FERMENTATION PATTERNS IN RABBIT CAECAL CULTURES SUPPLIED WITH PLANT POLYSACCHARIDES AND LACTATE

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Abstract


Fermentation patterns were investigated in anaerobic cultures of the rabbit caecal contents supplied with starch, hemicellulose, pectin, inulin and sodium lactate. Pectin and inulin were fermented more rapidly than starch and hemicellulose. Fermentation of pectin produced the highest amount of fermentation gas. The highest molar proportion of acetate was found in volatile fatty acids (VFA) of cultures supplied with pectin or without substrate. The highest molar proportions of propionate, butyrate and caproate were found in cultures supplied with inulin, sodium lactate and starch, respectively. Acetate and butyrate accounted for 68 - 85 % of metabolite carbon. Low proportions of propionate and valerate were specific for all substrates. The production of methane was roughly proportional to the VFA production (one mol of methane per 5.9 - 7.3 mols of VFA). Hydrogen recovery values varied from 50.5 to 61.0 %, suggesting the presence of H2-dependent acetogenesis.

Rabbit, caecum, fermentation, VFA production, in vitro cultures

The rabbit caecum is a fermentative chamber, where the microorganisms of the gut are able to metabolically alter the unabsorbed digesta from the stomach and small intestine, making it available to the host. The rabbit caecum is colonized by an abundant bacterial flora, ca 1010 cultivable cells per 1 g (Forsythe and Parker 1985). The principal substrates for caecal microorganisms are polysaccharides (hemicellulose, pectin, starch) and protein. Polysaccharides are converted to volatile fatty acids (VFA), methane, carbon dioxide and compounds incorporated into bacterial cells. The VFA are absorbed, supplying additional energy for the host.

Broiler rabbits fed high concentrate diets are highly susceptible to the development of digestive disturbances. Weanling rabbits are particularly sensitive. High mortality of young rabbits has severe economic impact on commercial rabbit production. The etiology of digestive disturbances is multifactorial. It is known, however, that apart from specific pathogenic agents, the proliferation in the caecum of bacteria that produce potent toxins is responsible for the major part of losses (Cheeke 1989). Cheeke and Patton (1980) proposed that enterotoxemia was caused by carbohydrate overload in the hindgut, providing a substrate allowing the proliferation of pathogens. On low fibre diets, hypomobility occurs, resulting in prolonged retention time of digesta in the caecum, changes in caecal pH, and ultimately causing changes in the caecal microbial population. On the other hand, it has been shown that caecal VFA and low pH inhibit pathogen’s proliferation in the caecum (Próhászka 1980). It follows from these facts that the caecal digestion is one of the factors limiting rabbit performance. The aim of this study was to obtain more information on rabbit caecal fermentation pattern. We compared production of microbial metabolites (VFA, gas, methane), in in vitro cultures of the rabbit caecal contents supplied with starch, hemicellulose, pectin, inulin and lactate.
Materials and Methods

Rabbits were New Zealand × Californian hybrids, fed ad libitum a commercial granulated feed, containing (%): barley - 18.5, oat - 10.0, wheat bran - 20.0, dehydrated lucerne - 25.0, sunflower meal - 21.0, soya-bean meal - 2.0, vitamin-mineral supplement - 3.5. The feed contained no coccidiostat. Twenty rabbits were killed at the age of 80 days, ca 4 h after the morning feeding. Their caeca were emptied by gentle squeezing and pooled caecal contents were used for inoculation of in vitro cultures.

Eight-hour incubations were carried out, on a water bath at 39°C, in 0.51 bottles, hermetically closed with rubber stoppers. The caecal contents (50 ml) were diluted with 100 ml of phosphate-bicarbonate buffer (Mc Dougall, 1949), containing isonitrogenous amounts (total 0.5 g N/l) of urea and yeast extract from Saccharomyces cerevisiae. Hemicellulose and maize starch were added at 3 g, pectin, inulin, and sodium lactate at 2 g per incubation. Hemicellulose was isolated from wheat straw (Marounék et al. 1988), the other substrates were purchased. As a control, the caecal contents were incubated under identical conditions without addition of the substrate. Each experimental or control arrangement was incubated in four replicates. Cultures were agitated manually every 30 min. To provide anaerobiosis, addition of sodium sulphide to the incubation fluid (0.5 g/l) and CO₂ atmosphere were used. The pH fell from ca 7.1-7.2 to about 5.5-6.3 after incubation.

