Contribution to canine babesiosis in the Czech Republic

Jarmila Konvalinová1, Ivo Rudolf2, Silvie Šikutová2, Zdeněk Hubálek2, Vlasta Svobodová3, Miroslav Svoboda1

1Clinic of Dog and Cat Diseases, Faculty of Veterinary Medicine, University of Veterinary and Pharmaceutical Sciences Brno, Czech Republic
2Institute of Vertebrate Biology, v.v.i., Academy of Sciences of the Czech Republic, Medical Zoology Laboratory, Valtice, Czech Republic
3Department of Parasitology, Faculty of Veterinary Medicine, University of Veterinary and Pharmaceutical Sciences Brno, Czech Republic

Received December 14, 2011
Accepted April 11, 2012

Abstract

From March to November 2010, a total of 68 samples of blood from 41 hunting and working dogs that never left the Czech Republic were examined. Some dogs were sampled repeatedly. Blood samples were examined by polymerase chain reaction for the presence of DNA of piroplasms with negative results. Specific IgG antibodies against Babesia canis were detected by indirect immunofluorescence test, and five dogs (12.21%) were seropositive. Titres ranged from 50 to 200. One dog was positive in two samplings within 3 months. The highest number of positive samples was taken in June. The results of this study suggest a likely contact of the examined dogs with the parasite; although in 2005, a total of 340 adult unfed Dermacentor reticulatus ticks in 34 pools screened by PCR for babesiae were negative.

Canine babesiosis, one of the most important emerging tick-borne diseases of dogs with worldwide distribution, is transmitted by intra-erythrocytic protozoan of the genus Babesia. Traditionally, identification of species is based on morphology and host specificity. According to these criteria, canine piroplasms are divided into two distinct species, the large (4–5 µm) Babesia canis and the small (2.5 µm) Babesia gibsoni. Based on the differences in geographical distribution, vector specificity, antigenic properties, pathogenicity and ss-ribosomal RNA gene three subspecies of B. canis are distinguished, namely B. canis canis transmitted by Dermacentor reticulatus in Europe, B. canis vogeli transmitted by Rhipicephalus sanguineus in tropical and subtropical regions, and highly pathogenic B. canis rossi transmissible by Haemaphysalis leachi in South Africa (Uilenberg 2006). B. canis canis is the most important agent of babesiosis in Europe.

The incidence of Dermacentor reticulatus in the Czech Republic is limited to the basins of the Morava and Dyje rivers in the Břeclav and Hodonín regions and along the border with Slovakia (Kubelová and Široký 2010) (Fig. 1). The activity of adults has two peaks, with the first being in the spring from early March (however, ticks can be observed as early as late February, depending on weather conditions – adults are sometimes found even on snow) to mid April. The second peak of adults’ activity starts in September. This tick species inhabits mainly lowland biotopes, waterlogged broadleaved forests, meadows, inundated areas of rivers and fringes of forests. Incidence of Dermacentor reticulatus is irregular and insular. In Central Europe, autochthonous canine babesiosis due to B. canis was recorded in several countries. Surprisingly, no autochthonous case of canine babesiosis was reported so far in the Czech Republic, although babesiosis is present in all the countries surrounding the Czech Republic and the competent vector of the disease frequently occurs (Svobodová and Svobodová 2004). In Slovakia, first cases of autochthonous babesiosis...
started to emerge in 1997; the first case of babesiosis in dog was documented in 2000 (Chandoga et al. 2002). The incidence of babesiosis nearest to the Czech Republic was observed in the neighborhood of Malacky, Slovakia. So far, only imported babesiosis has been observed in the Czech Republic – the first imported infection was described in 1992 (Kučera 1992).

Dynamics of the spreading of canine babesiosis in Europe markedly changed in the last few years. This is largely connected with the expanding area of *D. reticulatus* distribution. In fact, the expansion of the vector’s area and the increasing number of clinical cases of babesiosis has been observed also in all adjacent countries. Babesiosis has spread to Germany, Austria, Hungary and Poland as well as Switzerland (Földvári and Farkas 2005; Sréter et al. 2005; Duh et al. 2006; Zygnier and Wedrychowicz 2006; Zygnier et al. 2008; Hornok and Farkas 2009).

