Physicochemical Analyses of Burkina Fasan Honey

A. MEDA1, C. E. LAMIEN1, J. MILLOGO2, M. ROMITO3, O. G. NACOULMA1

1 Laboratoire de Biochimie et Chimie Appliquées, U.F.R./S.V.T, Université de Ouagadougou, Burkina Faso
2 Laboratoire de Biologie et d’Ecologie Végétales, U.F.R./S.V.T, Université de Ouagadougou, Burkina Faso
3 Biotechnology Division, Onderstepoort Veterinary Institute, South Africa

Received August 16, 2004
Accepted March 3, 2005

Abstract


This study intended to determine and compare the microscopic and physicochemical characteristics of Burkina Fasan honey (n = 27) with those described in the Codex Standard, and to also find correlations between individual constituents. Physicochemical properties were determined using the harmonised methods of the international honey commission.

Microscopic pollen analyses identified the samples as being derived from one Acacia, one Lannea, three Vitellaria, two Combretaceae, two mixed Poaceae honeydew and eighteen multifloral honey. Despite the tropical ambient temperature, all the samples were nevertheless well within the limits of the Codex Standard for levels of hydroxymethylfurfural, reducing sugars, proline and diastase activity. Only 7.4% (ash), 14.8% (free acidity and pH) and 22.2% (moisture) of samples exceeded the Codex-permitted limits. A highly significant correlation was found between pH and ash content (r = 0.77; P < 0.001).

The training of non-professional beekeepers in beekeeping practice is suggested to improve the quality of Burkina Fasan honey.

Quality control, chemical composition, pollen analysis, correlation

The reason for testing honey for quality control purposes is to verify the authenticity of the product and to reveal the possible presence of artificial components or adulterants, as well as to address processing and market needs (Krell 1996). This requires not only determining the moisture and mineral content (ash), but also the levels of hydroxymethylfurfural (HMF), acidity, diastase activity, apparent sugars and water insoluble solids (Bogdanov et al. 1999).

Value limits, as defined internationally by the European Honey Directive and the Codex Alimentarius, for honey of declared origin from tropical regions like Burkina Faso, are amounts of not more than 50 milliequivalents of free acidity, 20% moisture, 0.6 g·100 g⁻¹ for general honey ash and 80 mg·kg⁻¹ for HMF. In addition, values of not less than 8 for diastase activity, 60 g·100 g⁻¹ for reducing sugars and 180 mg·kg⁻¹ for proline levels are prescribed. Some of these limits differ for honeydew honey, viz. not less than 45 g·100 g⁻¹ for reducing sugars and not more than 1 g·100 g⁻¹ for ash content (Codex Alimentarius 2001; Bogdanov and Martin 2002; Bogdanov et al. 1999; The Council of the European Union 2002).

Despite the many scientific investigations into the physicochemical and enzymatic constituents of honey, further investigations are needed in countries like Burkina Faso where such data is lacking. Since 1983, studies have been carried out in various regions of this country to characterise the bee species present, with the intention of promoting beekeeping. An inventory of melliferous plant species and traditional beekeeping practices has also been described (Guinko et al. 1987; Guinko et al. 1989a; Nombré et al. 2002). The aim of this study was to evaluate and compare the quality of some samples based on physicochemical
properties of Burkina Fasan honey with the Codex Standard, and to determine whether any compositional relationships exist between local honey.

**Materials and Methods**

**Honey sampling**

27 honey samples were collected. 17 samples (1, 2, 3, 6, 10, 11, 13, 14, 15, 16, 18, 19, 24, 25, 26, 27) were mostly obtained from non-professional beekeepers, 3 (4, 8, 9) from the Fada Beekeeping Cooperative (east of Burkina Faso), 2 (12, 17) from the Apiculture Research Centre (Centre de Production, de Formation et de Recherche en Apiculture; CPFRA) and 5 (7, 20, 21, 22, 23) were local commercial honey. Samples were collected from separate hives within one month after extraction during July 2003. Samples from the cooperative and the research centre were harvested from December 2002 to July 2003. The commercial honeys were sampled in July 2003 without certainty of the harvest period. All the samples were stored between 0 °C and 4 °C.

