Exercise-induced Changes in the Clotting Times and Fibrinolytic Activity during Official 1 600 and 2 000 Meters Trot Races in Standardbred Horses

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Abstract


The aim of the study was to investigate the influence of a training program and submaximal exercise on the clotting times (prothrombin time, PT, activated partial prothrombin time, aPTT, thrombin time, TT) and fibrinogen in the horse in order to assess the probable role of physical conditioning in haemostatic processes suggested in jumping and long-distance running horses. We therefore studied 14 Standardbred horses divided into group A (6 horses) and group B (8 horses). The horses had been separately and specifically trained to take part in official 1 600 and 2 000 meters trot races, respectively. Blood samples were taken before and after the training period and after the official 1 600 and 2 000 meters trot races, under the following experimental conditions: at rest, immediately after the trial (within 2 minutes) and 30 min after the trial. Student T-test for paired data to two groups of untrained and trained horses from groups A and B showed a statistically significant decrease in fibrinogen only ($p < 0.006$ in group A and $p < 0.03$ in group B). During the official 1 600 meters trot race (group A), from among the clotting times only fibrinogen showed statistically significant differences ($p < 0.001$); during the official 2 000 meters trot race (group B), the clotting times and fibrinogen, with the exception of TT, showed statistically significant differences (from $p < 0.05$ to $< 0.001$). Our results suggest that specific training programs (as submaximal exercise) could influence the haemostatic changes occurring during an athletic competition.

Coagulation, fibrinogen, athletic horse, physical exercise

Few studies exist on the relationship between the blood clotting times and exercise. The increase in clotting and fibrinolytic activity due to exercise has been widely documented in humans, both for maximal (Dufaux et al. 1991; Lin et al. 1999; Van den Burg et al. 2000; Hilberg et al. 2003) and near-maximal efforts (Prisco et al. 1998; Hegde et al. 2001); in this species, a direct correlation between fibrinolysis and the exercise intensity has been observed (Ferguson et al. 1987). The increased fibrinolytic activity appears to counterbalance the exercise-induced increase in coagulability (Colwell 1986). As regards the equine species, data available on the effects of exercise on the clotting times are not univocal: some researchers have found evidence of fibrinogen degradation with physical activity (Marsh and Gaffney 1980), whereas others have not demonstrated a difference in coagulability or in fibrinolysis (Bayly et al. 1983ab). However, the lack of detection of exercise-induced hypercoagulability could be due to the time of blood sampling: in fact, blood was checked only before and after the exercise, when the clotting times may have returned to the baseline (McKeever et al. 1990). Recent studies have been conducted on the effect of near-maximal effort on the haemostasis of the horse, revealing different results: horses that took part in a long-distance running, did not show changes in hemostatic balance (Piccione et al. 2004a); while horses that underwent an official show jumping (Piccione
et al. 2004b) showed an increased blood clotting activity. The equivocal data reported in literature may suggest an impact of physical conditioning on the hemostatic processes, as investigated in humans (Van den Burg et al. 2000). Assuming that the training programs of jumping and long-distance running horses are different from those of the Standardbred, we wished to investigate the influence of the official 1600 and 2000 meters trot races on the clotting parameters (PT, aPTT, TT and fibrinogen) in the horse.

