TOTAL ANTIOXIDANT CAPACITY (TAC) VALUES AND THEIR CORRELATION WITH INDIVIDUAL ANTIOXIDANTS IN SERUM OF HEALTHY BEAGLES

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Abstract

The paper aims to establish a range for serum Total Antioxidant Capacity (TAC) and to determine the correlation between TAC and some individual antioxidants (vitamin A and E, lipid standardised vitamin E [Vit E/LS], β-carotene, total bilirubin and albumin) in a uniform population of beagle dogs. Emphasis was directed on their general health status to establish a basis for future investigation of the role of TAC in diseases of dogs as species most frequently studied in veterinary medicine. The animal body possesses a variety of protective antioxidant substances that act as a harmoniously and finely tuned mechanism to neutralise harmful oxidants. TAC measurements provide a tool for establishing links between antioxidant capacity and the risk of disease as well as for monitoring of antioxidant therapy. Serum samples of 19 healthy beagles were assayed for TAC on LP 700 photometer (Dr. Lange, Germany) with a commercially available TAS kit (“Total Antioxidant Status” – TAS; Randox, Crumlin, UK). TAS kit measures the capacity of all of the antioxidants present in serum or plasma sample. The assay is based on the suppression of the absorbance of the radical cation of 2,2’-azinobis (3-ethylbenzothiazoline-6-sulfonate) (ABTS•+) by antioxidants. Assay results are expressed as mmol/l of Trolox (6-hydroxy-2, 5, 7, 4-tetramethylchroman-2-carboxylic acid – a water-soluble analogue of α-tocopherol) equivalents.

The range for TAC expressed as mean ± SD resulted in 1.08 ± 0.08 mmol/l. TAC correlated positively with albumin (r = 0.18), vitamin E (r=0.14) and Vit E (LS) (r = 0.20), and negatively with total bilirubin (r = -0.30), vitamin A (r = -0.15) and β-carotene (r = -0.13) although no significant correlation has been found.

Antioxidant capacity, antioxidants, free radicals, dog

A wide range of substances, known as Reactive Oxygen Species (ROS), consisting of free radicals such as O2•-, OH•, and other non-radical oxygen derivatives such as hydrogen peroxide (H2O2), hypochlorous acid (HOCl) and singlet Oxygen (1O2) are constantly generated in vivo as an integral part of metabolism, as part of a controlled inflammatory reaction and by exposure to environmental factors. A free radical can be defined as any species capable of independent existence that contains one or more unpaired electrons (Halliwell 1996). Non-radical oxygen derivatives are substances capable of radical formation in intra- and extra-cellular environments (Cao and Prior 1998; Chapple 1997; Halliwell 1997). The pathological increase of ROS generation has already been recognised in over one hundred human and animal diseases including cancer, cardiovascular disease, diabetes mellitus, male infertility, renal disease and dialysis, cataracts, neurological, liver, periodontal, lung and inflammatory diseases (Halliwell 1996; Lantos et al. 1997; Moore et al. 1994; Pinzani et al. 1998). ROS circulate freely in the body with access to all organs and tissues. They cause tissue damage by a variety of different mechanisms, which...
include DNA damage, lipid peroxidation (through activation of cyclooxygenases and lipoxygenases), protein damage, oxidation of important enzymes, e.g. anti-proteases such as α1-antitrypsin, and stimulation of pro-inflammatory cytokine release by monocytes and macrophages (Chapple 1997). It is therefore not surprising that all oxygen consuming organisms have developed complex antioxidant systems to counteract ROS and to reduce their damage (Cao et al. 1995; Halliwell 1996). Antioxidants may be regarded as those substances which, present at low concentrations, compared to those of an oxidisable substrate, will significantly delay or inhibit oxidation of that substrate (Halliwell 1995). There are different classifications of antioxidants. According to their mode of action Chapple (1997) differentiates them into three main groups:

- “Preventative antioxidants” which prevent the formation of new ROS, e.g. caeruloplasmin, metallothioniene, albumin, myoglobin, ferritin, transferrin.
- “Scavenging antioxidants” remove ROS once formed, thus preventing radical chain reactions. These include reduced glutathione (GSH), vitamine E (α-tocopherol), vitamin C (ascorbic acid), β-carotene, uric acid and bilirubin.
- “Enzyme antioxidants” that function by catalysing the oxidation of other molecules. This group includes superoxide dismutase, glutathione reductase, glutathione peroxidase, catalas and metalloenzymes.

