Effect of Low-dose Chronic Melatonin Administration on Metabolic and Hormonal Variables in Young Laboratory Rats

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Received February 20, 2004
Accepted October 26, 2004

Abstract


The introduction of melatonin (MEL) as a drug remains open due to insufficient data about the effects of its long-term administration on metabolism. We explored effects of MEL given for 26 weeks at low doses, 4 µg/ml of tap water, on selected metabolic and hormonal indices in young female and male Sprague-Dawley rats.

Male and female rats aged 5 weeks were adapted to standard housing conditions and artificial light regimen L:D = 12:12 h. The animals were fed standard laboratory diet and drank tap water (controls) or MEL solution ad libitum. Body weight gain, food and water intake was regularly recorded. Oral glucose tolerance tests (OGTTs) were carried out before and 24 weeks after MEL administration. At the end of the experiment following an overnight fast the animals were sacrificed, selected organs and tissues were weighed, selected metabolic indices and hormone concentrations were determined in the serum, liver, heart muscle and bone marrow (femur).

Chronic MEL administration increased in females serum corticosterone concentration, decreased glycogen concentration in the liver and heart muscle, increased triacylglycerol concentration in the bone marrow. An increased absolute weight of periovarial fat and body weight gain were found in last 5 weeks of the experiment. In males, MEL decreased insulin concentration in the serum and body weight gain from week 2 to week 17 of its administration; it decreased relative liver weight in males and females. MEL did not alter food or water intake in both sexes. MEL treatment decreased glucose tolerance (i.e. reduced area under curve values in OGTTs) in males and prolonged the return of afterload glycemia values to initial level in females.

Alterations of carbohydrate metabolism regulation and of body weight gain were found in both sexes of young rats after long term melatonin administration.

Melatonin, long-term administration, metabolic indices, organ and body weights, males, females

Melatonin (MEL), the main product of the pineal gland, is a low molecular lipophilic hormone, very old from the phylogenetic point of view. Its physiological effects are pleiotropic and it is regarded as "regulator of regulators" (Reiter 1991a). MEL functions as an endogenous synchronizer of biological rhythms and modulates sleep. Therefore exogenous MEL is administered in the treatment of circadian rhythms and other functions imbalances resulting from subsonic or supersonic travel through a varied number of time zones, so called jet-lag syndrome and when working on shifts, so-called shift work-lag syndrome. MEL modulates functions of immune and endocrine system in the organism. Its antioxidizing and oncostatic properties are well-known (Tan et al. 1993 ab; Hill and Blask 1988). It displays a wide spectrum of metabolic and other physiological effects including hypothermic, sedative, hypnotic, analgetic, myorelaxing, cardio- and neuroprotective effects (Vijayalaxmi et al. 2002; Lagneux et al. 2000; Reiter et al. 1998).