Total VFA were estimated by titration, after steam distillation. Their molar composition was determined by gas-liquid chromatography, at 140°C, using a 2.4 m column of Supelcoport with 5% FFAP (Supelco). Samples of the headspace gas were taken at the end of the incubation and analysed using the same gas chromatograph with FID detector. At the same time, the manometric pressure in incubation vessels was measured. Methane production was calculated as the product of methane concentration and total gas production. The gas absorption chromatography was employed for the H₂ determination on the Carbosieve S column (Supelco) and a TCD detector. Metabolic hydrogen recovery (R) was computed according to Demeyer and Van Nevel (1975). The following equations were used:

\[ R = \frac{2H_{\text{accepted}}}{2H_{\text{released}}} \]

\[ 2H_{\text{accepted}} = 4M + 2P + 2B + 4V + 4C \]

\[ 2H_{\text{released}} = 2A + P + 4B + 3V + 6C, \]

where M, A, P, B, V and C are molar productions of methane, acetate, propionate, butyrate, valerate and caproate, respectively. The results summarized in tables are related to a difference between the beginning and the end of the incubation. The net amounts produced were obtained by correction of amounts determined for amounds present before incubation. The statisticial treatment of the data was performed by one factorial analysis of variance.

Results and Discussion

Table 1 shows that pectin and inulin were fermented more rapidly in in vitro incubations of the rabbit caecal contents than other substrates tested. Production of VFA from starch and hemicellulose was markedly lower, if we take into account the fact that these substrates were supplied in higher amounts at the beginning of the incubation. Approximately one half of the VFA production originated from the fermentation of carbon sources present in inocula. In all cultures, the propionate percentages were higher than the average proportion of propionate in the caecal contents used for inoculation. The average dry matter (%), pH and VFA concentration (mmol/l) in the rabbit caecal contents were 24.1, 6.57 and 62.5, resp. Acetate, propionate, butyrate, valerate, caproate represented on average 71.5, 7.9, 17.2, 1.9 and 1.5 mol.% of total VFA, respectively. The highest molar proportion of acetate was observed in cultures supplied with pectin or without substrate. The addition of any fermentable substrate to the caecal cultures changed the fermentation stoichiometry towards less acetate and more butyrate and caproate (in comparison with substrate-free controls). The highest molar percentages of propionate, butyrate and caproate were found in cultures supplied with inulin, sodium lactate and starch, respectively. Lactate is produced by some strains of caecal bacteria. It should be mentioned that in the rumen, acetate and propionate, not butyrate, are main products of the lactate fermentation (Nakamura and Takahashi 1971). Fermentation of pectin produced the highest volume of the fermentation gas, probably because of CO₂ formed in uronate decarboxylation. The production of methane was roughly proportional to the VFA production. Between 5.9 and 7.3 mols of VFA per mol
Table 1
Production of VFA, fermentation gas and methane in *in vitro* incubations of the rabbit caecal contents supplied with plant polysaccharides and lactate