The serum contains many different antioxidants that may be important for general health maintenance. These include ascorbic acid, α-tocopherol, β-carotene, uric acid, bilirubin, and albumin. In addition, trace amounts of antioxidant enzymes such as glutathione peroxidase and superoxide dismutase are found in serum to a lesser extent (Cao et al. 1993; Chapple 1997; Chapple et al. 1997). In diabetes, markers of oxidative stress have been shown to be elevated whereas vitamin C levels are reduced. Some studies have shown that treatment with antioxidants, particularly vitamin C, can reduce both oxidative stress and protein glycation and may help to reduce the risk of developing diabetic complications (Eriksson et al. 1995; Young et al. 1995). Recently Barros et al. (1999) reported low plasma vitamin C levels in cataractous dogs compared with healthy dogs. Decreased plasma levels of vitamin C may indicate a decrease in the antioxidant capacity of aqueous humour. Heliovarra et al. (1994) found low levels of vitamin E, β-carotene and selenium to be associated with increased risk of rheumatoid arthritis. The number of different antioxidant components in serum and tissues makes it relatively difficult to measure each antioxidant component separately. In addition, since there is a co-operation between various antioxidants, looking at one in isolation from rest may not accurately reflect their combined action. Therefore, the measurement of the total serum antioxidant capacity seems to represent a suitable biochemical parameter for evaluating the overall antioxidant status resulting from antioxidant intake or production and their consumption by the increasing levels of oxidative stress (Cao et al. 1993; Cao and Prior 1998; Chapple 1997; Pinzani et al. 1998; Whitehead et al. 1992). Several methods have been developed to evaluate the total antioxidant capacity of serum of plasma (Cao et al. 1993; Cao et al. 1995; Miler et al. 1993; Rice-Evans and Miller 1994; Whitehead et al. 1992). These methods are all essentially inhibition methods. A free radical species is generated, there is an end point by which the presence of the radical is detected, and the antioxidant activity of the added sample inhibits the end point by scavenging the free radical. Methods vary greatly as to the radical that is generated, reproducibility of the generation process, and the end point that is used (Rice-Evans and Miller 1994).

The aim of our study was to establish the range for serum Total Antioxidant Capacity (TAC) and to determine the correlation between TAC and some individual antioxidants (vitamin A and E, lipid standardised vitamin E [Vit E(LS)], β-carotene, total bilirubin and albumin) in healthy beagles.
Materials and Methods

Animals
19 beagles, 15 females and 4 males, ranging from 1 to 3 years, were considered healthy on the basis of history, results of physical examination, haematological parameters, i.e. complete blood count (CBC), white cell differential count (WCDC) and serum biochemical profile.

Samples
Venous blood samples were collected from fasted dogs into plain and EDTA – containing tubes. Samples in plain tubes stood for 30 min at 4 °C to clot, prior to centrifugation (3000 rpm for 10 min) and separation of serum. Serum samples were stored at -70 °C and assayed within 2 weeks in duplicate for TAC and various biochemical parameters indices (urea, creatinine, sodium, potassium, total bilirubin, total protein, albumin, alanin-aminotransferase [ALT], cholesterol, triglycerides, \( \beta \)-carotene and vitamins A and E). EDTA blood samples for CBC and WCDC determination were stored at room temperature and analysed from 1 to 5 h after sampling.

