

Use of Medetomidine for Sedation in the Laboratory Rats (*Rattus norvegicus*)

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AbstractHauptman K., Jekl V. Jr., Knotek Z.: *Use of Medetomidine for Sedation in the Laboratory Rats (*Rattus norvegicus*)*. Acta Vet. Brno 2003, 72: 583-591.

Sixty-three rats (*Rattus norvegicus*) assigned to ten groups of six to seven animals were evaluated for depth and quality of sedation induced by intramuscular medetomidine. The first group received a dose of 50 µg/kg of medetomidine and the dosages in the other groups were 150 µg/kg, 200 µg/kg, 250 µg/kg, 300 µg/kg, 350 µg/kg, 400 µg/kg, 450 µg/kg and 500 µg/kg respectively. Respiratory rate, peripheral pulse, blood oxygen saturation, and disappearance and recovery of reflexes were evaluated at 5-minute intervals for the period of 60 minutes from the medetomidine injection. Disappearance of the lateral reflex was observed within 15 minutes in the 3rd group and within 10 minutes in the 4th to 10th group. The peak of the sedation of the rat population was located between minute 10 and minute 20. All groups showed a drop in pulse rate, namely by 26% on average in the 2nd group, by 18% in the 3rd group, and by over 30% in groups 4 to 10. In groups 2 and 4, the maximum drop in pulse rate in minute 60 compared with minute 5 was by 30%; the drop was by 63% in group 10. Statistical evaluation of blood oxygen saturation did not reveal any statistically significant differences between the individual groups. The average blood oxygen saturation was $90.9 \pm 4.34\%$. Recovery of reflexes after the atipamezole injection was monitored at 1-minute intervals. All reflexes under evaluation were recovered within 5 minutes. Our recommendation regarding rat sedation is to apply medetomidine from 150 to 250 µg/kg. Higher dosages induce a relatively strong respiratory depression.

Rodents, anaesthesia, sedation monitoring

Small mammals represent a specific group of patients in clinical veterinary practice. For reasons of rather difficult patient handling, a number of procedures that are part of the routine checkup of small mammals (e.g. blood sample taking, skin biopsy, X-ray and ultrasonography examinations) require patient sedation. The drugs recommended for sedation of small mammals are the following: acepromazine, diazepam, midazolam, xylazine, ketamine, and tiletamine-zolazepam (Hess et al. 1984; Flecknell 1991; Mason 1997; Cantwell 2001). Some of them are designed for special indications e.g. diazepam and midazolam can be used in aggressive and nervous animals thanks to their anxiolytic and anticonvulsive effect and the possibility of antagonization by flumazenil (Clarke 1992). Acepromazine is used, among other things, for its antiemetic and spasmolytic effect (Thurmon et al. 1996). Xylazine is characterized by sedative and myorelaxation effects and a short-time analgesia (Klein and Klide 1989; Lukasik 1999). Ketamine in higher dosages induces perfect immobilization, which may, however, be occasionally associated with undesirable effects, especially convulsions (Hess et al. 1984). Tiletamine-zolazepam have a relatively fast onset of sedation. Also, the analgesic effect of the combination is milder and irreversible renal damage may occur in some animals (rabbit) (Brammer et al. 1991; Doerning et al. 1992).

To improve the quality of sedation, especially with respect to the induction and recovery speed, the alpha-2-adrenergic agonist medetomidine is used to a relatively great extent (Hu et al. 1992). The affinity of medetomidine to alpha-2 receptors is ten times higher than that

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of xylazine and the drug can be fully antagonized by atipamezole (Ko et al. 1997). The undesirable effects of medetomidine include cardiopulmonary depression including brachycardia, hypertension, and reduced cardiac output (Muir and Mason 1996). Medetomidine is used in different animal species (Jalanka 1989; Klein and Klide 1989; Jalanka et al. 1990; Flecknell and Liles 1996; Jonson-Delaney 1999; Langan et al. 2000). The dosages recommended for rats range between 30 and 1 000 $\mu\text{g}/\text{kg}$ (Jonson-Delaney, 1999; Cantwell 2001). On the other hand, there is a lack of more detailed information on the course of sedation of rats by medetomidine in a narrower range of dosages. The aim of this study was therefore to identify and propose an optimum dosage of medetomidine for sedation of rats (*Rattus norvegicus*), i.e. to narrow down the range of dosages commonly quoted in literature.