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Substrate</th>
<th>Starch</th>
<th>Hemicellulose</th>
<th>Pectin</th>
<th>Inulin</th>
<th>Na-lactate</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>(0.56)</td>
<td>(0.58)</td>
<td>(1.48)</td>
<td>(0.75)</td>
<td>(0.80)</td>
</tr>
<tr>
<td>Acetate (%)</td>
<td></td>
<td>66.0</td>
<td>69.4</td>
<td>71.2</td>
<td>64.9</td>
<td>60.5</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(5.5)</td>
<td>(1.2)</td>
<td>(2.1)</td>
<td>(3.1)</td>
<td>(6.7)</td>
</tr>
<tr>
<td>Propionate (%)</td>
<td></td>
<td>9.5</td>
<td>11.1</td>
<td>8.3</td>
<td>13.0</td>
<td>12.2</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(0.6)</td>
<td>(0.9)</td>
<td>(0.4)</td>
<td>(1.0)</td>
<td>(0.3)</td>
</tr>
<tr>
<td>Butyrate (%)</td>
<td></td>
<td>17.2</td>
<td>17.3</td>
<td>17.4</td>
<td>18.7</td>
<td>23.0</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(2.8)</td>
<td>(1.0)</td>
<td>(0.7)</td>
<td>(1.8)</td>
<td>(5.2)</td>
</tr>
<tr>
<td>Valerate (%)</td>
<td></td>
<td>2.5</td>
<td>1.4</td>
<td>1.5</td>
<td>1.7</td>
<td>2.3</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(0.9)</td>
<td>(0.1)</td>
<td>(0.1)</td>
<td>(0.6)</td>
<td>(0.7)</td>
</tr>
<tr>
<td>Caproate (%)</td>
<td></td>
<td>4.8</td>
<td>0.8</td>
<td>1.6</td>
<td>1.7</td>
<td>2.0</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(2.1)</td>
<td>(0.1)</td>
<td>(0.3)</td>
<td>(0.5)</td>
<td>(0.6)</td>
</tr>
<tr>
<td>Total gas (ml/flask)</td>
<td></td>
<td>373</td>
<td>365</td>
<td>548</td>
<td>447</td>
<td>188</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(38)</td>
<td>(36)</td>
<td>(10)</td>
<td>(25)</td>
<td>(30)</td>
</tr>
<tr>
<td>Methane (mmol/flask)</td>
<td></td>
<td>1.92</td>
<td>2.00</td>
<td>2.02</td>
<td>2.11</td>
<td>1.45</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(0.11)</td>
<td>(0.12)</td>
<td>(0.17)</td>
<td>(0.09)</td>
<td>(0.08)</td>
</tr>
<tr>
<td>VFA/methane (mol/mol)</td>
<td></td>
<td>5.90</td>
<td>6.63</td>
<td>7.34</td>
<td>7.05</td>
<td>6.88</td>
</tr>
<tr>
<td>2H-recovery (%)</td>
<td></td>
<td>61.0</td>
<td>55.2</td>
<td>50.5</td>
<td>57.4</td>
<td>59.7</td>
</tr>
</tbody>
</table>

Means of four incubations (8 h/39 °C). Standard deviations are given in parentheses beneath the mean values

1\(3g/150\) ml

2\(2g/150\) ml

3Without substrate

*Values within the same row differ if they do not share a common superscript (P < 0.05)

Means of four incubations (8 h/39 °C). Standard deviations are given in parentheses beneath the mean values

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2\(2g/150\) ml

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C per incubation. Acetate and butyrate accounted for 67.6-84.8 % of metabolite carbon. Other metabolites were quantitatively less important. The predominance of acetate and butyrate and low proportion of propionate were specific for all substrates.

It has been shown in rabbits that caecal VFA levels depend on the age of animals (Piattoni et al. 1995), on the time after feeding (Gidenne and Bellier 1992) and on dietary composition. Hoower and Heitmann (1972) found that an increased dietary fibre level did cause an increase in caecal VFA concentration, which was mainly attributable to butyric acid. The caecal butyrate production was greater than that of propionate, which represents a divergence from the fermentation ratios normally found in the rumen of ruminants. Gidenne (1992) related the VFA level and acetate proportion to the cell wall contents of the feed. It is shown in this study that four plant polysaccharides and lactate are fermented at different rates in in vitro incubations of the rabbit caecal contents and differ in proportions of fermentation end-products. Substrate-induced fermentation shifts, however, seem to be less pronounced than those in cultures of mixed rumen microorganisms (Lee and Hespell 1983). This difference probably reflects more uniform composition of rabbit caecal microflora in comparison with that of the rumen.

**Fermentace rostlinných polysacharidů a laktátu v kulturách obsahu slepého střeva králíků**

Fermentace v slepém střevu má vztah k užitkovosti králíků i jejich zdravotnímu stavu. Zjišťovali jsme parametry fermentace při anaerobních inkubacích obsahu slepého střeva se škrobem, hemicelulózou, pektinem, inulínem a Na-laktátem. Pektin a inulin byly fermentované střevním obsahem snáze než škrob a hemicelulóza. Při fermentaci pektinu se uvolňovalo největší množství fermentačního plynu. Nejvyšší molární zastoupení acetátu bylo zjištěno mezi těkavými mastnými kyselinami (TMK) v kulturách střevního obsahu s pektinem či bez substrátu. Nejvyšší molární zastoupení propiónátu, butyrátu a kaproátu bylo nalezeno v inkubacích s inulinem, laktátem a škrobem. Většina (68–85 %) uhlíku vzniklých metabolitů byla absažena v acetátu a butyrátu. Nízká produkce propiónátu a valerátu byla zjištěna u všech substrátů. Produkce metanu byla zhruba úměrná produkci TMK (mol metanu na 5,9–7,3 molu TMK). Hodnoty recovery vodíku se pohybovaly mezi 50,5 a 61,0 %, což naznačuje přítomnost acetogeneze z CO₂ a H₂.

**References**


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