TAC measurements
Serum samples of 19 healthy beagle dogs were assayed for TAC on LP 700 photometer (Dr. Lange, Germany) with a commercially available TAS kit (“Total Antioxidant Status” – TAS; Randox, Crumlin, UK), following the instructions of the kit. The assay is based on the reduction of free radicals (ABTS\(^{•+}\) – 2,2'-azinobis (3-ethylbenzothiazoline-6-sulfonate) measured as a decrease of absorbance at 600 nm at 3 min by antioxidants. The ABTS\(^{•+}\) radical cation is formed by the interaction of ABTS with ferrylmyoglobin radical species, generated by the activation of metmyoglobin with hydrogen peroxide. The suppression of the absorbance of the ABTS\(^{•+}\) radical cation by serum antioxidants was compared with that from a Trolox (6-hydroxy-2, 5, 7, -tetramethylchroman-2-carboxylic acid) which is included as part of the TAS kit. The results are expressed as mmol/l of Trolox equivalents. In addition, control serum (Randox, Crumlin, UK) with TAC value of 1.0 to 1.36 mmol/l was assayed in each batch of samples for the estimation of analytical imprecision (between-batch coefficient of variation).

Determination of haematological parameters
CBC and WCDC were determined by an automated laser haematology analyser Bayer-Technicon H\(^*\)1 (Bayer – Technicon, Tarrytown, New York) with species specific software (H\(^*\)1 Multi-Species V30 Software, Tarrytown, New York). CBC includes white blood cells (WBC), red blood cells (RBC), haemoglobin concentration (HGB), haematocrit (HCT), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH), mean corpuscular haemoglobin concentration (MCHC) and platelets (PLT). WCDC represent a six-part differential neutrophils (NEUT), lymphocytes (LYMPH), monocytes (MONO), eosinophils (EOS), basophils (BASO) and large unstained cells (LUC), both as a percentage (data not shown) and absolute count. The LUC category consists of a heterogeneous population of all large cells that fail to exhibit any peroxidase activity (atypical lymphocytes, immature granulocytes and blasts).

Determination of biochemical parameters
Urea, creatinine, sodium (Na), potassium (K), total bilirubin, total protein, albumin, uric acid and alanin-aminotransferase (ALT) were determined by automated chemistry analyser Ektachem 250 (Kodak, Rochester, New York). Cholesterol and triglycerides were determined by Abbott reagents using automated chemistry analyser ALCYONTM 300 (Abbott, Illinois, USA). Serum \( \beta \)-carotene concentrations were determined by spectrophotometric method at 450 nm. The preparation of the serum included saponification of the \( \beta \)-carotene with alcohol solution of potassium hydroxide (KOH) and extraction with n-heptane. Serum vitamin A and E concentrations were determined by fluorometric methods. The preparation of the serum included saponification of the vitamins A and E with alcohol solution of potassium hydroxide (KOH) and extraction with petrol ether. Lipid standardised vitamin E (Vit E[LS]) values were calculated as the ratios of vitamin E to the sum of cholesterol and triglycerides. They are expressed as \( \mu \)mol vitamin E/mmol total cholesterol plus triglycerides (Benzie et al. 1998; Jost et al. 1999).

Statistical evaluation
Statistical analyses were done using Statistica for Windows (STATSOFT Incorporation, 1993). Means and standard deviations were calculated for all haematological and biochemical parameters. Correlation coefficients (r) were calculated for TAC and some individual antioxidants. Correlations were considered significant at the level \( P < 0.05 \).

Results
The serum TAC values obtained for 19 beagle dogs varied between 0.93 and 1.27 mmol/l, with a mean value 1.08 ± 0.08 mmol/l, median value 1.10 mmol/l and valued for lower quartile 1.02 mmol/l and upper quartile 1.12 mmol/l. The analytical imprecision was
estimated by determination of between–batch coefficient of variation calculated on the basis of 20 analyses of control serum. The results are summarised in Table 1.

Table 1
Between-batch precision of TAC assay

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control serum</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number</td>
<td>20</td>
</tr>
<tr>
<td>Mean (mmol/l)</td>
<td>1.17</td>
</tr>
<tr>
<td>SD</td>
<td>0.03</td>
</tr>
<tr>
<td>CV (%)</td>
<td>2.8</td>
</tr>
</tbody>
</table>

Haematological and biochemical values of beagle dogs are presented in Tables 2, 3 and 4. Biochemical and haematological values were consistent with other published normal values (Bush 1998; Tvedten 1999; Baker et al. 1986). There are no data available for Vit (E (LS) in dogs in literature. The uric acid values were less than 12 µmol/l in all 19 beagle dogs.