Materials and Methods

Fourteen Standardbred horses from the same stud farm, average age 5 ± 3 years, average body weight 430 ± 30 kg, clinically healthy, were fed three times a day: at 07.00 h on hay, at 01.00 h on concentrates, and at 19.00 h both on hay and concentrates. Horses were divided into two groups, A and B: the two groups had been separately and specifically trained to take part in the official 1 600 and 2 000 meters trot races respectively; blood samples were taken before and after the training period. The horses from group A took part in the official 1600 meters trot race (average speed 831.4 m/min. in 1’19”3 time/km), while group B took part in the official 2000 meters trot race (average speed 831.6 m/min. in 1’20”2 time/km). Blood samples were collected through external jugular venipuncture under the following experimental conditions: at rest, immediately after the trial (within 2 minutes), and 30 min after the trial. Blood samples collected in tubes containing 3.8% sodium citrate were centrifuged within 2 hours from the sampling at 2600 rpm for 10 min. The Prothrombin Time (PT), the activated Partial Thromboplastin Time (aPTT), the Thrombin Time (TT), and fibrinogen were immediately assessed from the obtained plasma by means of standard kits made especially for the SEAC Clot 2 coagulometer. The Prothrombin Time kit (code 90000200, SEAC, Florence, Italy) is based on the assay principle that the addition of an adequately calcified amount tissue factor (factor III) to citrated plasma activates factor VII, inducing the formation of a stable plug. The assay procedure consists of placing 200 µl of tissue factor (at environmental temperature) in a test tube preheated to 37 °C, followed by incubation for 5 min at 37 °C, and subsequently adding 100 µl of test plasma. Upon the addition of test plasma, a stopwatch can be started and the clotting time can be measured. The activated Partial Prothrombin Time kit (code 90000180, SEAC, Florence, Italy) is based on the addition of a piastrinic substitute, as phospholipids, a soluble activator and calcium chloride, which induces plug formation. The assay procedure consists of placing 100 µl of test plasma, and 100 µl of aPTT reagent (preheated to 37 °C), respectively, in a test tube preheated to 37 °C, followed by incubation for 3 min at 37 °C, and adding 100 µl of calcium chloride. Upon the addition of calcium chloride, the stopwatch can be started and the clotting time can be measured. The Thrombin Time kit (code 90000221, SEAC, Florence, Italy) is based on the addition of a known amount of thrombin to citrated plasma, by which fibrinogen directly transforms into fibrin. The assay procedure consists of placing 200 µl of test plasma in a test tube preheated to 37 °C, followed by incubation for 2 min at 37 °C, and then adding 200 µL of the thrombin reagent. Upon the addition of the thrombin reagent, the stopwatch can be started and the clotting time can be measured. Eventually, the standard kit for the quantitative determination of fibrinogen (code 90000211; SEAC, Florence, Italy) is based on the addition of a relatively large amount of thrombin to diluted citrated plasma, so that the clotting time depends only on the fibrinogen contained in the sample. The assay procedure consists of placing, after the dilution of the sample at 1:10 (100 µl of plasma + 900 µl of buffer), 200 µl of diluted plasma in a test tube preheated to 37 °C, followed by incubation for 2 min at 37 °C, and then adding 100 µl of the fibrinogen reagent (preheated to 37 °C). Upon the addition of the fibrinogen reagent, the stopwatch can be started and the clotting time can be measured. For this assay only the results in seconds must be converted into mg/dl by using a conversion table supplied with the kit.

Student’s t-test for paired data was applied to the clotting times of untrained horses (before undergoing the 10-week training program), and trained horses from groups A and B respectively. Since the intragroup variance was not significant, the statistical elaboration of data was carried out on the mean values of the examined clotting times. The analysis of variance (one-way and repeated measures ANOVA) was applied in order to evaluate the statistically significant differences between the different experimental conditions (at rest vs. immediately after the trial, at rest vs. 30 min after the trial, and immediately after the trial vs. 30 min after the trial), in both groups A and B. Where ANOVA showed an acceptable level of significance (p < 0.05), Bonferroni test was applied for a post hoc comparison.

Results

Figs 1-4 show the pattern of the examined clotting times, together with the relative standard deviations and statistical significances, obtained before and after the training period in both groups A and B of horses and during the different experimental conditions (at rest, immediately after the trial, and 30’ after the trial).

Statistically significant differences were observed only for fibrinogen by comparing untrained and trained horses from both groups A and B (p < 0.006 in group A, and p < 0.03
in group B); for group A, only fibrinogen showed statistically significant differences ($p < 0.001$) immediately and 30’ after the trial compared to rest; in group B, PT ($p < 0.05$) and aPTT ($p < 0.01$) showed statistically significant differences 30’ after the trial compared to rest, while fibrinogen showed statistically significant differences ($p < 0.001$) immediately and 30’ after the trial compared to rest.