Materials and Methods

Animals and their health condition

The trial included 63 rats (*Rattus norvegicus*) of the Wistar strain, of which 16 were males and 47 females, aged from 10 to 14 months, with weight ranging from 320 to 460 g. The animals were SPF rats screened in order to eliminate pathogens according to FELASA standards. A control clinical examination including checking the fur, the colour of the mucosae, checking the size of submandibular and popliteal lymph nodes by palpation, auscultation of the cardiovascular and respiratory systems, and palpation of the abdominal cavity was performed prior to inclusion in the trial and immediately before sedation. The animals were clinically healthy individuals. They were randomised into 10 groups of 6 to 7 (see Table 1). The rats were not given any food for 4 hours prior to medetomidine injection; water intake remained unrestricted.

Table 1
Experimental animals

Group	Number of rats			Mean body weight (g)	Medetomidine IM ($\mu\text{g}/\text{kg}$)	Atipamezole* IM ($\mu\text{g}/\text{kg}$)
	Total	Males	Females			
1.	6	4	2	382 \pm 97.5	50	25
2.	7	3	4	429 \pm 27.9	100	50
3.	7	3	4	433 \pm 66.3	150	75
4.	7	4	3	447 \pm 95.7	200	100
5.	6	6	0	372 \pm 27.9	250	125
6.	6	4	2	430 \pm 147.4	300	150
7.	6	6	0	352 \pm 24.0	350	175
8.	6	6	0	335 \pm 28.1	400	200
9.	6	6	0	340 \pm 39.5	450	225
10.	6	5	1	388 \pm 27.9	500	250

* atipamezole (Antisedan inj., Pfizer) was injected 60 minutes after medetomidine (Domitor inj., Pfizer)

Sedative and antidote

Medetomidine (Domitor inj., Pfizer) was intramuscularly applied into the left hind leg (*m. semimembranosus et semitendinosus*). The patients assigned to the different groups received 50 $\mu\text{g}/\text{kg}$ (group 1), 100 $\mu\text{g}/\text{kg}$ (group 2), 150 $\mu\text{g}/\text{kg}$ (group 3), 200 $\mu\text{g}/\text{kg}$ (group 4), 250 $\mu\text{g}/\text{kg}$ (group 5), 300 $\mu\text{g}/\text{kg}$ (group 6.), 350 $\mu\text{g}/\text{kg}$ (group 7), 400 $\mu\text{g}/\text{kg}$ (group 8), 450 $\mu\text{g}/\text{kg}$ (group 9), 500 $\mu\text{g}/\text{kg}$ (group 10) of medetomidine. Sixty minutes after the medetomidine injection, the rats were administered atipamezole into the same muscle region of the right hind leg (Antisedan inj., Pfizer), the dosage being the half of the initial dose of medetomidine. The parameters monitored were the onset of resuming a sternal position spontaneously, recovery of the palpebral reflex, and recovery of surface and depth sensibility.

Reflex and sensibility check

The rats were placed on a thermal pad of 39 °C during the sedation. The times of resuming a lateral position, of the loss of the palpebral reflex, and of the loss of the surface and depth sensibility were recorded. The lateral position was regarded as resumed when the animal was not able to go back to a sternal position from the lateral position induced by the progressive sedation. The palpebral reflex was evoked by touching the lower eyelid and was

regarded as present when the animal responded to stimulation by closing the eyelids. Surface tactile sensibility was checked by clamping the skin in the region of the last thoracic vertebra with forceps. Depth sensibility was provoked by clamping the interdigital area of the hind leg with forceps.

Pulse, respiratory rate, and blood oxygen saturation monitoring

After resuming a lateral position spontaneously, a finger clamp of the pulse oximeter (V3301 Pulse oximeter, SurgiVet, USA) was attached to the left front leg of the animal. The probe was located in the *a. antebrachialis superficialis*, *a. radialis superficialis*, *a. ulnaris*, *v. cephalica*, *v. ulnaris* area. Pulse rate and respiratory rate values as well as values of blood oxygen saturation (SpO₂) were recorded at 5-minute intervals for 60 minutes. Respiratory rate in the rats was evaluated by the control of breast movements. For the rats from groups 2 to 10, in whom the lateral reflex disappeared and respiratory rate, pulse rate, and blood oxygen saturation were evaluated, the obtained values were adjusted to the mean using the paired t-test (Microsoft Excel 2000). The disappearance and recovery of the individual reflexes were evaluated using the Fischer test (Sisa Table version).