Qualitative microscopic analysis of honey samples and the determination of frequency classes of pollen grains were done as described (Moar 1985). Acetolysed slides were made from 10 g samples of honey (Louveaux et al. 1978) and these were compared with published photographs of different pollens (Association des Palynologues de Langue Francaise 1974; Bonnefille and Riollet 1980) and with reference slides from the Laboratory of Biology and Ecology, University of Ouagadougou.

**The physicochemical properties**

Were determined according to the Harmonised Methods of the International Honey Commission (Bogdanov 1999). The individual constituents were determined using standard procedures as described (Bogdanov 1999):

- Moisture was determined using a honey hand refractometer (HHR-2N, ATAGO, Tokyo, Japan).
- pH and free acidity were determined by titration to pH 8.3.
- Ash content was determined after the sample was burnt in an electric furnace (Thermolyne type 48000, U.S.A).
- Hydroxymethylfurural (HMF) content was based on UV absorbance at 284 nm (CECIL CE 2041 spectrophotometer 2000 series, CECIL Instruments, Cambridge, England) using the method of White (Bogdanov 1999).
- Apparent reducing sugars were determined as described (Ross 1959).
- Diastase activity was determined using the method of Schade et al. (Bogdanov 1999).
- Proline content was determined using the method of Ough et al. as adapted by Bogdanov (1999).

**Statistical analyses:**

All the determinations were carried out in triplicate and the means and standard deviations were calculated using MS Excel software. Correlation coefficients (R) for two variables were calculated using Sigmastat 2.0 Jandel Scientific software (Person Product Moment Correlation function).

**Results and Discussion**

Several quality variables for 27 local honey samples were analysed and recorded viz. pH, HMF, moisture, diastase, reducing sugars, free acidity, ash, proline levels and microscopic pollen analysis (Table 1).

Pollens analyses allowed for the identification of seven unifloral, 2 mixed Poaceae honeydew and 18 multifloral honey samples. The unifloral honeys were classed as being 2 Combretaceae honeys (64.9 % and 82.8 %), 3 Vitellaria honeys (81.4%, 90.1% and 84.8%) and 1 Acacia (59.2%) and 1 Lannea honey (94.5%). The pollen analyses showed more multifloral (67%) than unifloral (26%) and honeydew honeys (7%).

The free acidity varied from 20.3 ± 0.4 to 60.8 ± 0.4 meq·kg$^{-1}$. When considering the new limit for free acidity permitted by the Codex (2001) and the European Community Directive (The Council of the European Union 2002), only the 2 eastern Poaceae honeydew honey samples from the eastern part and 1 Vitellaria (south-western part) and 1 multifloral (central part) honey were outside the legislation limits. A high free acidity value was obtained for some honeydew honey from Morocco (Diez et al. 2004) which indicated a tendency to ferment.

Honey pH values varied from 3.5 ± 0.1 to 4.7 ± 0.1. Published reports indicate that pH should be between 3.2 and 4.5 (Bogdanov 1995). According to these values, 1 Vitellaria honey from the central part, 1 Poaceae honeydew honey and 2 multifloral honeys from the south-western part were outside this range. The mean values, however, only indicated that the central Vitellaria honey were outside this range (4.6 ± 0.1). Some honeys, such as chestnut and fir honey have been shown to have high pH values viz. 5-6 and 4.6-5.9,
respectively (Bogdanov 1995). The pH values of the 2 honeydew honeys were similar to those of honeydew honey from the Czech Republic (4.53) (Čelechovská and Vorlová 2001) and some Moroccan honeydew honey (Diez et al. 2004).

The moisture content varied from 15.0 ± 0.1 to 25.1 ± 0.0%. The *Acacia* honey, 1 Combretaceae honey and 4 multifloral honeys exceeded the permitted limit of 20% (Codex Alimentarius 2001) and can be mainly explained by the premature extraction of these honeys. This can lead to a greater risk for fermentation.