**Discussion**

The comparison between untrained and trained horses based on the analysis of the obtained results revealed the same pattern of fibrinogen for both groups A and B, which showed a significant decrease (see above) thus confirming that training induces the enhancement of fibrinolytic activity. The increased fibrinolysis observed after conditioning

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**Fig. 1.** The pattern of PT, together with the relative standard deviations and statistical significances, obtained before and after the training period both in group A (official 1 600 meters trot race), and group B (2 000 meters trot official race) of horses and under different experimental conditions (official 1 600 and 2 000 meters trot races). Significance: *Vs Rest ($p < 0.05$).

**Fig. 2.** The pattern of aPTT, together with the relative standard deviations and statistical significances, obtained before and after the training period both in group A (1 600 meters trot official race), and group B (official 2 000 meters trot official race) of horses and under different experimental conditions (official 1 600 and 2 000 meters trot races). Significance: *Vs Rest ($p < 0.01$).
is in agreement with some studies conducted on humans (Stratton et al. 1991; Streiff and Bell 1994; Szymanski et al. 1994) but not with others who did not register any effect of the training on the constitutive and sub-maximal plasma levels of coagulation variables, including fibrinogen (El-Sayed et al. 1995; Van den Burg et al. 1997, 2000). The discrepant results may be due to differences in the experimental setup. In fact, the study of hemostatic variables requires a meticulous standardization, and a delay in assessing blood samples may affect the outcome of the studies (McKeever et al. 1990). In our study, clotting variables were assessed within 2 min from the trial, so as to detect transient changes in haemostasis, while in previous studies which did not reveal exercise-induced modifications, blood samples were collected when values may have returned to baseline levels (Bayly et al. 1983ab). Thus the differences in blood sampling procedures (Huisveld et al. 1992), as well as the intensity (strenuous or sub-maximal), duration and type of training could alter the results. The significant decrease observed in fibrinogen immediately after the trial and 30’ after the trial compared to rest both in the 1 600 and 2 000 meters competitions, confirms the enhancement of fibrinolytic activity during exercise as
documented in literature, along with the optimization of blood clotting processes in the equine species as well (McKeever et al. 1990; Piccione et al. 2004b). However, the level of this increase correlates with both the intensity and duration of exercise (Rosing et al. 1970; Davis et al. 1976). The importance of the intensity of physical activity in influencing hemostatic balance has been documented in humans; submaximal efforts, contrary to the long-duration efforts, actually did not involve changes in aPTT and PT in this species (Bourrey and Santoro 1988). The influence of exercise on the clotting times has been reported in horses (McKeever et al. 1990; Domina et al. 1998), in cats (Hartman 1972), and in humans (Bourrey and Santoro 1988). Changes in hemostatic balance consequent upon exercise have been widely documented, but it is still unclear how hemostatic mechanisms actually respond to this stimulus in the equine species. The statistically significant increase observed for PT and aPTT during the 2000 meters competition is in disagreement with previous researches which showed the shortening of the clotting times in trained trotting horses compared to untrained subjects (Quintavilla et al. 1994), according to studies on human athletes (Herren et al. 1992; Ibbotson et al. 1993). The shortening of PT and aPTT has also been observed in long-distance running horses (Monreal et al. 1995), proving that data provided by literature in this field are equivocal.

On the basis of the analysis of the obtained results we may say that the patterns of the clotting times and fibrinogen in response to submaximal efforts are influenced by specific training programs, which should condition the hemostatic changes occurring during an athletic competition. The knowledge of changes which affect hemostatic balance is therefore essential for monitoring the hemostatic adjustments induced by physical exercise in the athletic horse, as well as for a proper interpretation of the critical circulatory situations which commonly occur in the equine species.

References


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