The aim of the study was to evaluate differences in haematological and biochemical indicators in young rabbits of breeds embedded in the Czech genetic resources. Seven breeds (Moravian Blue, Czech Spotted, Czech Solver, Czech White, Czech Red, Moravian White of Brown Eyes and Czech Black Guard Hairs) represented by six males of each breed were used. Weaned rabbits were fattened under identical conditions till the age of 91 days. Haematological values were measured by using automatic analyzer; biochemical indicators were determined photometrically. From the haematological indicators, mean cell volume was significantly \((P \leq 0.05)\) higher in Czech Black Guard Hairs (83.00 fl) and the lowest (73.75 fl) in Czech Red. Significantly higher \((P \leq 0.05)\) concentration of haemoglobin (143.3 g·l\(^{-1}\)) was recorded in Czech Spotted; values of 125.6 and 123.4 g·l\(^{-1}\) were found in Czech White and Czech Black Guard Hairs, respectively. From the biochemical indicators, cholesterol and triacylglycerols significantly differed in rabbit breeds. Significantly \((P \leq 0.05)\) higher level of cholesterol (3.16 mmol·l\(^{-1}\)) was determined in Moravian Blue, decreasing with body size in other breeds. Significant correlations of 0.390 were determined between live weight and serum cholesterol content. Significantly different \((P \leq 0.05)\) content of triacylglycerols was found in Czech White and Moravian White of Brown Eyes (1.30 vs. 0.75 mmol·l\(^{-1}\)). Our results showed for the first time haematological and biochemical indicators in rabbits from Czech genetic resources and the results revealed that some of these variables could be affected by the rabbit genotype.

### Abstract

Biochemical and haematological examinations provide valuable information on objective assessment of health status, in order to detect health disorders or for monitoring stress factors already at preclinical stage (Hinton et al. 1982). Changes in physiological biochemical and haematological values can also be used as indicators of welfare in rabbit breeding (Hoy and Verga 2006). Biochemical and haematological values may be influenced by a number of environmental factors, such as feed components and animal management. Differences between rabbit genotypes in biochemical and haematological values were not studied in details with the exception of status in Watanabe heritable hyperlipaemic rabbits (Kondo and Watanabe 1975), and similar model rabbits (Kurosawa et al. 1995).

The effect of rabbit genotype on blood picture and serum biochemical indicators were demonstrated in crossbreeds of Californian, Checkered giant and New Zealand White (NZW) rabbits. Crossbred rabbits had significantly higher values of haemoglobin, erythrocyte number and leukocyte number compared to NZW rabbits (Burnett et al. 2006). Similarly, Chineke et al. (2006) described rabbit genotype differences for mean values leukocyte number and haematocrit in hybrids of breeds NZW, Chinchilla and Dutch belted, however, all genotypes used in the study were similar in haemoglobin, erythrocyte number and mean cell volume (MCV). Jurčík et al. (2007) stated significant differences in haemoglobin, total protein and urea content in transgenic rabbits compared with nontransgenic rabbits.

The aim of this study was to evaluate differences in haematological and biochemical indicators in young rabbits of different breeds embedded in the Czech genetic resources.
Materials and Methods

The experiment was carried out on breeds included in the Czech Program of Conservation and Development of Genetic Resources and Agrobiodiversity. Seven breeds of Moravian Blue (MB), Czech Spotted (CS), Czech Solver (CB), Czech White (CW), Czech Red (CR), Moravian White of Brown Eyes (MW) and Czech Black Guard Hairs (CH) rabbits with 16 weaned rabbits (males and females ratio 1:1) per breed were used. The rabbits were fattened from 42 till 91 days of age under identical conditions in commercial wire cages for two rabbits, with the floor density of 0.09 m² per rabbit. The rabbits were fed pelleted feed mixture with the following nutrient content: crude protein 184 g/kg, crude fibre 169 g/kg, starch 117 g/kg and fat 36.8 g/kg. Both water and feed were available ad libitum. Environmental conditions were maintained throughout the fattening period as follows: temperature of 16–17 °C, relative humidity of 65% and 12-h light regime. The rabbits were fasted (after fattening period) overnight and slaughtered in the experimental slaughterhouse. No rabbit showed any clinical signs of disease and their body condition was good. Haematological and biochemical indicators were evaluated in rabbits at the age of 92 days. Blood was sampled from 42 rabbits (six males of the average weight of each breed) during slaughtering from the jugular vein; two samples of blood from each rabbit, one containing K₂EDTA for haematology, and one for biochemical tests collected into tubes without anticoagulant agent. The blood serum after centrifugation of samples (1000 g for 10 min) was stored at -70 °C until analysis.