Table 2
CBC values expressed as mean ± SD

<table>
<thead>
<tr>
<th>WBC (10^9/l)</th>
<th>RBC (10^12/l)</th>
<th>HGB (g/l)</th>
<th>HCT (l/l)</th>
<th>MCV (fl)</th>
<th>MCHC (g/l)</th>
<th>PLT (10^9/l)</th>
</tr>
</thead>
<tbody>
<tr>
<td>9.84 ± 2.53</td>
<td>7.43 ± 0.46</td>
<td>173 ± 9</td>
<td>0.54 ± 0.028</td>
<td>72.5 ± 1.6</td>
<td>321 ± 5</td>
<td>357 ± 61</td>
</tr>
</tbody>
</table>

No significant correlation (P > 0.05) was found between TAC and individual antioxidants such as albumin, total bilirubin, vitamin A, β-carotene, vitamin E and Vit E(LS).

Table 3
WCDC values expressed as mean ± SD

<table>
<thead>
<tr>
<th>NEUT (10^9/L)</th>
<th>LYMPH (10^9/L)</th>
<th>MONO (10^9/L)</th>
<th>EOS (10^9/L)</th>
<th>BASO (10^9/L)</th>
<th>LUC (10^9/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>5.95 ± 1.74</td>
<td>3.20 ± 1.01</td>
<td>0.30 ± 0.13</td>
<td>0.29 ± 0.17</td>
<td>0.024 ± 0.0016</td>
<td>0.056 ± 0.035</td>
</tr>
</tbody>
</table>

Table 4
Biochemical values expressed as mean ± SD

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Value</th>
<th>Parameter</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Urea (mmol/l)</td>
<td>3.0 ± 1.0</td>
<td>Albumin (g/l)</td>
<td>33.9 ± 3.2</td>
</tr>
<tr>
<td>Potassium (mmol/l)</td>
<td>4.1 ± 0.2</td>
<td>Cholesterol (mmol/l)</td>
<td>4.8 ± 0.88</td>
</tr>
<tr>
<td>Sodium (mmol/l)</td>
<td>150 ± 3</td>
<td>Triglycerides (mmol/l)</td>
<td>0.3 ± 0.08</td>
</tr>
<tr>
<td>Creatinine (µmmol/l)</td>
<td>57.3 ± 10.8</td>
<td>Vitamin A (µmol/l)</td>
<td>11.2 ± 4.2</td>
</tr>
<tr>
<td>ALT (µkat/l)</td>
<td>0.66 ± 0.33</td>
<td>β-carotene (µmol/l)</td>
<td>0.2 ± 0.1</td>
</tr>
<tr>
<td>Total bilirubin (µmol/l)</td>
<td>7.0 ± 0.09</td>
<td>Vitamin E (µmol/l)</td>
<td>54.0 ± 15.8</td>
</tr>
<tr>
<td>Total protein (g/l)</td>
<td>62.5 ± 2.8</td>
<td>Vit E (LS)</td>
<td>10.5 ± 3.2</td>
</tr>
</tbody>
</table>

Correlation coefficients between TAC and some individual antioxidants and P values are given in Table 5.
Discussion

Limited number of studies and experimental data about free radicals and serum total antioxidant capacity in dogs are available in spite of a great interest and progress in this area of medicine. Our study included a uniform population of beagle dogs with particular emphasis on their general health status to establish a basis for future investigation of role of TAC in diseases of dogs as the species most frequently studied in veterinary medicine. Based on a number of haematological and biochemical values that were all within physiological ranges, the population investigated was considered healthy. This might be an indirect proof for obtained TAC values to be regarded as normal for dogs. The serum TAC values were obtained by using TAS kit that enables rapid and reproducible measurement of total antioxidant capacity of serum or plasma. The range for TAC in investigated population, expressed as mean ± SD, was 1.08 ± 0.08 mmol/l and in accordance with data obtained by Gaal and Kopal (1997) with the mean value of 1.10 ± 0.45 mmol/l, when the same method was used. There were no data available about the breed and general health status of the population investigated in this comparable study.