Babesiosis is a serious dog disease. Typical symptoms of acute babesiosis include apathy, anorexia, fever and general weakness. The disease leads to haemolytic anaemia along with thrombocytopenia, lymphadenopathy and splenomegaly. Jaundice and haematuria can occur as well. Clinical signs are often very variable and the disease can have mild to peracute course that results in death within 2 days. Incubation period of *B. canis* is 10 to 21 days (Boozer and Macintire 2003). The infection induces an antibody reaction which is usually not strong enough to eliminate all babesiae in a host organism. Animals therefore become chronic carriers of the infection (Vercammen et al. 1997). In most cases, antibodies occur within 8 to 10 days after the infection. Puppies under 2 months of age can have colostral antibodies. Poor immune reaction is typical for puppies under 8 months of age. Antibody levels start to decrease 5 to 8 months after the animal went through the infection. Protection of dogs that underwent the disease against reinfection with the same *Babesia* species lasts 5 to 8 months on average. Antibodies acquired after the infection with one *Babesia* species do not protect against the infection with other species (Boozer and Macintire 2003; Uilenberg 2006). In certain studies, parasitaemia was detected in up to 36% of serologically negative dogs (Taboada 1998). Animals that recover from the infection and live in endemic localities acquire the so-called pre-immunity, i.e. non-sterile immunity. This means that the parasite survives in the host organism and eliminates reinfections. To the best of the authors’ knowledge, no comprehensive study on *B. canis canis* and its main tick vector *D. reticulatus* nor systematic survey of dogs from endemic localities for the presence of antibodies to *B. canis* was conducted in the Czech Republic.

The aim of our study was to examine a group of dogs living in the region where emergence of *B. canis* infection might be expected. The presence of *D. reticulatus* vector was confirmed in that locality. Moreover, it is located near to Slovakia where the disease commonly occurs. Examinations of dogs followed up the pilot study which was carried out to assess prevalence of *B. canis canis* in *D. reticulatus* ticks in the South Moravia region (Czech Republic), where the vector is widespread and enzootic focus of tularaemia occurs (Hubálek et al. 1996).

**Materials and Methods**

From March to November 2010, a total of 41 dogs of 11 breeds (Siberian husky being the most frequent breed) were examined. The sample included 21 males (one of them castrated) and 19 females, aged 1 to 12 years. The body weight of these dogs ranged from 6 to 42 kg. All animals came from the Břeclav district (Břeclav and Lanžhot localities) where *D. reticulatus* occurs. They were hunting and working dogs that never left this territory. As the dogs often worked in the field, they were more likely to be infested with ticks. All animals were clinically healthy. Blood samples of some of them were collected repeatedly. Of a total of 41 dogs, blood samples of 21 animals were collected once, 7 dogs were sampled twice and 13 animals thrice. A total of 68 blood samples were collected. Samples were taken at monthly intervals at the least. Blood was sampled in March, April, June and November.

Blood samples were taken from v. cephalica antebrachii. Samples of full blood (inserted in EDTA) and blood serum were obtained from each dog. Full blood samples were examined by PCR method. DNA was
extracted from the samples using the commercial kit QIAGEN NucleoSpin Blood (Machery-Nagel, Germany) as prescribed by the manufacturer. To amplify the diagnostic fragment of the 91 piroplasm SSU rRNA gene, we designed the forward primer TB-F (5´-CTTCAGCACCTTGAGAGAAAT-3´) and the reverse primer TB-R (5´-TCDATCCCRWACCGATGCRBAC-3´). Amplification condition were: 5 min at 94 °C, 39 cycles each of 94 °C for 45 s, 62 °C for 30 s, and 72 °C for 45 s, with the addition of a final extension period of 10 min at 72 °C. DNA isolated from the dog with confirmed imported B. canis infection (it was a patient at our clinic) was used as a positive control. Specific IgG antibodies against Babesia canis were detected by indirect immunofluorescence using the commercial Babesia canis IFA IgG Antibody Kit (Fuller Laboratories Fullerton, California, USA). The kit manufacturer states that titres 50 and more suggest recent or current infection. Host-seeking adult D. reticulatus were collected by flagging low vegetation during April 2005. All tick specimens were frozen at -60 °C until examination. Immediately before DNA isolation, ticks were surface sterilized with 70% ethanol (PCR quality), then pooled (10 ticks per pool) and mechanically disrupted using a sterile glass microblender. The total genomic DNA was extracted with QIAamp DNA Tissue Kit (Qiagen, Hilden, Germany) according to manufacturer’s instructions. Molecular detection of B. canis was performed as described previously (Jefferies et al. 2003) including primers PIRO-A1 (5´- AGGGAGCCTGAGAGACGGCTACC - 3´) and PIRO-B (5´- TTAAATACGAATGCCCCCAAC - 3´) which amplify an approximately 450 bp long conservative region of the 18S rRNA gene of babesiae.

Results

A total of 68 blood samples taken from 41 dogs were examined by PCR. No sample contained DNA of B. canis. Specific antibodies were detected in 5 dogs (12.2%). Serological examination based on indirect immunofluorescence detected 6 positive serum samples. Titres ranged from 50 to 200. One dog was positive in two samplings within 3 months. The highest number of positive samples was taken in June. The results are demonstrated in Table 1. A total of 340 adult D. reticulatus ticks (210 females and 130 males) in 34 pools were screened for babesiae. Specific PCR products of babesial DNA were not detected in any of the examined pools.