Haematological examination was carried out on samples of blood stabilized within 24 h after removal (samples were stored at 4 °C) using automatic haematological analyzer Coulter model ZF (Coulter Electronics Ltd., UK). Erythrocyte and leukocyte number, haemoglobin concentration and haematocrit were determined. On the basis of erythrocyte and haematocrit values MCV was calculated. Biochemical indicators in serum as the total protein (TP), albumin, urea, glucose, cholesterol, triacylglycerols (TAG) and non-esterified fatty acids (NEFA) were determined photometrically in a spectrophotometer Libra S 22 (Biochrom Ltd., UK) by using a standard commercial kits (Randox Laboratories Ltd., Crumlin, UK).

Statistical analysis was performed using the program SAS (SAS Institute Inc., 2003). The results were processed by one-way ANOVA, all values were expressed as mean ± SE. Differences between breeds were tested by Duncan test at the level of significance $P \leq 0.05$. Relationship between serum biochemistry characteristics and live weight was evaluated by estimating Pearson correlation coefficient.

Results

Results of haematological indicators monitored in different rabbit breeds are listed in Table 1. Breed-related significant ($P \leq 0.05$) differences of haematological values were shown in MCV and haemoglobin values. The values of MCV were the highest in CH (83.00 fl) and the lowest in CR (73.75 fl). In other breeds no significant difference was found and the value of MCV ranged from 77.5 to 80.5 fl. Haemoglobin concentrations were in the range from 123.4 to 143.3 g·l⁻¹. The highest value of haemoglobin was recorded in the CS. The significant difference ($P \leq 0.05$) in haemoglobin concentration was recorded in the CS. The significant difference ($P \leq 0.05$) in haemoglobin concentration was recorded

Table 1. Values of haematological indicators in rabbit breeds of Czech genetic resources

<table>
<thead>
<tr>
<th>Variable</th>
<th>MB</th>
<th>CS</th>
<th>CB</th>
<th>CW</th>
<th>CR</th>
<th>MW</th>
<th>CH</th>
</tr>
</thead>
<tbody>
<tr>
<td>Erythrocytes (T·l⁻¹)</td>
<td>6.38</td>
<td>6.68</td>
<td>6.33</td>
<td>5.83</td>
<td>6.50</td>
<td>6.20</td>
<td>5.77</td>
</tr>
<tr>
<td>Leukocytes (G·l⁻¹)</td>
<td>2.62</td>
<td>3.60</td>
<td>2.74</td>
<td>2.06</td>
<td>2.44</td>
<td>3.68</td>
<td>2.52</td>
</tr>
<tr>
<td>Mean cell volume (fl)</td>
<td>78.6ab</td>
<td>78.75ab</td>
<td>80.5ab</td>
<td>80.25ab</td>
<td>73.75b</td>
<td>77.5ab</td>
<td>83.00a</td>
</tr>
<tr>
<td>Haemoglobin (g·l⁻¹)</td>
<td>132.6abc</td>
<td>143.3a</td>
<td>139.2ab</td>
<td>125.6abc</td>
<td>131.0abc</td>
<td>134.8abc</td>
<td>123.4a</td>
</tr>
<tr>
<td>Haematocrit (l·l⁻¹)</td>
<td>49.48</td>
<td>53.04</td>
<td>51.62</td>
<td>46.58</td>
<td>48.44</td>
<td>46.92</td>
<td>47.88</td>
</tr>
</tbody>
</table>

a,b,c means in the same row with different superscripts differ ($P \leq 0.05$)
Breed: MB - Moravian Blue, CS - Czech Spotted, CB - Czech Solver, CW - Czech White, CR - Czech Red, MW - Moravian White and CH - Czech Black Guard Hairs rabbit
only in CW, and CH. The values of erythrocytes, leukocytes and haematocrit were found in physiological values and were not significantly \((P \leq 0.05)\) affected by the genotype of rabbit. The results of biochemical examination of serum in different rabbit breeds are shown in Table 2. Concentration of cholesterol and TAG was significantly different \((P \leq 0.05)\). The highest concentration of cholesterol was determined in the large breed of MB (3.16 mmol·l\(^{-1}\)), whereas in other rabbit breeds cholesterol content did not significantly decrease with body size. Significant difference \((P < 0.05)\) in the TAG content was found in CW and MW (1.30 vs. 0.75 mmol·l\(^{-1}\)); other breeds did not show a significant difference. Other biochemical indicators such as glucose, NEFA, TP, albumin and urea did not differ between rabbit breeds. Correlation analyses indicated low values between live weight and selected biochemical indicators except for serum cholesterol concentration. This correlation was medium (0.390) and significant (0.012).

