that this factor consists of two components, PRGF₁₆ and PRGF₁₇. Each of these two components is sufficient for manifestation of transforming activity, whereas for transformed phenotype repressing activity, either non-resolved PRGF, or both components acting simultaneously are required (Gašperík et al. 1994).

The aim of our further studies is the testing of PRGF and its two components in vivo. Especially the transformed phenotype repressing activity of PRV observed in vitro might be under certain circumstances utilized in prevention of tumour formation or progression. In the first step of this research program, PRGF and its two components have been tested in suckling mice and rats in attempts to demonstrate, or rule out their oncogenicity in vivo. It has been surprisingly shown, that PRGF, especially the PRGF₁₆ component facilitates the growth and development of suckling mice and rats. Long-term keeping of mice under PRGF₁₆ treatment have not resulted in appearance of tumours. We are aware of the fact that based on these results there is no reason to believe, that PRGF possess absolutely no oncogenic activity in vivo. More profound studies are required for such conclusion. Of special interest is the interaction of PRGF with endocrine and immune systems. Nevertheless, PRGF being shown to enhance the growth and facilitate the development of newborn mice and rats appears to be very tempting for further studies, as it might find some applications in animal husbandry. As PRGF is at present only one known herpes virus-related growth factor, characterized in our laboratory, the findings presented in this paper, as well as the results reported in Golaïs et al. (1990), Golaïs et al. (1992) and Gašperík et al. (1994) could not be related to any other published studies.

Rastový faktor vírusu pseudorabies (PGRF) urýchluje rast a postembryonálny vývoj myší a potkanov

PRGF₁₆, jedna z dvoch zložiek PGRF, po subkutánnej aplikácii urýchluje rast a postnatálny vývoj novorozených myší a potkanov outbrednej línie Wistar. V skupinách
zvierat, ktorým sme aplikovali PGRF<sub>A</sub>, nastal urýchlený rast a skôršie osrstenie ako v skupinách kontrolných. Účinok PGRF<sub>A</sub> závisel od veku zvierat: najúčinnejší bol u mláďat dvojdňových, ale u mláďat starších ako 12 dní bol neúčinný. Dve monoklonové protitáky proti GII glykoproteínu výrdu pseudorabies neutralizujú rast stimulujujúcu aktivitu PRGF<sub>A</sub>. V práci sa diskutuje použitie PGRF<sub>A</sub> ako nového potenciálneho stimulátora rastu.

References