Table 1. Titres of specific antibodies against Babesia canis in positive dogs (IFA)

<table>
<thead>
<tr>
<th>Breed</th>
<th>Sex</th>
<th>Age (years)</th>
<th>Blood examination and titre of specific antibodies</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dachshund</td>
<td>Male</td>
<td>3</td>
<td>Blood examination and titre of specific antibodies</td>
</tr>
<tr>
<td>Jagdterrier</td>
<td>Female</td>
<td>11</td>
<td>March</td>
</tr>
<tr>
<td>German Shepherd</td>
<td>Male</td>
<td>5</td>
<td>Negative</td>
</tr>
<tr>
<td>Dachshund</td>
<td>Male</td>
<td>8</td>
<td>Negative</td>
</tr>
<tr>
<td>Siberian Husky</td>
<td>Male</td>
<td>8</td>
<td>NT</td>
</tr>
</tbody>
</table>

NT- not tested

Discussion

Canine piroplasms are increasingly more frequently brought to the north (Gothe and Schmid 1995; Losson et al. 1999). The geographical distribution of the causative agent and thus the occurrence of babesiosis are largely dependent on the distribution of the competent vector and susceptible host, therefore being regarded as endemic for certain regions (Martinod et al. 1986). We encounter clinical cases of babesiosis increasingly more often at our clinic. So far, they have been only cases of imported babesiosis, mostly from Slovakia. One of the risk groups is represented by search and rescue dogs that are often used to work abroad. Autochthonous infection has not been observed in the Czech Republic yet.

Examination performed in April 2005 did not detect B. canis in D. reticulatus ticks picked up in the localities where they occur. In 2010, we examined a group of dogs coming...
from the locality with incidence of *D. reticulatus* near the border with Slovakia. Those
dogs often worked in the wild, which made them more likely to get into contact with ticks.
None of the examined dogs ever left the Czech Republic. Serological examination proved
that 5 dogs were positive for antibodies against *B. canis*. In one of these dogs positive titre
was observed repeatedly after 3 months (March and June; titer 50 in both cases). In April,
one positive result (titre 100) was recorded. The highest number (4) of positive results
was observed in June when also the highest titre (200) was recorded. Spring activity of
*D. reticulatus* spans over March throughout April, having the second peak in late summer.
Antibodies start to be produced 1 to 2 weeks after contact with the infectious agent. All
the examined animals were clinically healthy. Our results indicate a likely contact of the
examined dogs with the parasite. If the infectious dose was low, the infection could induce
only antibody reaction without the outburst of the disease. In such cases, the parasite’s
DNA in the samples could be under the detection limit, or the parasite was eliminated.

Babesiae were not detected in the blood of the examined dogs by PCR. This indicates
that the parasite was either absent in the samples or there was such a low level of its DNA
that it was not possible to detect it by this type of assay. Diagnostics of babesiosis is based
on direct detection of the parasite in blood smear or on using PCR method. Serology is used
rather for seroepidemiologic studies than clinical diagnostics. Certain studies indicate that
up to 36% of dogs with parasitaemia can be serologically negative (Taboada 1998). In
localities with babesiosis, serologically positive dogs should not be used for breeding, even
if parasite was not detected in them. In these animals a low level of parasitaemia under
the detection limit of microscopy or PCR cannot be ruled out. Subclinical infections of this
kind cause problems in breeding kennels and pose a risk in cases of transfusion therapy
(Taboada 1998; Birkenheuer et al. 2003; Boozer and Macintire 2003; Irwin 2005).

As far as incidence of babesiosis is concerned, the Czech Republic has a unique position
nowadays compared to the adjacent countries where the vector’s area is expanding and
babesiosis is spreading out. Long-term incidence of *Dermacentor reticulatus* in the Czech
Republic was confirmed only in a relatively small area around the Morava and Dyje rivers
in the southeastern part of the country (Fig. 1). Although babesiosis is commonly detected
in Slovakia near the Czech national border, no autochthonous clinical case of babesiosis
has been confirmed in the Czech Republic yet. The examination of 340 ticks in 2005 did
not demonstrate the presence of the parasite’s DNA. In 2010, we detected antibodies

![Fig. 1. Localities with the incidence of Dermacentor reticulatus – Hodonín, Břeclav, Lanžhot](image-url)
against babesiosis in five dogs (12.2%). Although babesiae were not detected directly by PCR, the results of our study indicate that the presence of *B. canis* in the Czech Republic cannot be excluded. Epidemiological surveillance including distribution of competent vector, detection of the disease agent, seroprevalence study of dogs, and monitoring of acute and imported cases are needed to elucidate whether canine babesiosis could become established in the Czech Republic.

**References**


Chandoga P, Goldová M, Baranová D, Kozák M 2002: First cases of canine babesiosis in the Slovak Republic. Vet Rec 150: 82-84


Uilenberg G 2006: *Babesia* - A historical overview. Vet Parasitol 138: 3-10