Results

Group 1 (50 µg/kg of medetomidine) was excluded from statistical processing of pulse rate and respiratory rate and blood oxygen saturation since all rats assigned to the group showed only very weak signs of sedation (reduced interest in food and other animals in the cubicle). Neither the lateral nor the other reflexes monitored disappeared and the animals kept trying to take up a sternal position. Due to motive activity of the animals, the finger probe of the pulse oximeter could not be attached. There was one rat in group 2 (100 µg/kg of medetomidine) in whom sedation did not succeed and neither pulse rate nor blood oxygen saturation could be taken.

The average values of respiratory and pulse rate measured in the individual groups of rats are presented in Tables 2 and 3. In all monitored groups respiratory rate and pulse rate decreased gradually. The statistical differences between the times are stated in Tables 2 and 3. In group 2 statistically significant ($P < 0.05$) decreases in respiratory rate occurred between minute 15 and minute 20 and between minute 55 and minute 60. In group 3 a statistically significant ($P < 0.05$) decrease in respiratory rate occurred between minute 5 and minute 10, between minute 10 and minute 15, and between minute 30 and minute 35. In group 4 statistically significant ($P < 0.05$) decreases in respiratory rate occurred between minute 15 and minute 20. In the groups 5 to 7 a statistically significant ($P < 0.05$) decrease in respiratory rate occurred between minute 5 and minute 10. In group 8 there was no statistically significant difference between subsequently taken respiratory rates. In group 9 a statistically significant ($P < 0.05$) decrease in respiratory rate occurred between minute 50 and minute 55. In group 10 statistically significant ($P < 0.05$) decreases in respiratory rate occurred between minute 20 and minute 25, between minute 25 and 30, and between minute 45 and 50. The drop in respiratory rate in minute 60 compared with minute 5 in groups 2 to 4 was by 30 % at the most. On the other hand, the same drop was by 63% in group 10.

The average drop in pulse rate was by 26% in group 2, by 18% in group 3, and by over 30% in groups 4 to 10. The evaluation of blood oxygen saturation showed no significant differences between the groups. The average blood oxygen saturation was $90.9 \pm 4.34\%$.

From the dosage of 150 µg/kg of medetomidine (group 3) up, the lateral and the palpebral reflexes as well as surface tactile sensibility disappeared in all rats. Group 3 was subjected to a statistical comparison with all remaining groups. The disappearance of the lateral and the palpebral reflex in any of the groups did not differ from group 3 at the level of statistical significance. The differences in the disappearance of surface tactile sensibility and the corneal reflex are presented in Table 4. Depth tactile sensibility remained unimpaired in all animals without exception. The corneal reflex remained unimpaired for quite long including the cases of higher medetomidine dosages (15 minutes after injection of the anaesthetic). After the period, the reflex had disappeared in all rats from groups 5 and 7 to 10, but remained intact in one animal from group 6 (300 µg/kg) for as long as 25 minutes after injection of the

Table 2
Mean respiratory rate in rats sedated with medetomidine (Domitor inj., Pfizer)