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The results of studies investigating MEL effect on carbohydrate and lipid metabolism in experimental animals are controversial, glucose metabolism in particular. In certain studies MEL increased glycemia in rats (Csaba and Barath 1971) and Syrian hamsters (Ortega-Corona et al. 1991) while in the others did not alter it (Bailey et al. 1974) or, on the contrary, decreased it (Iizuka 1996). MEL administration to pinealectomized rats had similar ambiguous effects on glycemia levels (Milcu et al. 1971; Csaba and Barath 1971; Diaz and Blazquez 1986). In rabbits MEL decreased basal plasma glucose but increased it after glucose load (Dhar et al. 1983). The effect of MEL on glucose level may be explained by modification of insulin secretion and/or change of cell sensitivity to insulin. The results in this area are also controversial as inhibitory (Peschke et al. 1997) or no effects (Frankel and Strandberg, 1991) of MEL on insulin secretion from isolated pancreatic islets in rats and mice were described. Bizot-Espiard et al. (1998) found in an in vivo study that glycemia decrease after insulin load was not affected by MEL administration or pinealectomy, pinealectomy did not significantly alter basal glucose and insulin concentration in the plasma or hepatic glucose and its utilization by tissues when compared with sham-operated rats. MEL is assumed to act directly on target cells e.g. hepatocytes and pancreatic β-cells (Acuna-Castroviejo et al. 1994; Peschke et al. 2000) and it is possible that MEL and the pineal gland affect glucose metabolism through modulation of activity of suprachiasmatic hypothalamic nucleus (Margraf and Lynch 1993). Effect of exogenous MEL on lipid parameters was more exactly defined. In genetically and diet-induced hypercholesterolemic rats MEL administration decreased serum cholesterol (Aoyama et al. 1988; Mori et al. 1989). Esquifino et al. (1997) recorded a decrease in cholesterol in the serum, liver, adrenal glands and testes after pineal extract administration to hyperprolactinemic rats while pinealectomy had the opposite effect. Pharmacological doses of MEL administered in tap water during a period of 3 months reduced increased concentrations of total and LDL-cholesterol and increased plasma HDL-cholesterol in young rats fed hypercholesterolemic diet. MEL also decreased malondialdehyde and 4-hydroxyalkenal content in the liver, brain and spleen in rats fed hypercholesterolemic and normal diet (Hoyos et al. 2000). Single subcutaneous MEL administration at a dose of 1 mg/kg body weight decreased decreased concentration of total, free, esterified and HDL cholesterol and decreased the level of free fatty acids in the blood of rats (Fabis et al. 2002). Mechanism of MEL effects on cholesterol metabolism remains unknown. MEL is assumed to affect cholesterol metabolism via influence on cytokine secretion from macrophages - e.g. interleukin 2 (Morrey et al. 1994; Garcia-Mauriño et al. 1997, 1998).

This study is a follow-up research of metabolic effects of MEL (4 µg/ml tap water) administered for 10 weeks to Sprague-Dawley rats of both sexes aged 5 weeks (Marková et al. 2003). It is focused on metabolic effects of MEL administered for 26 weeks to young Sprague-Dawley rats including effects on organ weights, adipose intraabdominal tissue and body weight gain.

Materials and methods

Female and male Sprague-Dawley rats (SD) (Faculty of Medicine, P.J. Šafárik University, Košice, Slovak Republic) aged 36-37 days, weighing 115-140 g were used in the experiment. The animals were adapted to standard housing conditions (temperature 23 ± 2 °C, relative humidity 60-70%); artificial light:dark regimen LD 12:12, with lights on at 07.00 h, with intensity 150 lux per cage (TESLA, fluorescent lamps, 40 W). The rats were fed standard MP laboratory diet (Top-Dovo, Dobrá voda, Slovak Republic), drank tap water and MEL solution, respectively, ad libitum. Three to five rats were housed per cage.

MEL (Sigma, Diesenhofen, Germany) was administered in tap water at a concentration of 4 mg/ml discontinuously – from 03.00 h to 08.00 h – in the period of increased sensitivity of the organism to MEL (from 08.00 h to 03.00 h the animals were drinking tap water) for the period of 26 weeks. Ten mg of MEL were dissolved in 0.2 ml of 30% ethanol and mixed up with tap water to the required concentration. The solution of MEL was freshly prepared three times a week. The bottles with MEL solution were covered with a dark foil. The drinking water of control group contained 0.01% ethanol.