The most commonly monitored parameters for determining honey freshness include HMF levels and diastase and invertase activity (Oddo et al. 1999; Bogdanov and Martin 2002). The latter two are included as international quality standards for honey (Codex Alimentarius Commission 1969; European Economic Community 1974). According to our findings HMF levels ranged from 2.0 ± 0.2 to 41.9 ± 0.1 mg·kg⁻¹ and the diastase activity varied from 6.5 ± 0.5 to 62.3 ± 2.3. All the honey samples were well inside the current Codex Standard for HMF and confirmed the young age of the samples. The central multifloral honey showed the best mean HMF value (10.6 ± 10.5 mg·kg⁻¹) with a high mean diastase number (22.0 ± 5.9) in comparison with those from the eastern and the south-western parts (Table 1). The multifloral honey sample with 6.5 Schade units could be qualified as a honey with low natural enzyme content (Codex Alimentarius 2001).

The proline content varied from 437.82 ± 23.04 to 2169.37 ± 18.39 mg·kg⁻¹. The mean proline values of the multifloral honey were very similar (Table 1). Some authors have reported that high values for proline are typical for honeydew honeys (Diez et al. 2004). In our study, the proline content of 2 honeydew honey were 437.8 ± 23.0 and 687.6 ± 19.8 mg·kg⁻¹ but were not the highest values found. However, these values were higher than those of some groups of Moroccan honeydew honeys, which reportedly varied from 69 to 556 mg·kg⁻¹ (Diez et al. 2004). Authors such as Louveaux (1985) believe that the majority of the proline comes from bee salivary secretions. Proline content has been shown to vary considerably between different honeys (Bogdanov et al. 1999). In our study, the highest proline contents were obtained with the 3 *Vitellaria* honeys which also had the highest antioxidant activities (unpublished data). It has been shown that some amino acids have antioxidant properties (Wu et al. 2003).

The values for reducing sugars (67.30 ± 1.9 to 96.20 ± 6.4 g·100g⁻¹) were within the limits listed in the Codex Alimentarius (2001). These values, however, seemed to be higher than those for commercial honey from Australia, China, Egypt, Germany, Morocco, Pakistan, Qatar, USA, Italy and Yemen (Al-Jedah et al. 2003).

The ash content varied from 0.130 ± 0.056 to 0.947 ± 0.048 g·100g⁻¹. The two highest ash values were observed in the 2 mixed Poaceae honeydew honeys. According to the Codex Standard for honeydew or mixed honeydew ash content (Bogdanov 1995), these values confirmed the findings of the pollen analyses of the two samples. Some authors have reported that honeydew and/or mixed honeys have the highest ash content (Kirkwood et al. 1960; Bogdanov et al. 1999). High ash content has been obtained in Moroccan and Czech honeydew honeys (Diez et al. 2004). One central *Vitellaria* honey and one eastern multifloral honey had ash contents outside this range.

Kirkwood et al. (1960), using the discriminant function \(X = x_1 + x_2 + x_3\) in which \(x_1\) is the pH, \(x_2\) is the percentage of ash and \(x_3\) is the percentage of reducing sugars, found values of \(X = 86.7\) for authentic floral honey and \(X = 57.6\) for honeydew honey. Accordingly, the two mixed honeydew honeys (\(X = 82.9\) and \(X = 75.5\)) could be classed here as floral or mixed honey.