Several human studies investigated a correlation between TAC and different individual antioxidants, i.e. uric acid as major antioxidant in humans, albumin, vitamin A and E and bilirubin (Ca o and Prior 1998; Chapple 1997; Dailly et al. 1998; Lands et al. 2000; Moore et al. 1994). As no data could be obtained about the correlation between TAC and specific individual antioxidants in dogs, our study also focused on interrelation between TAC and albumin, total bilirubin, vitamin A and E, ß-carotene and Vit E(LS). As abnormally high serum lipid concentrations can lead to false interpretation of serum vitamin E concentration we also correlated Vit E(LS) – lipid standardised vitamin E, calculated as the ratio of vitamin E to the sum of cholesterol and triglycerides (Benzie et al. 1998; Jost et al. 1999).

Most of human studies showed a correlation of TAC and individual antioxidant parameters. Although statistically significant, the correlations obtained were relatively small and considered related to a complex, non-linear interaction between various components while studies on groups of human patients with particular disease i.e. cystic fibrosis show different pattern of interrelation like significant correlation with uric acis but not with albumin, vitamin A or E or lymphocyte glutathione concentration (Lands et al. 2000). In experimentally induced diabetes in rats TAC significantly correlated with plasma albumin levels (Feillet-Coudray et al. 1999) or direct, highly significant correlation of TAC and bilirubin was found in neonatal plasma (Gopinathan et al. 1994). As no studies

<table>
<thead>
<tr>
<th>Individual antioxidant</th>
<th>Correlation coefficient (r) with TAC</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Albumin</td>
<td>0.18</td>
<td>0.46</td>
</tr>
<tr>
<td>Total bilirubin</td>
<td>-0.30</td>
<td>0.20</td>
</tr>
<tr>
<td>Vitamin A</td>
<td>-0.15</td>
<td>0.55</td>
</tr>
<tr>
<td>ß-carotene</td>
<td>-0.13</td>
<td>0.59</td>
</tr>
<tr>
<td>Vitamin E</td>
<td>0.14</td>
<td>0.58</td>
</tr>
<tr>
<td>Vit E(LS)</td>
<td>0.20</td>
<td>0.43</td>
</tr>
</tbody>
</table>

Table 5

Least squares linear regression correlation of TAC values with some individual antioxidants

TAC negatively correlated with total bilirubin (r = -0.30), vitamin A (r = -0.15) and ß-carotene (r = -0.13). There was a positive, but small correlation with albumin (r = 0.18), vitamin E (r = 0.14) and Vit (LS) (r = 0.20)
of interrelation between TAC and individual antioxidants in dogs could be found in available literature we were not able to make comparisons with related studies although our data showed somehow different results compared to human studies. None of the correlations was significant whereas some correlated negatively, like total bilirubin (r = -0.30), vitamin A (r = -0.15) and β-carotene (r = -0.13) and others positively, like albumin (r = 0.18), vitamin E (r = 0.14) and Vit E(LS) (r = 0.20). Compared to human studies there was no attempt to make correlation between TAC and uric acid due to its extremely low serum concentrations (reference range between 0 and 59 µmol/l) and clinical insignificance except probably in Dalmatians. They are known to have a defect in uric acid metabolism due to a failure to convert uric acid to soluble salts along with impaired transport of urate across the hepatocyte membrane and in hyperuricaemic dogs with portosystemic vascular anomalies due to liver insufficiency (Center and Magne 1990).

Most investigators conclude that TAC appears to represent a mixed antioxidant response, rather than response to a single antioxidant. While being responsive to oxidative stress, the mechanisms of the response may differ between clinical situations, such that the clinical significance of changes in serum TAC remains to be defined. Further investigations should be oriented towards determination of TAC and its correlation with different individual antioxidants in a group of canine population with different, accurately defined pathological and clinical syndromes.

References