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The animals were divided into 4 groups: females and males that were administered MEL (MEL-treated females, MEL-treated males, 9 animals in each group) and control females and males that were not treated with MEL (CONT females, CONT males, 7 animals per group). The rats were weighed twice a week and once a month daily food and water intake was observed (a total of 5 measurements). Oral glucose tolerance tests (OGTTs) were carried out before MEL administration and 24 weeks thereafter as follows: 1 g of glucose per 1 kg of body weight in 20% solution was administered intragastrically to animals following overnight fasting. Twenty-five ml of the blood was collected from rat tail vein before glucose administration and 15, 30, 60 and 120 minutes after glucose load. Glucose concentration was determined enzymatically in the blood and presented as graphs. The OGTTs were evaluated by calculation of area under curve (AUC) according to the formula: 

$$AUC (\text{mmol/h}) = \frac{G_0 + 2G_{30} + 3G_{60} + G_{120}}{4}$$

where $G_0$-$G_{120}$ are the respective blood glucose values. After 26 weeks following overnight fasting the animals were sacrificed by quick decapitation between 08.00 h and 11.00 h; selected organs (liver, heart muscle, spleen, thymus, adrenal glands) and white fat (periovarial and epididymal) were weighed and the following parameters were determined in the serum, liver, heart muscle and bone marrow (femur):

- in the serum from mixed blood: concentrations of glucose, triacylglycerols, total cholesterol, phospholipids, corticosterone and insulin.
- in the liver: concentrations/contents of glycogen, triacylglycerols, total cholesterol, phospholipids, malondialdehyde (as an indicator of lipoperoxidation).
- in the bone marrow: concentration of triacylglycerols, phospholipids and malondialdehyde.
- in the heart muscle: glycogen concentration/conten.

Phospholipids were measured from lipid phosphorus according to Bartlett (1959), total cholesterol according to Zlatkis et al. (1953), glycogen according to Roe and Dailey (1966), malondialdehyde was measured in reaction with thiobarbituric acid (Satch 1978), for triacylglycerol and glucose measurement commercial sets of Lachema (Brno, Czech Republic) were used, insulin by radioimmunoassay with the use of commercial set of Linco Research (St. Charles, MO, USA), corticosterone was measured using fluorimetry according to Guillemin et al. (1958).

Results were statistically evaluated by one-way analysis of variance and Kruskall-Wallis test. The criterion for the choice of relevant test was the value of Bartlet’s number. Data are presented as means ± standard error of the mean (S.E.M) and significant differences between groups as: * (a) for $p \leq 0.05$; ** (b) for $p \leq 0.01$; *** (c) for $p \leq 0.001$.

The experiment was carried out from May to November.

Results

During the experiment food and water intake in MEL treated females and males did not differ from the controls. An average daily food and water intake was higher in males in both groups than in females in both groups. Food intake (g): CONT males 27.6 ± 0.8 vs CONT females 22.5 ± 1.7 (p ≤ 0.05); MEL treated males 26.9 ± 0.8 vs MEL treated females 21.4 ± 1.6 (p ≤ 0.05). Water intake respectively MEL solution (ml): CONT males 39.3 ± 1.5 vs CONT females 29.1 ± 0.8 (p ≤ 0.001); MEL treated males 40.8 ± 1.3 vs MEL treated females 29.6 ± 0.3 (p ≤ 0.001).

Mean daily MEL intake in females was 118.5 µg and in males 163.1 µg. An average final body weight (g) in male rats of both groups was significantly higher than in females of both groups (CONT males 347.6 ± 16.0 vs CONT females 159.1 ± 6.7 (p≤0.001); MEL treated males 348.2 ± 11.9 vs MEL treated females 173.1 ± 5.1 (p ≤ 0.001). Twenty six weeks MEL administration prominently increased absolute weight of the periovarial fat in females (Table 2) together with their body weight from day 144 to day 182 of the experiment (Fig. 1). Body weight of MEL treated males significantly reduced from day 14 to day 116; in the following period the body weight was comparable to that of controls (Fig. 1). The weight of epididymal fat was not significantly influenced by MEL compared to controls (Table 2). Chronic MEL administration
significantly increased corticosterone concentration in the serum as well as concentration of triacylglycerols in the bone marrow and decreased glycogen concentration in the liver and heart muscle in females. MEL decreased insulin concentration in the serum in males significantly ($p \leq 0.01$) (Table 1). MEL significantly decreased relative weight of the liver in both females and males; the weights of the other organs were not significantly changed (Table 2). Twenty-four weeks lasting MEL administration modified the curves of OGTTs in males in which significant reduction of AUC ($p \leq 0.01$) was noticed (Fig. 3, Table 3). In females MEL prolonged the return of glycemia to the initial level in OGTTs but did not change the values of AUC (Fig. 2, Table 3).