Considering the physicochemical characteristics obtained for different Burkina Fasan honey samples, 22.2% (moisture), 14.8% (free acidity and pH), and 7.4% (ash) of samples exceeded the Codex permitted limits, with a highly significant correlation between pH and
<table>
<thead>
<tr>
<th>Region of Burkina Faso</th>
<th>Honey type (Floral origin) and sample number</th>
<th>Free acidity (meq·kg⁻¹)</th>
<th>pH (3.2-4.5)</th>
<th>HMF (mg·kg⁻¹)</th>
<th>Moisture (%)</th>
<th>Diastase number (DN)</th>
<th>Reducing sugars (g·100g⁻¹)</th>
<th>Ash (mg·kg⁻¹)</th>
<th>Proline (g·100g⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Eastern</td>
<td>Combretaceae (n = 1)</td>
<td>30.8 ± 0.4</td>
<td>3.8 ± 0.1</td>
<td>21.8 ± 0.5</td>
<td>15.4 ± 0.0</td>
<td>15.4 ± 0.8</td>
<td>83.2 ± 2.4</td>
<td>0.2 ± 0.0</td>
<td>870.1 ± 23.59</td>
</tr>
<tr>
<td></td>
<td>Acacia (n = 1)</td>
<td>40.3 ± 0.4</td>
<td>3.6 ± 0.1</td>
<td>6.3 ± 0.2</td>
<td>21.9 ± 0.0</td>
<td>26.6 ± 0.8</td>
<td>83.5 ± 1.5</td>
<td>0.1 ± 0.1</td>
<td>790.4 ± 69.1</td>
</tr>
<tr>
<td></td>
<td>Poaceae-Honeydew (n = 2)</td>
<td>59.0 ± 2.5</td>
<td>4.5 ± 0.1</td>
<td>27.5 ± 5.9</td>
<td>16.2 ± 0.1</td>
<td>22.2 ± 1.9</td>
<td>73.9 ± 5.5</td>
<td>0.8 ± 0.2</td>
<td>562.7 ± 176.6</td>
</tr>
<tr>
<td></td>
<td>Multifloral (n = 5)</td>
<td>32.4 ± 2.5</td>
<td>4.0 ± 0.1</td>
<td>19.6 ± 7.5</td>
<td>16.7 ± 2.0</td>
<td>17.4 ± 2.5</td>
<td>83.8 ± 1.3</td>
<td>0.3 ± 0.2</td>
<td>897.1 ± 215.2</td>
</tr>
<tr>
<td>Central</td>
<td>Vitellaria (n = 2)</td>
<td>25.4 ± 7.2</td>
<td>4.6 ± 0.1</td>
<td>3.0 ± 01.3</td>
<td>16.3 ± 0.1</td>
<td>13.3 ± 0.6</td>
<td>70.9 ± 5.1</td>
<td>0.6 ± 0.1</td>
<td>1881.4 ± 407.2</td>
</tr>
<tr>
<td></td>
<td>Lannea (n = 1)</td>
<td>28.3 ± 0.4</td>
<td>3.5 ± 0.1</td>
<td>22.2 ± 0.3</td>
<td>15.1 ± 0.0</td>
<td>19.8 ± 1.1</td>
<td>83.3 ± 3.1</td>
<td>0.2 ± 0.1</td>
<td>890.8 ± 21.3</td>
</tr>
<tr>
<td></td>
<td>Combretaceae (n = 1)</td>
<td>41.8 ± 1.1</td>
<td>4.0 ± 0.1</td>
<td>9.8 ± 0.3</td>
<td>20.1 ± 0.0</td>
<td>62.3 ± 2.3</td>
<td>77.7 ± 1.6</td>
<td>0.5 ± 0.0</td>
<td>1090.5 ± 38.9</td>
</tr>
<tr>
<td></td>
<td>Multifloral honey (n = 5)</td>
<td>37.9 ± 8.2</td>
<td>3.9 ± 0.3</td>
<td>10.6 ± 10.6</td>
<td>20.1 ± 4.3</td>
<td>22.0 ± 5.9</td>
<td>85.5 ± 7.1</td>
<td>0.4 ± 0.1</td>
<td>910.0 ± 267.9</td>
</tr>
<tr>
<td>South-western</td>
<td>Vitellaria (n = 1)</td>
<td>52.3 ± 0.4</td>
<td>3.7 ± 0.1</td>
<td>26.7 ± 0.3</td>
<td>16.0 ± 0.1</td>
<td>21.7 ± 0.8</td>
<td>84.7 ± 2.6</td>
<td>0.5 ± 0.0</td>
<td>1968.1 ± 39.7</td>
</tr>
<tr>
<td></td>
<td>Multifloral (n = 8)</td>
<td>34.1 ± 7.0</td>
<td>4.1 ± 0.3</td>
<td>22.9 ± 14.4</td>
<td>18.4 ± 1.0</td>
<td>18.3 ± 7.1</td>
<td>84.2 ± 7.7</td>
<td>0.4 ± 0.1</td>
<td>898.2 ± 272.0</td>
</tr>
</tbody>
</table>