### Discussion

Haematological characteristics

In our study, significant differences were not detected in the erythrocyte number. However, Burnett et al. (2006) demonstrated a larger erythrocyte number in rabbit hybrids compared to NZW. This result was probably influenced by heterosis, rather than a breed marker. Leukocyte number was at a lower range of physiological value (2-12 G·l\(^{-1}\)) as reported by Flecknell (2000). Tůmová et al. (2007) stated a higher number of leukocytes (7.02 G·l\(^{-1}\)) in broiler rabbits in similar environmental conditions and feeding. Similar results were found by Hewitt et al. (1989), Burnett et al. (2006) or Archetti et al. (2008). Lower leukocyte number in all rabbit breeds in

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Table 2. Values of biochemical indicators in rabbit breeds of Czech genetic resources and correlation coefficients between live weight and biochemical indicators

<table>
<thead>
<tr>
<th>Variable</th>
<th>MB</th>
<th>CS</th>
<th>CB</th>
<th>CW</th>
<th>CR</th>
<th>MW</th>
<th>CH</th>
<th>r</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total protein (g·l(^{-1}))</td>
<td>70.66</td>
<td>65.40</td>
<td>64.71</td>
<td>67.17</td>
<td>62.43</td>
<td>63.99</td>
<td>60.28</td>
<td>0.259</td>
</tr>
<tr>
<td></td>
<td>± 5.61</td>
<td>± 4.31</td>
<td>± 4.29</td>
<td>± 1.62</td>
<td>± 6.86</td>
<td>± 6.29</td>
<td>± 6.56</td>
<td>(0.111)</td>
</tr>
<tr>
<td>Albumin (g·l(^{-1}))</td>
<td>37.99</td>
<td>43.30</td>
<td>38.43</td>
<td>43.78</td>
<td>42.15</td>
<td>54.38</td>
<td>40.29</td>
<td>0.177</td>
</tr>
<tr>
<td></td>
<td>± 4.31</td>
<td>± 6.32</td>
<td>± 5.78</td>
<td>± 7.11</td>
<td>± 6.67</td>
<td>± 4.03</td>
<td>± 5.99</td>
<td>(0.280)</td>
</tr>
<tr>
<td>Urea (mmol·l(^{-1}))</td>
<td>5.19</td>
<td>3.73</td>
<td>4.29</td>
<td>3.72</td>
<td>4.97</td>
<td>5.27</td>
<td>5.08</td>
<td>0.178</td>
</tr>
<tr>
<td></td>
<td>± 2.37</td>
<td>± 0.74</td>
<td>± 1.47</td>
<td>± 0.85</td>
<td>± 0.95</td>
<td>± 1.44</td>
<td>± 1.10</td>
<td>(0.218)</td>
</tr>
<tr>
<td>Glucose (mmol·l(^{-1}))</td>
<td>4.97</td>
<td>5.63</td>
<td>4.84</td>
<td>3.94</td>
<td>4.29</td>
<td>5.16</td>
<td>5.01</td>
<td>0.142</td>
</tr>
<tr>
<td></td>
<td>± 1.33</td>
<td>± 1.38</td>
<td>± 2.59</td>
<td>± 1.41</td>
<td>± 6.67</td>
<td>± 1.81</td>
<td>± 2.25</td>
<td>(0.386)</td>
</tr>
<tr>
<td>Cholesterol (mmol·l(^{-1}))</td>
<td>3.16(^a)</td>
<td>1.97(^b)</td>
<td>1.83(^b)</td>
<td>2.36(^b)</td>
<td>1.46(^b)</td>
<td>2.03(^b)</td>
<td>1.73(^b)</td>
<td>0.390</td>
</tr>
<tr>
<td></td>
<td>± 0.80</td>
<td>± 0.50</td>
<td>± 0.56</td>
<td>± 0.94</td>
<td>± 0.59</td>
<td>± 0.65</td>
<td>± 0.60</td>
<td>(0.012)</td>
</tr>
<tr>
<td>Triacylglycerols (mmol·l(^{-1}))</td>
<td>0.91(^b)</td>
<td>0.89(^b)</td>
<td>1.19(^b)</td>
<td>1.30(^b)</td>
<td>0.94(^b)</td>
<td>0.75(^b)</td>
<td>1.12(^b)</td>
<td>0.115</td>
</tr>
<tr>
<td></td>
<td>± 0.15</td>
<td>± 0.18</td>
<td>± 0.29</td>
<td>± 0.27</td>
<td>± 0.11</td>
<td>± 0.12</td>
<td>± 0.59</td>
<td>(0.477)</td>
</tr>
<tr>
<td>NEFA (g·l(^{-1}))</td>
<td>1.17</td>
<td>0.95</td>
<td>0.75</td>
<td>0.85</td>
<td>0.84</td>
<td>0.82</td>
<td>0.69</td>
<td>- 0.125</td>
</tr>
<tr>
<td></td>
<td>± 0.40</td>
<td>± 0.37</td>
<td>± 0.37</td>
<td>± 0.41</td>
<td>± 0.11</td>
<td>± 0.41</td>
<td>± 0.27</td>
<td>(0.439)</td>
</tr>
<tr>
<td>Live weight (g)</td>
<td>2603(^a)</td>
<td>2280(^b)</td>
<td>2486(^b)</td>
<td>2617(^a)</td>
<td>1840(^c)</td>
<td>2250(^b)</td>
<td>1887(^c)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>± 381</td>
<td>± 397</td>
<td>± 351</td>
<td>± 200</td>
<td>± 192</td>
<td>± 226</td>
<td>± 102</td>
<td></td>
</tr>
</tbody>
</table>