### Discussion

In the experiments, preceding this study, MEL was administered to male and female Sprague-Dawley rats aged 5 weeks at the concentration 4 µg/ml in tap water over 2.5 months. In males MEL decreased the concentration of serum triacylglycerols and glycogen liver content, reduced weights of several organs, epididymal fat and body weight gain. Increased liver phospholipid content, weight of the heart muscle and perivascular fat without any changes in body

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#### Table 1
The effect of 26-week-long MEL administration on selected metabolic parameters

<table>
<thead>
<tr>
<th></th>
<th>CONT-female rats n = 7</th>
<th>MEL-female rats n = 9</th>
<th>CONT-male rats n = 7</th>
<th>MEL-male rats n = 9</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>SERUM</strong></td>
<td></td>
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<tr>
<td>GLU (mmol/l)</td>
<td>6.17 ± 0.46</td>
<td>7.14 ± 0.66</td>
<td>6.47 ± 0.32</td>
<td>5.63 ± 0.31</td>
</tr>
<tr>
<td>TAG (mmol/l)</td>
<td>0.62 ± 0.05</td>
<td>0.65 ± 0.05</td>
<td>0.52 ± 0.03</td>
<td>0.59 ± 0.06</td>
</tr>
<tr>
<td>CH (mmol/l)</td>
<td>1.44 ± 0.09</td>
<td>1.38 ± 0.06</td>
<td>0.96 ± 0.04</td>
<td>0.93 ± 0.03</td>
</tr>
<tr>
<td>PL (mmol/l)</td>
<td>2.12 ± 0.13</td>
<td>2.03 ± 0.07</td>
<td>1.13 ± 0.11</td>
<td>1.08 ± 0.06</td>
</tr>
<tr>
<td>CTS (pmol/ml)</td>
<td>350.43 ± 31.43</td>
<td>761.05 ± 119.38 *</td>
<td>286.12 ± 64.37</td>
<td>255.99 ± 31.32</td>
</tr>
<tr>
<td>INS (ng/ml)</td>
<td>0.34 ± 0.03</td>
<td>0.28 ± 0.05</td>
<td>0.40 ± 0.08</td>
<td>0.24 ± 0.03 *</td>
</tr>
<tr>
<td><strong>LIVER</strong></td>
<td></td>
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</tr>
<tr>
<td>GLU (µmol/g)</td>
<td>5.22 ± 0.36</td>
<td>3.59 ± 0.48 *</td>
<td>3.50 ± 0.51</td>
<td>3.47 ± 0.35</td>
</tr>
<tr>
<td>TAG (µmol/g)</td>
<td>53.23 ± 5.83</td>
<td>46.39 ± 4.02</td>
<td>44.87 ± 6.63</td>
<td>44.24 ± 5.06</td>
</tr>
<tr>
<td>CH (µmol/g)</td>
<td>21.37 ± 1.08</td>
<td>19.63 ± 0.35</td>
<td>19.27 ± 1.06</td>
<td>20.18 ± 0.90</td>
</tr>
<tr>
<td>PL (µmol/g)</td>
<td>44.52 ± 1.78</td>
<td>43.98 ± 1.61</td>
<td>46.70 ± 0.71</td>
<td>45.47 ± 0.91</td>
</tr>
<tr>
<td>MDA (nmol/g)</td>
<td>19.28 ± 2.68</td>
<td>14.83 ± 1.98</td>
<td>12.46 ± 2.16</td>
<td>14.97 ± 1.76</td>
</tr>
<tr>
<td><strong>BONE MARROW</strong></td>
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<tr>
<td>TAG (µmol/g)</td>
<td>55.60 ± 20.19</td>
<td>120.81 ± 16.54 *</td>
<td>80.68 ± 15.05</td>
<td>106.55 ± 23.89</td>
</tr>
<tr>
<td>PL (µmol/g)</td>
<td>16.69 ± 0.76</td>
<td>16.33 ± 0.51</td>
<td>14.76 ± 0.98</td>
<td>15.18 ± 0.60</td>
</tr>
<tr>
<td>MDA (nmol/g)</td>
<td>56.45 ± 4.54</td>
<td>57.16 ± 8.73</td>
<td>43.62 ± 16.00</td>
<td>34.62 ± 3.66</td>
</tr>
<tr>
<td><strong>HEART MUSCLE</strong></td>
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<tr>
<td>GLU (µmol/g)</td>
<td>8.30 ± 1.26</td>
<td>5.12 ± 0.61 *</td>
<td>6.92 ± 0.94</td>
<td>7.61 ± 0.45</td>
</tr>
</tbody>
</table>