TAM (min)	Rats (n)											
	Medetomidine ($\mu\text{g}/\text{kg}$) IM						Respiratory rate (x/min)					
	100 $\mu\text{g}/\text{kg}$	150 $\mu\text{g}/\text{kg}$	200 $\mu\text{g}/\text{kg}$	250 $\mu\text{g}/\text{kg}$	300 $\mu\text{g}/\text{kg}$	350 $\mu\text{g}/\text{kg}$	400 $\mu\text{g}/\text{kg}$	450 $\mu\text{g}/\text{kg}$	500 $\mu\text{g}/\text{kg}$	6	6	6
	7	7	7	6	6	6	6	6	6	6	6	6
	x \pm SD	x \pm SD	x \pm SD	x \pm SD	x \pm SD	x \pm SD	x \pm SD	x \pm SD	x \pm SD	x \pm SD	x \pm SD	x \pm SD
5	71.3 \pm 16.86	94.3 ^a \pm 25.31	78.3 \pm 13.49	71.0 ^a \pm 16.91	71.7 ^a \pm 17.82	69.0 \pm 14.57	61.7 \pm 17.04	66.0 \pm 16.54	101.3 \pm 15.47			
10	72.2 \pm 16.98	90.1 ^b \pm 22.81	71.1 \pm 18.93	59.0 ^b \pm 20.58	60.0 ^b \pm 18.24	45.0 ^b \pm 5.02	63.0 \pm 17.70	61.7 \pm 19.12	106.0 \pm 18.76			
15	72.2 ^a \pm 12.88	79.4 ^c \pm 21.99	74.6 ^a \pm 20.29	55.0 \pm 21.94	64.0 \pm 15.95	47.0 \pm 7.97	57.7 \pm 10.69	55.0 \pm 18.75	96.0 \pm 18.02			
20	66.3 ^b \pm 11.20	77.1 \pm 19.18	60.9 ^b \pm 14.46	57.0 \pm 16.43	55.0 \pm 13.90	46.0 \pm 7.27	53.0 \pm 16.72	50.0 \pm 11.17	87.3 ^a \pm 20.73			
25	67.0 \pm 13.19	81.4 ^d \pm 17.58	60.0 \pm 12.49	53.0 \pm 14.90	54.7 \pm 13.00	47.0 \pm 5.90	54.3 \pm 16.85	48.0 \pm 8.49	62.3 ^b \pm 19.08			
30	66.3 \pm 8.43	73.4 ^e \pm 17.95	64.3 \pm 13.29	49.3 \pm 11.15	53.0 \pm 10.33	45.0 \pm 9.86	55.0 \pm 18.36	44.0 \pm 9.03	44.7 ^c \pm 14.68			
35	65.0 \pm 10.41	74.3 \pm 17.68	62.6 \pm 10.31	47.7 \pm 15.67	51.0 \pm 12.44	44.0 \pm 6.20	51.0 \pm 12.44	46.0 \pm 11.17	47.3 \pm 23.35			
40	62.7 \pm 9.44	73.1 \pm 17.12	58.3 \pm 12.35	49.0 \pm 16.28	53.7 \pm 17.68	42.0 \pm 3.79	53.0 \pm 8.83	42.0 \pm 8.49	40.0 \pm 14.03			
45	64.7 \pm 8.07	76.9 \pm 14.60	58.3 \pm 9.62	44.0 \pm 9.03	52.0 \pm 18.46	41.0 \pm 4.52	49.0 \pm 8.83	43.0 \pm 12.25	37.0 ^d \pm 7.01			
50	62.3 \pm 9.91	71.7 \pm 11.86	60.0 \pm 13.86	43.0 \pm 8.83	51.0 \pm 14.13	41.0 \pm 4.52	46.0 \pm 12.39	41.0 ^a \pm 12.82	30.0 ^e \pm 5.37			
55	66.5 ^c \pm 11.31	66.9 \pm 8.47	58.0 \pm 12.00	43.0 \pm 9.61	47.0 ^c \pm 19.87	39.0 \pm 6.29	47.0 \pm 11.01	36.0 ^b \pm 8.49	36.0 \pm 5.37			
60	59.3 ^d \pm 11.22	68.7 \pm 11.44	56.3 \pm 11.57	45.0 \pm 10.18	53.0 ^d \pm 17.56	44.0 \pm 4.90	46.0 \pm 9.80	39.0 \pm 8.27	36.0 \pm 3.79			

TAM – time after medetomidine injection
a-i significant difference (p<0.05) within the same group

Table 3
Peripheral pulse in rats sedated with medetomidine (Domitor inj., Pfizer)