% of samples within the prescribed range

85.2 | 85.2 | 100 | 77.8 | 100 | 100 | 92.6 | 100

HMF = hydroxymethylfurfural; M = mean; sd = standard deviation; n = number of sample; * = Codex Standard limits for floral honeys.
ash content (R = 0.77; P < 0.001) (Fig. 1). This finding has also been described by Čelechovská and Vorlová (2001). A negative correlation was obtained between HMF and moisture (R = -0.51; P < 0.05) (not shown). There was no significant relationship between the other variables (not shown).

In conclusion, our study obtained physicochemical data for several Burkina Fasan honey derived from flowers of Combretaceae, *Acacia*, *Lannea* and *Vitellaria* species. The pollen analyses of the 2 mixed Poaceae honeydew honey were confirmed by the ash content and partly by the discriminant function of Kirkwood et al. (1960).

This study has shown that honey derived from both non-professional and professional Burkina Fasan beekeepers as well as commercial honey, are of a good quality in respect to physicochemical variables like HMF, diastase, proline and reducing sugar levels. Sample freshness was determined because of the tropical ambient temperature of the country, using HMF levels and diastase activity. A highly significant correlation was shown between pH and ash content and almost every sample was within the current standard for moisture and free acidity levels. The high levels of reducing sugars warrants further investigations to determine acceptable limits for other Burkina Fasan honey. Honey quality (based on moisture, HMF, free acidity and pH levels) could be improved by the training of non-professional Burkina Fasan beekeepers in honey harvesting and storage.

**Fyzikálně-chemická analýza medů z Burkina Faso**

Tato studie byla provedena za účelem zjištění a srovnání mikroskopických a fyzikálně-chemických vlastností medů z Burkina Faso (27) s medy popsanými v Codex Alimentarius, a také zjistit vzájemné korelace mezi jednotlivými složkami. Fyzikálně-chemické vlastnosti byly stanoveny s využitím harmonizovaných metod International Honey Commission.

Mikroskopickou analýzou pylu bylo určeno že vzorky pocházejí z medů rostlin: 1 *Acacia*, 1 *Lannea*, 3 *Vitellaria*, 2 Combretaceae, směš 2 Poaceae a 18ti medů z více rostlin. Navzdory tropickým teplotám prostředí byly všechny vzorky v limitech Codex Standard pro: hydroxymethylfurfural, redukující cukry, prolin, aktivitu amylázy. Kodexem stanovené limity byly překročeny jen hodnotami popela (7.4%), kyselosti a pH (14.8%) a vlhkosti (22.2%). Mezi pH a obsahem popela byl zjištěn výsoce signifikantní vztah (r = 0.77; P < 0.001). Autorky navrhují školení neprofesionálních včelařů v chovu včel za účelem zlepšení kvality medu z Burkina Faso.

![Fig. 1. Regression line for ash contents and pH values of various honey samples analysed.](image)
Acknowledgment

This research was supported financially by the Swedish International Foundation for Science (IFS) by means of a fellowship (E-3331-E) granted to the main author.

References


BOGDANOV, S 1999. Harmonised methods of the international honey commission. Swiss Bee Research Center, FAM, Liebefeld, Bern, Switzerland


BONNEFILLE, R, RIOLLET, G 1980: Pollens des savanes d’Afrique orientale. Editions du CNRS, France