Data in Table 1 are expressed as means ± SEM, significant differences between groups are designed as * for $p \leq 0.05$ (MEL-group vs CONT-group). Abbreviations: CONT-controls, MEL-melatonin, GLU-glucose, GLY-glycogen, TAG-triacylglycerols, CH-cholesterol, PL-phospholipids, MDA-malondialdehyde, CTS-corticosterone, INS-insulin, n-number of animals in groups.

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![Fig. 2](image_url)  
**Fig. 2.** The course of average OGTTs in control and MEL-treated female Sprague-Dawley rats. Data in Fig. 2 are expressed as means ± SEM. Abbreviations: GLU-glucose, CONT-controls (n = 6), MEL-melatonin (n = 9).
weight gain when compared to controls were recorded in MEL treated females. Normal glucose tolerance after load prevailed in both sexes (Marková et al. 2003). Prolonged period of MEL administration (26 weeks) to rats aged 5 weeks in the present experiment resulted in modifications in carbohydrate metabolism. Prolonged MEL treatment increased serum corticosterone concentration only in females.

In the experiments by Wolden-Hanson et al. (2000) three months of MEL administration at low doses (0.4 µg/ml) in tap water to young Sprague-Dawley male rats did not alter plasma corticosterone. However, in 10-month-old (middle-aged) rats MEL restored morning concentration of the aforementioned hormone to the values of young animals. The authors associate the above change with increased glucocorticoid secretion due to MEL effect on the hypothalamic-hypophyseal-adrenal axis and decreased corticosterone clearance, respectively. No bibliographical data were available about the effect of prolonged melatonin administration on serum corticosterone in female rats. Daily MEL

<table>
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<th>Table 2</th>
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<tbody>
<tr>
<td>LIVER</td>
</tr>
<tr>
<td>Absolute (g)</td>
</tr>
<tr>
<td>7.14 ± 0.21</td>
</tr>
<tr>
<td>2.88 ± 0.132</td>
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<tr>
<td>HEART MUSCLE</td>
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<tr>
<td>Absolute (mg)</td>
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<tr>
<td>0.30 ± 0.008</td>
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<tr>
<td>SPLEEN</td>
</tr>
<tr>
<td>Absolute (mg)</td>
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<tr>
<td>0.19 ± 0.01</td>
</tr>
<tr>
<td>THYMUS</td>
</tr>
<tr>
<td>Absolute (mg)</td>
</tr>
<tr>
<td>0.07 ± 0.008</td>
</tr>
<tr>
<td>ADRENALS</td>
</tr>
<tr>
<td>Absolute (mg)</td>
</tr>
<tr>
<td>0.03 ± 0.002</td>
</tr>
<tr>
<td>PERIOVARIAL FAT</td>
</tr>
<tr>
<td>Absolute (g)</td>
</tr>
<tr>
<td>1.35 ± 0.26</td>
</tr>
<tr>
<td>EPIDIDYIMAL FAT</td>
</tr>
<tr>
<td>Absolute (g)</td>
</tr>
<tr>
<td>0.89 ± 0.13</td>
</tr>
</tbody>
</table>