TAM (min)	Rats (n)											
	Medetomidine ($\mu\text{g}/\text{kg}$) IM						Pulse frequency (x/min)					
	100 $\mu\text{g}/\text{kg}$	150 $\mu\text{g}/\text{kg}$	200 $\mu\text{g}/\text{kg}$	250 $\mu\text{g}/\text{kg}$	300 $\mu\text{g}/\text{kg}$	350 $\mu\text{g}/\text{kg}$	400 $\mu\text{g}/\text{kg}$	450 $\mu\text{g}/\text{kg}$	500 $\mu\text{g}/\text{kg}$	6	6	6
5	280.5 ^a ± 13.81	242.7 ± 25.29	227.1 ± 17.11	253.2 ^a ± 19.56	222.2 ± 19.66	215.3 ± 23.23	213.8 ^a ± 19.89	232.8 ± 22.60	222.3 ^a ± 26.63	6	6	6
10	260.7 ^b ± 26.87	235.1 ^a ± 22.51	220.9 ^a ± 21.22	239.3 ^b ± 30.02	212.3 ^a ± 20.77	212.3 ± 21.92	202.8 ^b ± 21.19	230.5 ± 26.79	197.7 ^b ± 36.66	6	6	6
15	259.7 ^c ± 24.57	222.7 ^b ± 23.52	214.6 ^b ± 19.52	228.8 ^c ± 26.37	201.0 ^b ± 27.36	204.5 ± 29.76	187.7 ^c ± 29.35	223.0 ^a ± 23.90	185.0 ^c ± 35.37	6	6	6
20	237.5 ^d ± 20.52	214.0 ^c ± 21.15	195.6 ^c ± 16.03	211.2 ^d ± 34.25	189.3 ^c ± 26.82	185.2 ± 32.65	178.0 ^d ± 20.06	207.3 ^b ± 23.61	177.5 ± 42.76	6	6	6
25	227.8 ^e ± 19.37	214.7 ^d ± 13.61	191.1 ^d ± 17.28	201.8 ^e ± 33.78	177.7 ± 25.94	181.2 ± 30.39	164.5 ^e ± 20.52	191.0 ^c ± 23.49	176.5 ± 42.01	6	6	6
30	216.5 ^f ± 14.80	211.0 ^e ± 11.73	181.3 ± 26.11	190.2 ^f ± 34.02	173.5 ^d ± 14.27	173.8 ± 25.36	158.0 ± 16.11	174.2 ^d ± 27.79	171.2 ^d ± 35.55	6	6	6
35	213.2 ± 11.62	207.1 ^f ± 12.63	179.0 ^e ± 23.03	184.2 ^g ± 32.03	162.2 ^e ± 16.41	174.7 ± 28.42	155.3 ± 10.15	163.5 ^e ± 29.62	162.5 ^e ± 35.74	6	6	6
40	209.7 ± 16.51	203.1 ^g ± 14.35	166.4 ^f ± 13.39	181.0 ± 26.74	156.2 ^f ± 14.32	165.7 ^a ± 24.89	150.7 ± 8.38	156.7 ^f ± 30.16	164.8 ^f ± 30.02	6	6	6
45	208.0 ± 18.73	200.3 ± 10.45	163.1 ± 15.74	173.3 ± 33.43	147.5 ^g ± 16.84	154.8 ^b ± 27.37	145.7 ^f ± 16.13	151.2 ^g ± 28.06	162.8 ± 28.40	6	6	6
50	200.7 ^g ± 24.47	200.1 ± 16.42	156.1 ± 16.76	170.5 ^h ± 32.97	142.3 ^h ± 10.41	156.2 ^c ± 24.36	140.2 ^g ± 14.62	147.8 ± 30.29	157.5 ± 28.93	6	6	6
55	201.3 ^h ± 19.80	197.1 ± 14.71	153.7 ± 16.20	159.5 ⁱ ± 28.79	136.0 ⁱ ± 12.39	150.5 ± 28.50	139.2 ± 17.65	144.3 ± 28.46	158.3 ^g ± 30.12	6	6	6
60	206.8 ± 21.26	198.6 ± 19.27	153.1 ± 14.62	161.0 ± 27.22	136.2 ± 12.09	147.0 ± 27.62	138.0 ± 23.22	140.5 ± 30.89	152.7 ^h ± 27.49	6	6	6

TAM – time after medetomidine injection

a-i – significant difference ($p < 0.05$) within the same group

Table 4
Disappearance of reflexes and sensibility in rats after medetomidine injection (Domitor inj., Pfizer)