\(^a,b,c\) means in the same row with different superscripts differ \((P \leq 0.05)\).

\(r\) – correlation coefficient between live weight and a indicator, significance of correlations is in a bracket, NEFA – non-esterified fatty acids

Breed: MB - Moravian Blue, CS - Czech Spotted, CB - Czech Solver, CW - Czech White, CR - Czech Red, MW - Moravian White and CH - Czech Black Guard Hairs rabbit
this study presumably might have been connected with lower immune stage of pure
breeds, generally known for their lower resistance and viability compared to hybrid
rabbits.

Significant differences were found for MCV values in breeds CR and CH. These
breeds did not differ in body size, so MCV might be the breed characteristic. Values
of MCV determined in our study were higher than 58–79.6 fl or 52–60 fl reported
by Hewitt et al. (1989) and by Archetti et al. (2008), respectively. Our results
also confirmed findings of Burnett et al. (2006), who found significant differences
in MCV values between rabbit breeds. Content of haemoglobin found in our study
was consistent with results of Hewitt et al. (1989) and Archetti et al. (2008). On
other side, Tůmová et al. (2007) found lower haemoglobin content (76.2–73.0 g·l−1)
in young broiler rabbit. In our study, haematocrit levels were not affected by rabbit
genotype but our results differed from findings of Chineke et al. (2006) who detected
significant variation between rabbit breeds.

Biochemical indicators

We did not find significant differences in the TP contents between rabbit breeds.
Values of TP were in accordance with results of Kaneko et al. (1997), Rupić et
al. (1999), and Burnett et al. (2006). On the contrary, Archetti et al. (2008) found
lower values in growing broiler rabbits (32–61 g·l−1). The albumin levels were not
affected by rabbit genotype, our results correspond with the results of Flecknell
(2000) and Burnett et al. (2006). Similarly, the blood urea did not vary according
to rabbit genotype, the content of urea corresponded to the results of Jurčík et al.
(2007) and Archetti et al. (2008). Values of glucose were not significantly different in
rabbit breeds, average glucose values were similar to the results reported by Kaneko
et al. (1997). Cholesterol content in the examined rabbit breeds corresponded with the
range in growing rabbits presented by Burnett et al. (2006) and Javed et al. (2009).
However, Kaneko et al. (1997) and Flecknell (2000) indicated the physiological
range 0.14–1.86 and 0.1–2.0 mmol·l−1, respectively. Large breed MB had significantly
the highest cholesterol level. In CW, cholesterol did not vary from the MB and values
obtained in other breeds. Values of TAG in our study were lower than in most references
such as Rupić et al. (1999), Corino et al. (2002) and Javed et al. (2009). In our study,
significant differences in TAG concentration were found with the highest values in CW
and the lowest in MW. The effect of rabbit breed on TAG content was not described
in literature. Results of NEFA were not affected by rabbit genotype, and were higher
than those found by Corino et al. (2002) or Rommers et al. (2004). Level of NEFA
increases during fasting, higher NEFA levels have been considered as indicators of a
negative energy balance (Emery et al. 1992), which may explain the higher levels of
NEFA in our experiment after fasting rabbits before slaughtering.

Correlation coefficients between live weight and serum biochemical indicators were low
and not significant, only correlations between serum cholesterol level and live weight were
medium and significant. The relationship of cholesterol and body size could be influenced
by rabbit genotype (Lukefahr et al. 1989; Hernandez et al. 2008). Genetic changes
of cholesterol concentration in rabbits were detected by Kurossawa et al. (1985) and
Atkinson et al. (1989). Serum cholesterol levels were related to growth and carcass traits
also in cattle (Wheeler et al. 1987) and poultry (Wilcox et al. 1963).

Differences in basic haematological and biochemical indicators of rabbit breeds after
dozens of generations of pure breeding, characterized by the exterior features, have
not been systematically studied. In our study, the effect of rabbit breed was revealed in
MCV, haemoglobin, cholesterol and TAG. Significant correlations were found between
live weight and serum cholesterol levels. It is evident that rabbit breeds differ in lipid
metabolism.
Acknowledgements
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References