Data in Table 2 are expressed as means ± SEM, significant differences between groups are designed as: * for \( p \leq 0.01 \); * for \( p \leq 0.05 \) (MEL-group vs CONT-group). Relative weight (%) = absolute weight (g or mg)/ body weight × 100. MEL-melatonin, CONT-controls, n - number of animals in groups.

![Fig. 3. The course of average OGTTs in control and MEL treated male Sprague-Dawley rats.](image)

Data in Fig. 3 are expressed as means ± SEM. Abbreviations: GLU-glucose, CONT-controls (n = 7), MEL-melatonin (n = 9).
administration for the period of 6 months did not significantly affect cortisol concentration in older women (Pawlikowski et al. 2000).

Acute decrease in serum insulin in MEL treated males was found; this fact is assumed to be one of the reasons of impaired glucose tolerance after oral load. Decreased serum insulin concentration after prolonged MEL administration to young rats of both sexes was not observed by other authors (Rasmussen et al. 1999, 2001; Mustonen et al. 2002). In middle-aged male Sprague-Dawley rats continuous MEL administration (4 µg/ml of tap water) decreased age-associated increased plasma insulin and leptin concentration to the values of young animals (Rasmussen et al. 1999, 2001; Wolden-Hanson et al. 2000).

Decreased serum insulin concentration may be attributed to inhibition of insulin secretion by MEL as reported by Peschke et al. (1997, 2000) in pancreatic islets of healthy rats in the in vitro experiments. MEL released from subcutaneous implants decreased in young diabetic male OLETF (Otsuka Long-Evans Tokushima Fatty) rats increased serum insulin, leptin and triacylglycerol concentration to the values of healthy controls but did not decrease hyperglycemia (Nishida et al. 2002).

Insulin and leptin act centrally as long-term food intake inhibitors and energy expenditure enhancers in mammals (Williams et al. 2001). Twenty-six-week-lasting MEL administration in the present study might have affected serum leptin concentration and together with decreased insulin secretion (concentration) it might decrease the energy expenditure. Increased weight of periovarial fat tissue was recorded as well as increased body weight gain in MEL treated females in the last stage of the experiment. In MEL treated males decreased body weight settled to the level of controls from week 17 to the end of the experiment (week 26).

MEL did not alter average food and water intake in males and females. In MEL-treated males and controls average food and water intake was higher when compared to MEL treated females and control. This fact corresponds with their different mean body weight. Body fat amount is considered the most important factor determining plasma leptin levels; there is a positive correlation between plasma leptin level and body mass index (Maffei et al. 1995). The reason of sexually distinct response of intraabdominal fat tissue to exogenous MEL in males and females in our experiments may have arisen from differences in leptin levels.

Chronic MEL administration to female rats prolonged the return of glycemia to the starting level in OGTTs, the connection with increased serum corticosterone level might be assumed. Decreased liver weight may be associated with decreased liver glycogen content in females. Changes in carbohydrate metabolism in MEL treated males were more outstanding than in females. Higher total intake of MEL in males resulting from higher daily intake of water could cause a decrease of serum insulin concentration (as reported by Peschke et al. 1997, 2000). It resulted in decreased glucose tolerance, manifested as

<table>
<thead>
<tr>
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<th>CONT-</th>
<th>MEL-</th>
<th>CONT-</th>
<th>MEL-</th>
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<tbody>
<tr>
<td>females</td>
<td>females</td>
<td>males</td>
<td>males</td>
<td></td>
</tr>
<tr>
<td>(n = 6)</td>
<td>(n = 9)</td>
<td>(n = 7)</td>
<td>(n = 9)</td>
<td></td>
</tr>
<tr>
<td>AUC (mmol/h)</td>
<td>15.50 ± 1.85</td>
<td>15.74 ± 0.47</td>
<td>16.33 ± 0.50</td>
<td>14.33 ± 0.45 **</td>
</tr>
</tbody>
</table>