Reflexy	TAM (min)	Medetomidine dosage (µg/kg) IM												
		Number of rats in group (n)												
		50	100	150	200	250	300	350	400	450	500			
Lateral reflex	5	0	2	4	4	6	3	6	5	5	5	6	6	6
	10	0	4	6	7	6	6	6	6	6	6	6	6	6
	15	0	5	7	7	6	6	6	6	6	6	6	6	6
	20-50	0	6	7	7	6	6	6	6	6	6	6	6	6
	55-60	0	5	7	7	6	6	6	6	6	6	6	6	6
	5	0	0	0	1	2	2	6*	5*	3	4	5	6	6
	10	0	3	3	7	5	3	6*	6*	4	5	6	6	6
	15	0	5	5	7	6	5	6	6	6	6	6	6	6
	20-25	0	5	6	7	6	6	6	6	6	6	6	6	6
	30-45	0	5	7	7	6	6	6	6	6	6	6	6	6
Surface sensibility	50	0	5	6	7	6	6	6	6	6	6	6	6	6
	55	0	5	5	7	7	6	6	6	6	6	6	6	6
	60	0	5	5	7	6	6	6	6	6	6	6	6	6
	5	0	5	4	6	6	5	6	6	6	6	6	6	5
	10-50	0	6	7	7	6	6	6	6	6	6	6	6	6
	55-60	0	5	7	7	6	6	6	6	6	6	6	6	6
	5	0	0	0	1	0	0	5	4	0	3	6	6	6
	10	0	1	0	2	6	0	5	5	2	6	6	6	6
	15	0	2	0	5	6	3	5	6	3	6	6	6	6
	20	0	3	3	5	6	5	6	6	6	5	6	6	6
25	0	3	3	5	6	5	6	6	6	6	6	6	6	
30	0	3	4	5	6	6	6	6	6	6	6	6	6	
35	0	3	5	5	6	6	6	6	6	6	6	6	6	
40-45	0	3	3	5	6	6	6	6	6	6	6	6	6	
50	0	3	2	4	6	6	6	6	6	6	6	6	6	
55	0	2	3	5	6	6	6	6	6	6	6	6	6	
60	0	2	1	4	6	5	6	6	6	6	6	6	6	

TAM – time after medetomidine injection * significant difference (p<0.05) in comparison with the 3rd group (150 µg/kg)

tranquillizer. In group 3 a significant ($P < 0.05$) difference in disappearance of the corneal reflex occurred between minute 15 and minute 20. In group 4 a significant ($P < 0.05$) difference in disappearance of the corneal reflex occurred between minute 10 and minute 15. In group 5 a significant ($P < 0.05$) difference in disappearance of the corneal reflex occurred between minute 10 and minute 15. In group 6 significant ($P < 0.05$) differences in disappearance of the corneal reflex occurred between minute 5 and minute 10 and between minute 10 and minute 15. In group 5 a significant ($P < 0.05$) difference in disappearance of the corneal reflex occurred between minute 5 and minute 10. In group 3 a significant ($P < 0.05$) difference in disappearance of the corneal reflex occurred between minute 10 and minute 15.

The recovery of the monitored reflexes after the atipamezole injection was within 3 minutes in group 2, within 2 minutes in groups 3 and 4, within 3 minutes in group 5, within 5 minutes in group 6, within 4 minutes in group 7, within 2 minutes in group 8, within 4 minutes in group 9, and within 3 minutes in group 10 (see Table 5).

Discussion

We do not regard the palpebral or the corneal reflex as suitable for evaluating sedation due to the lack of reliability of their evaluation. Moreover, evaluating the loss of the corneal reflex by palpation may result in corneal damage (Smith 1993). There are authors who do not evaluate the corneal and the palpebral reflex at all (Hu et al. 1992). It was evaluation of the lateral reflex and of surface and depth tactile sensibility that we regarded as suitable for this purpose. We defined the onset of sedation as the moment of loss of the lateral reflex. In our population of rats the dosage of 50 $\mu\text{g}/\text{kg}$ of medetomidine did not suffice for achieving good-quality sedation. In contrast to cats and dogs, small mammals require higher dosages of medetomidine (Thurmon et al. 1996; Mason 1997; Muir et al. 1999). The medetomidine dosages recommended for rats by Johnson-Delaney (1999) range between 150 and 250 $\mu\text{g}/\text{kg}$. Increasing the dosages further did not improve the quality of sedation while evoking a relatively strong respiratory depression.

A drop in the pulse rate was recorded in rats of all the monitored groups. This result is in accordance with the effects of medetomidine on the cardiovascular system of dogs and cats (Lukasik 1999; Muir and Mason 1996) as well as of wild animals (Jalanka et al. 1990). Not even the highest dosage of 500 $\mu\text{g}/\text{kg}$ of medetomidine did, however, cause a decrease in blood oxygen saturation in the monitored population.

Lukasik (1999) writes of recovery of reflexes after atipamezole injection within 5 to 10 minutes. In our trial, the reflexes in rats had recovered within 5 minutes.