Data in Table 3 are expressed as means ± SEM, significant differences between groupu are designated as ** for p ≤ 0.01. Abbreviations: AUC - area under the curve, CONT - controls, MEL - melatonin.
significant reduction of AUC-values in OGTTs. Intravenous glucose tolerance tests realized in various times of day did not differ in non-influenced, pinealectomized, melatonin agonist- or melatonin antagonist-treated young Wistar rats. In such acute experiments, administration of melatonin agonist did not alter the basal levels of plasma glucose and insulin nor the hepatic glucose production; pinealectomy as chronic melatonin deficiency model did not produce changes of parameters observed, too (Bizot-Espiard et al. 1998). Melatonin agonist or antagonist administration during euglycemic-hyperinsulinemic clamp in young Wistar rats did not change the fall of glycemia after insulin injection (Bizot-Espiard et al. 1998).

Decreased liver weight in MEL treated males was not accompanied with its decreased glycogen or lipid content. Lipid metabolism parameters in the serum, liver and bone marrow in rats of both sexes were not changed by chronic MEL administration except increased triacylglycerol concentration in the bone marrow in females.

Prolonged 26 weeks lasted MEL administration to young Sprague-Dawley rats of both sexes in our experiments did not have the effect described by Rasmussen et al. (1999, 2001) and Wolden-Hanson et al. (2000) after long-term MEL administration to middle-aged rats. We recorded alterations of carbohydrate metabolism parameters, associated with different changes of body weight gain, in males and females.

Acknowledgements
The project 1/0442/03 was supported by Grant Science Agency - VEGA, Ministry of Education, Slovak Republic. The experiment was conducted according to the principles provided in the Law No. 115/1995 § 24 of Slovak Republic for the Care and Use of Laboratory Animals.

We thank RNDr. Július Benický, PhD. from the Institute of Experimental Endocrinology, the Slovak Academy of Sciences, Bratislava for his help in insulin determination by RIA as well as Ing. Mária Čermáková, Mrs Renáta Ivanová and Ms Jana Sýkorová of our staff for their technical assistance.
References

ACUNA-CASTROVIEJO, D, REITER, RJ, MENENDEZ-PELAEZ, A, PABLOS, MI, BURGOS, A 1994: Characterization of high-affinity melatonin binding sites in purified cell nuclei of rat liver. J Pineal Res 16: 100-112


CSABA, G, BARATH, P 1971: Are Langerhans' islets influenced by the pineal body? Experientia 27: 962


DHAR, M, DAYAL, SS, RAMESH BABU, CS, ARORA, SR 1983: Effect of melatonin on glucose tolerance and some metabolic characteristics of high-affinity melatonin binding sites in purified cell nuclei of rat liver. J Pineal Res 16: 100-112

ESQUIFINO, A, AGRASAL, C, VELÁZQUEZ, E, VILLANUA, MA, CARDINALI, DP 1997: Effect of melatonin on serum cholesterol and phospholipid levels, and on prolactin, thyroid-stimulating hormone and thyroid hormone levels, in hyperprolactinemic rats. Life Sci 61: 1051-1058


GARCIA-MAURINO, S, GONZALES-HABA, MG, CALVO, JR, GOBERNA, R, GUERRERO, JM 1998: Involvement of nuclear binding sites for melatonin in the regulation of IL-2 and IL-6 production by human blood mononuclear cells. J Neuroimmun 92: 76-84


MARGRAF, RR, LYNCH, GR 1993: Melatonin injections affect circadian behavior and SCN neurophysiology in Djungarian hamsters. Am J Physiol 264: 615-621