Využití medetomidinu pro sedaci u laboratorního potkana *Rattus norvegicus*

U 63 potkanů (*Rattus norvegicus*), rozdělených po 6-7 do deseti skupin jsme hodnotili hloubku a kvalitu sedace navozené intramuskulární aplikací medetomidinu. V první skupině byl medetomidin aplikován v dávce 50 $\mu\text{g}/\text{kg}$, v dalších skupinách 100 $\mu\text{g}/\text{kg}$, 150 $\mu\text{g}/\text{kg}$, 200 $\mu\text{g}/\text{kg}$, 250 $\mu\text{g}/\text{kg}$, 300 $\mu\text{g}/\text{kg}$, 350 $\mu\text{g}/\text{kg}$, 400 $\mu\text{g}/\text{kg}$, 450 $\mu\text{g}/\text{kg}$ a 500 $\mu\text{g}/\text{kg}$. Dechová frekvence, periferní puls, saturace krve kyslíkem, ztráta a opětovný návrat reflexů byly vyhodnocovány po dobu 60 minut od okamžiku aplikace medetomidinu v pětiminutových intervalech. Ke ztrátě laterálního reflexu došlo u 3. skupiny do 15 minut u 4.–10. skupiny do 10 minut. Za vrchol sedace považujeme u sledovaného souboru potkanů interval mezi 10. a 20. minutou. U všech skupin došlo k poklesu pulsové frekvence. U 2. skupiny se snížila v průměru o 26 %, u 3. skupiny o 18 %, u 4.–10. skupiny o více než 30 %. U skupin 2. až 4. došlo ke snížení dechové frekvence v 60. minutě oproti 5. minutě maximálně o 30 %. U 10. skupiny došlo ke snížení o 63 %. Při statistickém hodnocení saturace krve kyslíkem jsme nezjistili žádné statisticky významné rozdíly mezi jednotlivými

Table 5
Recovery of reflexes and sensibility in rats after medetomidine and atipamezole

Reflexes	Recovery of reflexes after atipamezole injection (min)	Atipamezole ($\mu\text{g}/\text{kg}$) IM															
		50	100	150	200	250	300	350	400	450	500	Number of rats in group (n)					
		6	7	7	7	6	6	6	6	6	6	6	6	6	6		
Lateral reflex	1	6	5	4	5	0	1	0	0	0	0	0	0	0	Number of rats with recovery of reflexes (n)		
	2	6	5	7	7	5	3	2	6	6	3	5	5	5			
	3	6	7	7	7	6	5	4	6	6	5	6	6	6			
	4	6	7	7	7	6	5	6	6	6	5	6	6	6			
	5	6	7	7	7	6	6	6	6	6	6	6	6	6			
Surface sensibility	1	6	5	7	5	1	2	1	1	1	0	0	0	0			
	2	6	5	7	7	5	4	2	6	6	3	5	5	5			
	3	6	7	7	7	6	6	4	6	6	5	6	6	6			
	4	6	7	7	7	6	6	6	6	6	6	6	6	6			
	5	6	7	7	7	6	6	6	6	6	6	6	6	6			
Palpebral reflex	1	6	5	5	5	0	1	0	0	0	0	0	0	0			
	2	6	5	7	7	5	3	2	6	6	3	5	5	5			
	3	6	7	7	7	6	5	4	6	6	5	6	6	6			
	4	6	7	7	7	6	5	6	6	6	6	6	6	6			
	5	6	7	7	7	6	6	6	6	6	6	6	6	6			
Corneal reflex	1	6	5	7	7	2	3	1	1	1	0	0	0	0			
	2	6	5	7	7	5	5	3	6	6	3	5	5	5			
	3	6	7	7	7	6	5	5	6	6	6	6	6	6			
	4	6	7	7	7	6	5	6	6	6	6	6	6	6			
	5	6	7	7	7	6	6	6	6	6	6	6	6	6			

skupinami. Průměrná saturace krve kyslíkem byla 90.9 ± 4.34 %. Opětovný návrat reflexů po aplikaci atipamezolu byl kontrolován v minutových intervalech. K návratu všech hodnocených reflexů došlo do 5 minut. K sedaci potkanů doporučujeme dávky medetomidinu v rozmezí 150 – 250 $\mu\text{g}/\text{kg}$. Vyšší dávky způsobují poměrně silnou depresi dýchání.

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References

- BRAMMER, DW, DOERING, BJ, CHRISP, CE, RUSH, HG 1991: Anesthetic and nephrotoxic effect of Telazol in New Zealand white rabbits. *Lab Anim Sci* **41**: 432-435
- CANTWELL, SL 2001: Ferret, rabbit, and rodent anesthesia. *Veter Clin North Amer Exotic Anim Pract* **4**: 169-191
- CLARKE, KW 1992: Premedication and sedation. In: HILBERRY, ADR (Ed.): *Manual of Anaesthesia for Small Animal Practice*. 3rd ed., BSAVA, Cheltenham, pp. 39-49
- DOERING, BJ, BRAMMER, DW, CHRISP, CE, RUSH, HG 1992: Nephrotoxicity of tiletamine in New Zealand white rabbits. *Lab Anim Sci* **42**: 267-269
- FLECKNELL, P 1991: Anaesthesia and postoperative care of small practice. In *Practice* **9**: 180-189
- FLECKNELL, PA, LILES, JH 1996: Halotane anaesthesia in the rabbit: A comparison of the effects of medetomidine, acepromazine, and midazolam on breath-holding during induction. *J Ass Veter Anaest* **23**: 11-14
- HESS, L, DVOŘÁČEK, I, SVOBODNÍK, J 1984: Hlodavci. In: HESS L., DVOŘÁČEK I., SVOBODNÍK, J (Ed.): *Anestezie laboratorních zvířat*, Avicenum, Praha, pp. 158-189
- HU, C, FLECKNELL, PA, LILES, JH 1992: Fentanyl and medetomidine anaesthesia in the rat and its reversal using atipamezole and either nalbuphine or butorphanol. *Lab Anim* **26**: 15-22
- JALANKA, HH 1989: Medetomidine and ketamine-induced immobilization of snow leopards (*Panthera uncia*): Doses, evaluation, and reversal by atipamezole. *J Zoo Wildlife Med* **20**: 154-162
- JALANKA, HH, BENGT, O, ROEKEN, BO 1990: The use of medetomidine, medetomidine – ketamine combinations, and atipamezole in non-domestic mammals: A review. *J Zoo Wildlife Med* **21**: 259-282
- JONSON-DELANEY, C 1999: Medetomidine in small mammals. *Exotic DVM* **5**: 35-36
- KLEIN, LV, KLIDE, AM 1989: Central alfa2-adrenergic and benzodiazepine agonists and their antagonists. *J Zoo Wildlife Med* **20**: 138-153
- KO, JCH, MCGRATH, CJ, NICKLIN, CF 1997: Answers to your questions about medetomidine and atipamezole. *Vet Med* **92**: 415-425
- LANGAN, JN, SCHUMAKER, J, POLLOCK, C, OROSZ, SE, JONES, MP, HARVEY, RC 2000: Cardiopulmonary and anesthetic effect of medetomidine-ketamine-butorphanol and antagonism with atipamezole in servals (*Felis serval*). *J Zoo Wildlife Med* **34**: 329-334
- LUKASIK, VM 1999: Premedication and sedation. In: SEYMOUR, C, GLEED, R (Ed.): *Manual of Small Animal Anaesthesia and Analgesia*. BSAVA, Cheltenham, pp. 71-85
- MASON, DE 1997: Anesthesia, analgesia, and sedation for small mammals. In: HILLYER, E V, QUESENBERRY, KE (Ed.): *Ferrets, Rabbits, and Rodents Clinical Medicine and Surgery*. W. B. Saunders Comp., Philadelphia, pp. 378-391
- MUIR, WW, FORD, JL, KARPA, GE, HARRISON, EE, GADAWSKI, JE 1999: Effect of intramuscular administration of low doses of medetomidine and medetomidine-butorphanol in middle-aged and old dogs. *JAVMA*, **215**: 1116-1120
- MUIR, WW, MASON, D 1996: Cardiovascular system. In: THURMON, JC, TRANQUILLI, WJ, BENSON, GJ (Ed.): *Lumb and Jones' Veterinary Anesthesia*, 3rd ed., Williams and Wilkins, Baltimore, pp. 62-114
- SMITH, W 1993: Responses of laboratory animals to some injectable anaesthetics. *Lab Anim* **27**: 30-39
- THURMON, JC, TRANQUILLI, WJ, BENSON, GJ 1996: Preanesthetics and anesthetic adjuncts. In: THURMON, JC, TRANQUILLI, WJ, BENSON, GJ (Ed.): *Lumb and Jones' Veterinary Anesthesia*. 3rd edition, Williams and Wilkins, Baltimore, pp. 183-209